

Methods for the examination of biomolecular structure and dynamics.

X-ray diffraction,
mass spectrometry,
infrared spectroscopy.

Larsick Smoller

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Overview of the methods for the examination of biomolecular structure and dynamics

Structure

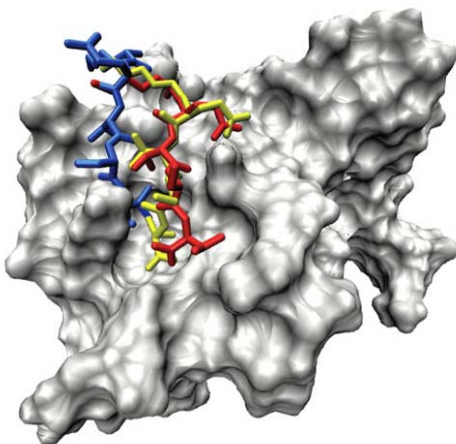
X-ray crystallography
NMR spectroscopy
Mass spectrometry
Infrared spectroscopy
Luminescence spectr.
...

Dynamics

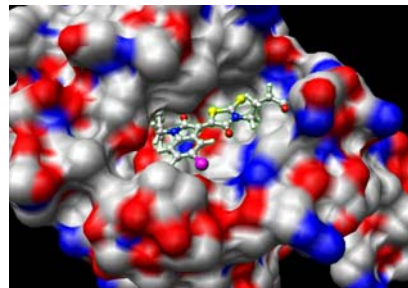
Fluorescence lifetime
Fluorescence polarization
ESR (EPR) spectroscopy
Dynamic light scattering
FCS (Fluorescence correlation spectroscopy)
...

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What is the relevance of the structural information?



Understanding the enzymatic function,
Docking of small molecules
...

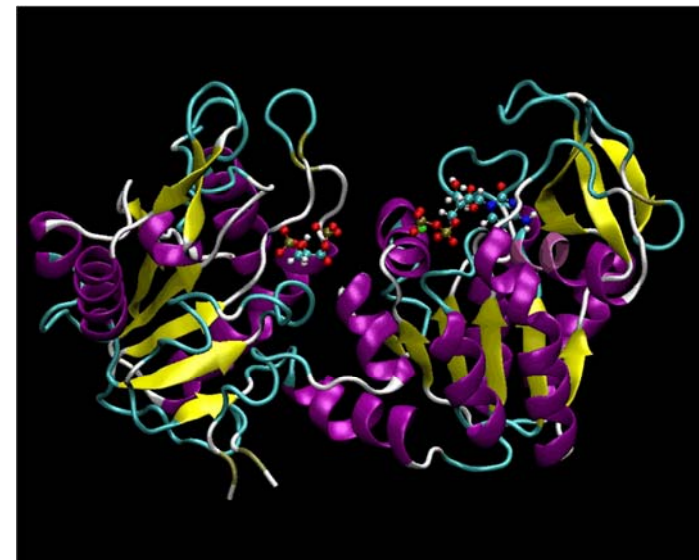


From: <http://www.cipsm.de>

<http://en.academic.ru>

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Structure - dynamics



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X-ray diffraction

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X-ray diffraction

- **What** (kind of information will it provide)?
3D structure of the molecules.
x,y,z coordinates of all (heavy) atoms.
- **Why** is it interesting/useful in medicine?
Understanding of biological processes
e.g. DNA transcription, enzymatic function,
molecular recognition, effect of mutations
Designing drug molecules (binding, docking)
- **How?**

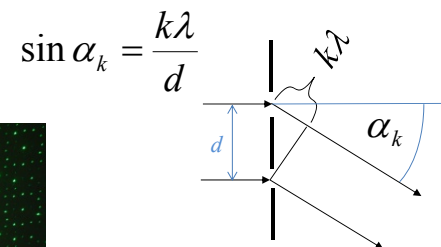
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Reminder: diffraction of light

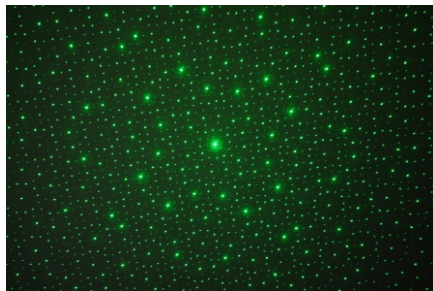
Diffraction of light on an optical grating:

Grating constant:

$$d \gtrsim \lambda$$



measure α_k if λ is known
 $\Rightarrow d$ can be calculated



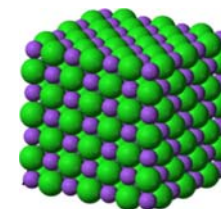
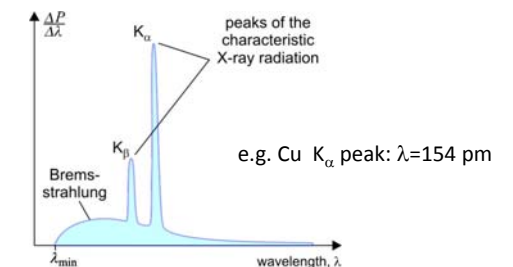
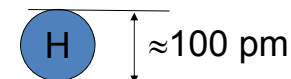
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X-ray diffraction

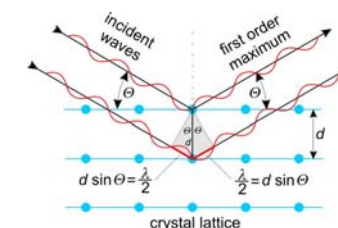
Which grating (lattice) fits to the x-ray?

$$d \gtrsim \lambda$$

$\lambda_{\text{x-ray}}$ 10-200 pm



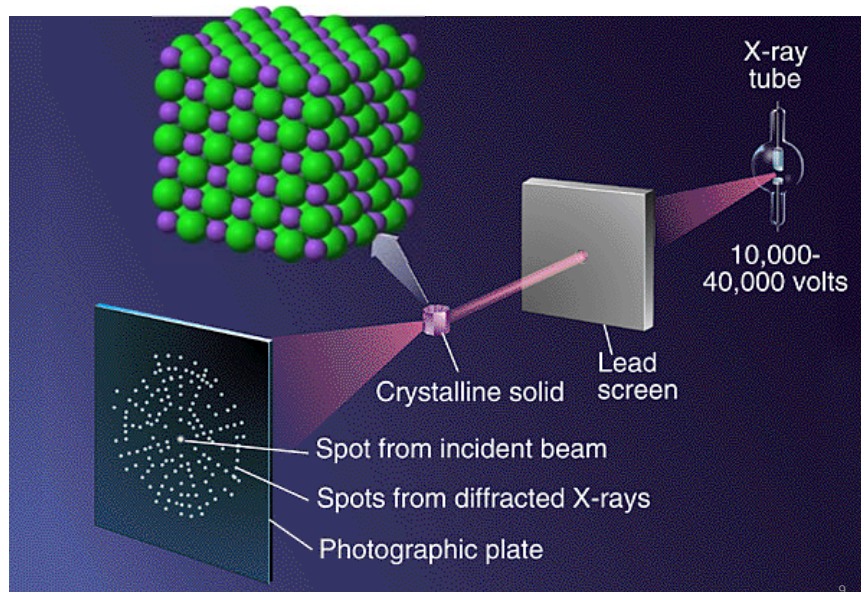
NaCl lattice constant: 564 pm



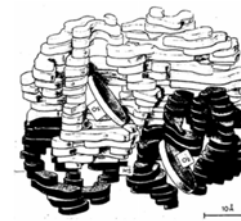
$$n\lambda = 2d \sin\theta$$

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Scematic structure of the x-ray diffractometer

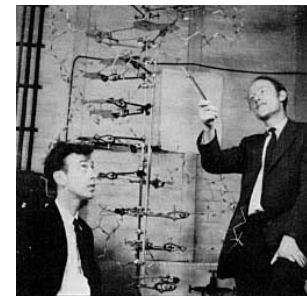


Investigation of 3D structure of macromolecules by x-ray diffraction

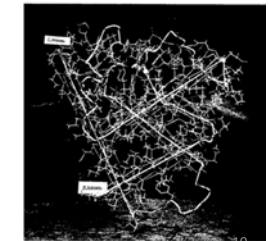
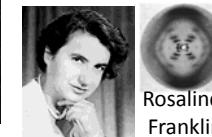


Nobel prize 1962
Globular protein
M. F. Perutz,
J. C. Kendrew

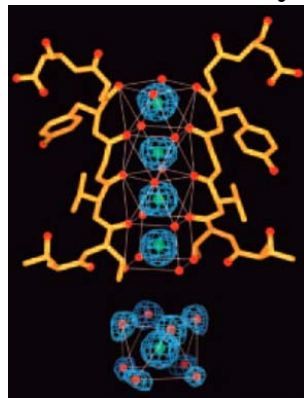
mioglobin: ~1200
atoms



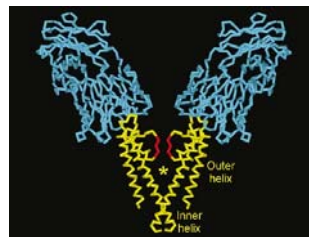
Nobel prize 1962
Structure of DNA
Francis Crick
James Watson
Maurice Wilkins



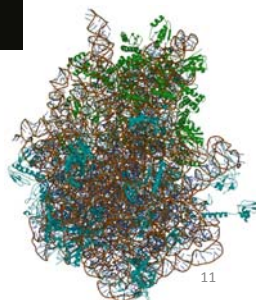
Investigation of 3D structure of macromolecules by x-ray diffraction



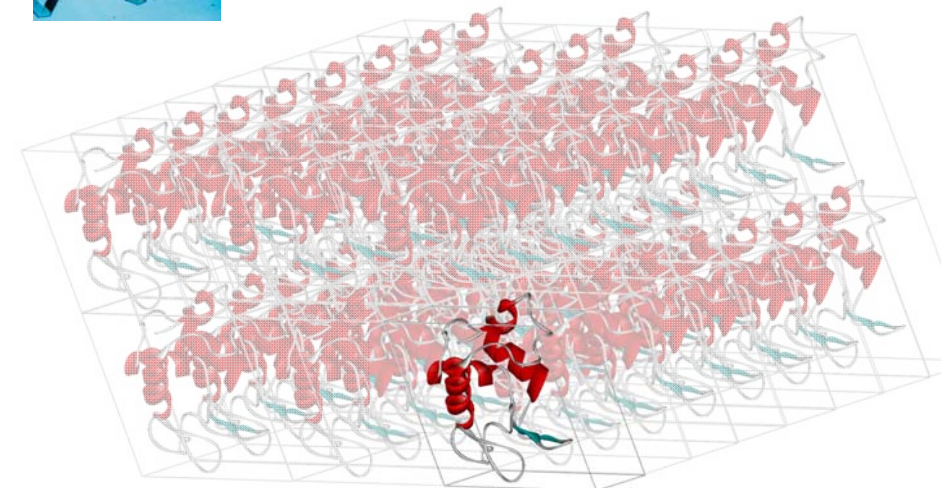
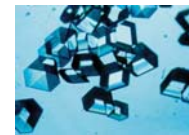
Nobel prize 2003
Membrane channel
Roderick MacKinnon

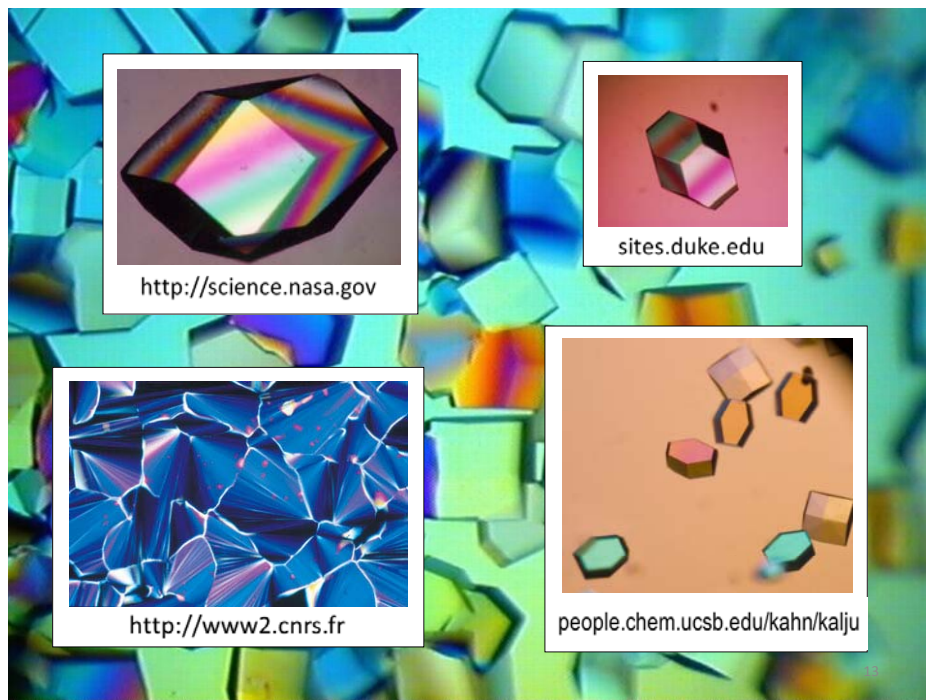


Nobel prize 2009
Ribosome
V. Ramakrishnan, T. A. Steitz,
A. E. Yonath
30S subunit: ~35000 atoms,
50S subunit: ~64000 atoms

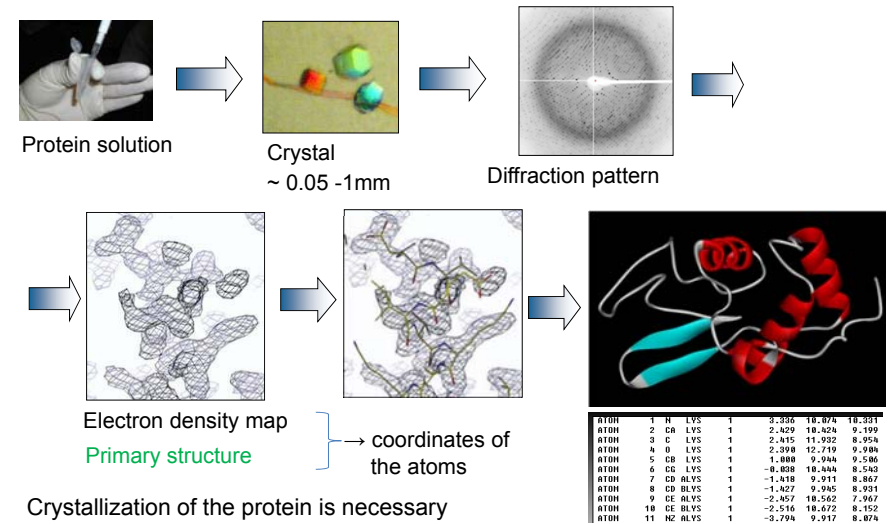


Protein crystals





Determination of 3D structure of proteins by x-ray diffraction



- Crystallization of the protein is necessary
 - Proteins with stable conformations can be investigated (globular proteins)
- The time and space average of the **electron density map** is obtained experimentally

RCSB Protein Data Bank - Mozilla Firefox

File Edit View History Bookmarks Tools Help

http://www.rcsb.org/pdb/home/home.do

RCSB Protein Data Bank

105676 x-ray cryst.
11399 NMR

3th May 2016: 118280 structures!

An Information Portal to Biological Macromolecular Structures

As of Tuesday Feb 15, 2011 at 4 PM PST there are 71264 Structures

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PDB ID lookup or Text search of the complete structure file

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Deposition

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X-ray | NMR

Validation Service

A Resource for Studying Biological Macromolecules

The PDB archive contains information about experimentally-determined structures of proteins, nucleic acids, and complex assemblies. As a member of the wwPDB, the RCSB PDB curates and annotates PDB data according to agreed upon standards.

The RCSB PDB also provides a variety of tools and resources. Users can perform simple and advanced searches based on annotations relating to sequence, structure and function. These molecules are visualized, downloaded, and analyzed by users who range from students to specialized scientists.

Hide Welcome Message

Featured Molecules

List View of Archive By: Title | Date | Category

New Features

Structure Summary Page

Binding Affinity in Tabular Format

Latest features released:

Website Release Archive

RCSB PDB News

Weekly | Quarterly | Yearly

2011-02-15

Upcoming Meeting: AAA Meeting and Family Day

RCSB PDB and SBKB will promote molecular explorations of bio the AAAS Annual Meeting (Feb 17-21, Washington DC).

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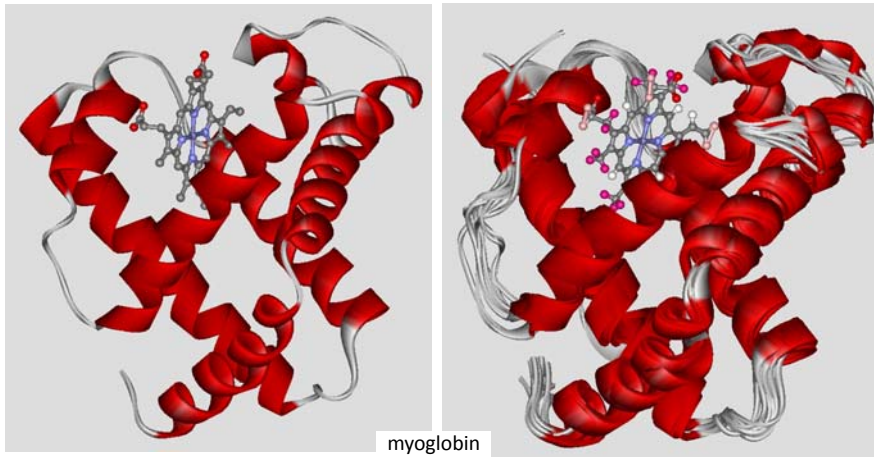
Create High Resolution

Protein data bases

- PDB** (<http://www.rcsb.org>)
Protein Data Bank
3D structures (>100 thousand)
from x-ray and
NMR spectroscopic measurements
- Swiss-prot** (<http://www.expasy.org/>)
Primary structure (sequence)
Proteomic tools to estimate
the secondary structure (homology modelling)
the chemical parameters (e.g. isoelectric point...)
or to compare different sequences

X-ray crystallography ↔ NMR

Is the crystal structure relevant for the proteins in physiological solutions?



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Mass spectrometry

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Mass spectrometry

- **What** (kind of information will it provide)?
Sorting of the molecules (or their fragments) on the basis of their mass (exactly mass/charge ratio m/z).
Identifying the fragment and the molecules
- **Why** is it interesting/useful in medicine?
Proteomics, diagnostics, screening, intraoperative pathology
- **How?**

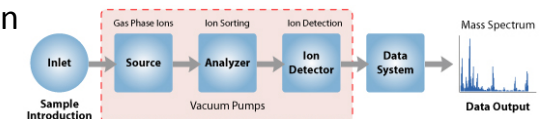
* z : the electric charge of the molecule in elementary charge units

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The principle of the mass spectrometer

Important steps:

1. Producing gaseous molecules from the sample
2. Ionization
3. Fragmentization (break the molecules into smaller fragments)
4. Analysis (sorting)
5. Detection
6. Data evaluation



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Producing gaseous ions from the sample

Mostly used methods:

ESI (electrospray ionisation)

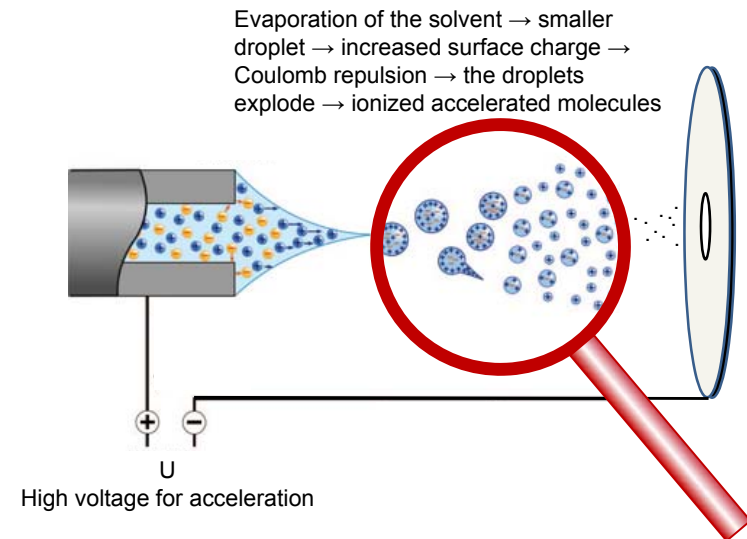
Electron ionization

Laser desorption

MALDI (matrix assisted laser desorption ionisation)

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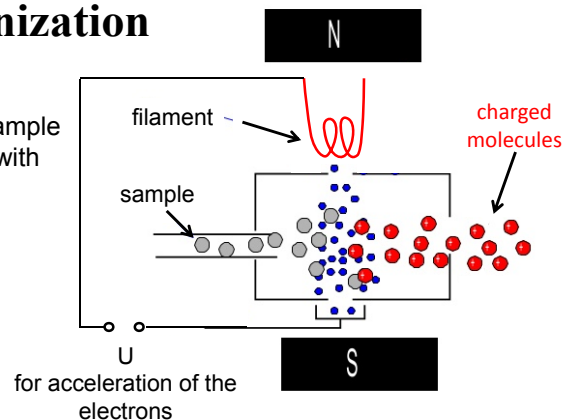
Principle of the electrospray ionization



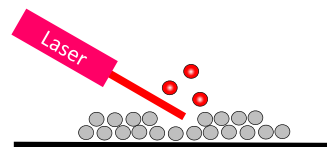
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Electron ionization

The evaporated sample molecules collide with electrons



Laser desorption

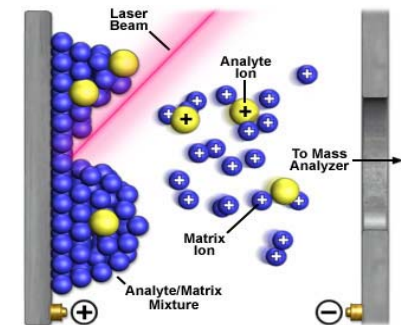


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MALDI matrix assisted laser desorption ionisation

The laser light will be absorbed by the atoms (or molecules of the matrix (not the sample!))

Ideal for big molecules.



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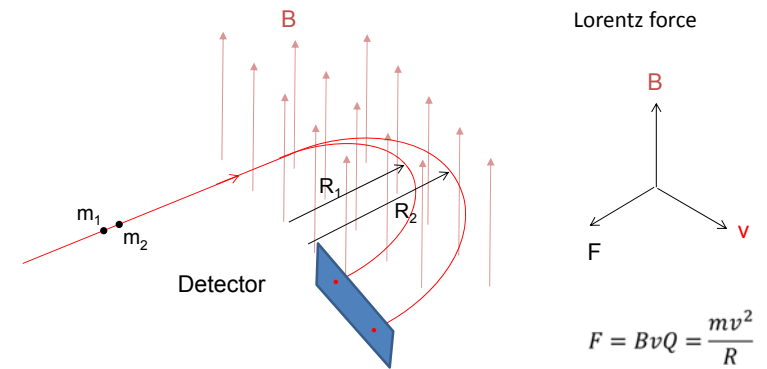
Analysis (Sorting)

Magnetic
Quadrupole
TOF
...

...

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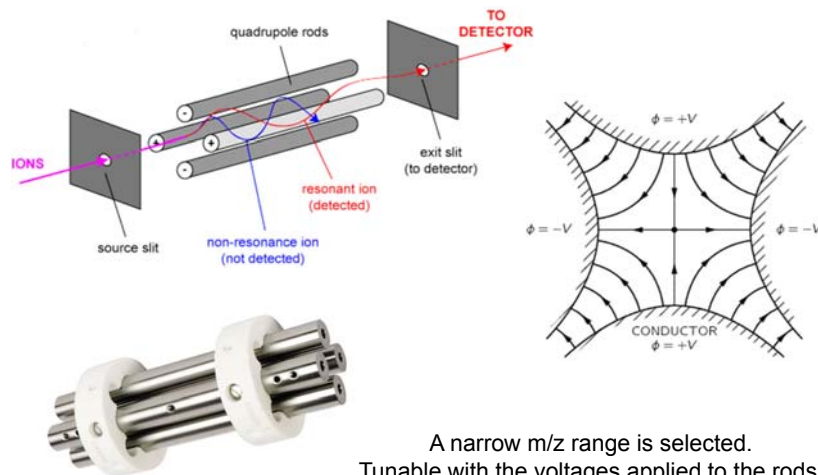
Magnetic mass analyzer



Outdated due to the high mass and high energy consumption of the magnet

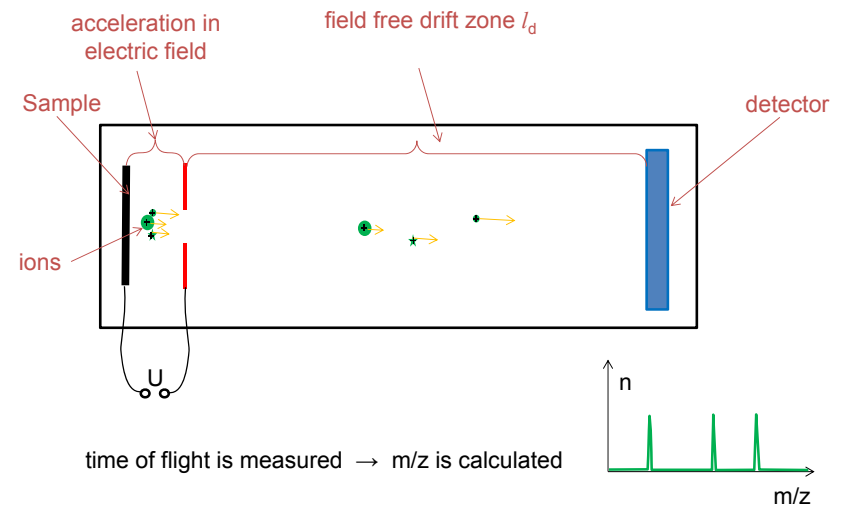
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Quadrupole mass analyzer



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Principle of the *Time of Flight* (TOF) analyzer



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Time of flight MS

The time of flight:

The velocity:

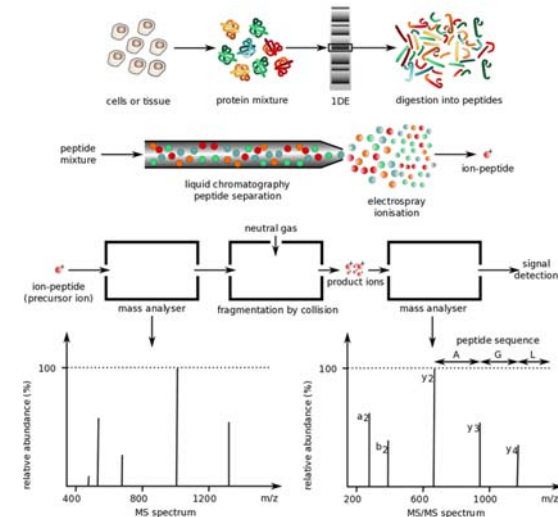
m/z ratio:



Pulsed ionisation is needed (e.g. laser desorption)

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Applications: proteomics



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Applications: diagnostics

^{13}C -urea breath test to detect *Helicobacter pylori* (causing gastric ulcer)

^{13}C -urea pill
(stable isotopic
labelling)



After 30 min the increase of the $^{13}\text{CO}_2$ in the exhaled air indicates the presence of urease enzyme (produced by *Helicobacter pylori*).

Mass spectrometry is one of the methods to determine the amount of $^{13}\text{CO}_2$ from the breath sample.



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Applications: screening

Newborn screening for
metabolic diseases



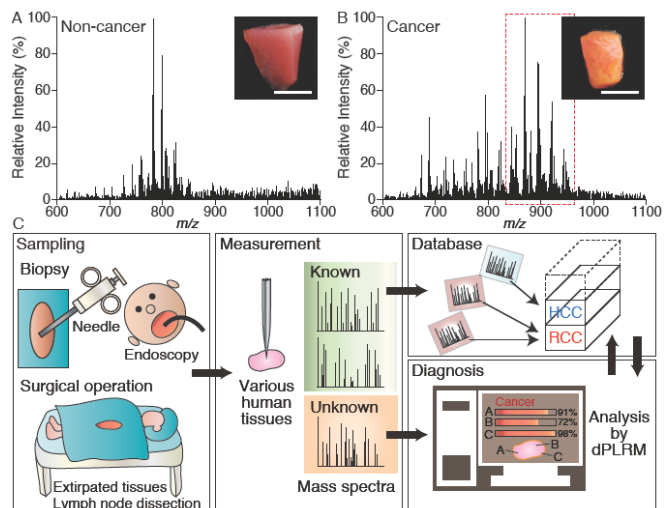
TABLE 1. Metabolic disorders detectable in newborns aged 1–5 days by using tandem mass spectrometry

Disorder	Primary metabolic indicator
Amino Acids	
Phenylketonuria	Phe
Maple syrup urine disease	Leu/Ile, Val
Homocystinuria (cystathione synthase deficiency)	Met
Hypermethioninemia	Met
Citrullinemia	Cit
Argininosuccinic aciduria	Cit
Tyrosinemia, type I	Tyr
Fatty Acids	
Medium-chain acyl-CoA dehydrogenase deficiency	C8, C10, C10:1, C6
Very-long-chain acyl-CoA dehydrogenase deficiency	C14:1, C14, C16
Short-chain acyl-CoA dehydrogenase deficiency	C4
Multiple acyl-CoA dehydrogenase deficiency	C4, C5, C8:1, C8, C12, C14, C16, C5DC
Carnitine palmitoyl transferase deficiency	C16, C18:1, C18
Carnitine/acylcarnitine translocase defect	C16, C18:1, C18
Long-chain hydroxy acyl-CoA dehydrogenase deficiency	C16OH, C18:1OH, C18OH
Trifunctional protein deficiency	C16OH, C18:1OH, C18OH
Organic Acids	
Glutaric acidemia, type I	C5DC
Propionic acidemia	C3
Methylmalonic acidemia	C3
Isovaleric acidemia	C5
3-hydroxy-3-methylglutaryl CoA lyase deficiency	C5OH
3-methylcrotonyl CoA carboxylase deficiency	C5OH

Notes: The list of primary metabolic indicators is not all-inclusive and serves only as a guideline. It is based on results obtained from laboratories experienced in tandem mass spectrometry technology and that serve as diagnostic metabolic laboratories in the United States and other countries. The identified disorders have been detected from analyses of dried blood-spot specimens collected during the newborn period. Certain disorders require complex metabolic profiles and intermetabolic relation to detect disease with low false-positive and no false-negative rate.

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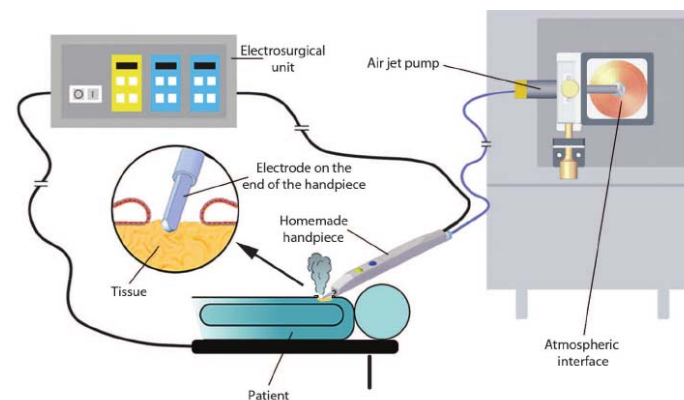
Applications: cancer diagnostics



Takeda et. al.: Innovations in Analytical Oncology - *Status quo* of Mass Spectrometry-Based Diagnostics for Malignant Tumor *Journal of Analytical Oncology*, 2012, 1, 74-80

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Applications: intraoperative MS



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IR spectroscopy

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Infrared spectroscopy

- **What** (kind of information will it provide)?
vibrational frequencies of the molecules
conformation, configuration of the molecules.
- **Why** is it interesting/useful in medicine?
Identification of molecules (drugs)
Detection of the changes in the molecular structure
Diagnostics
- **How?**

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Infrared spectroscopy

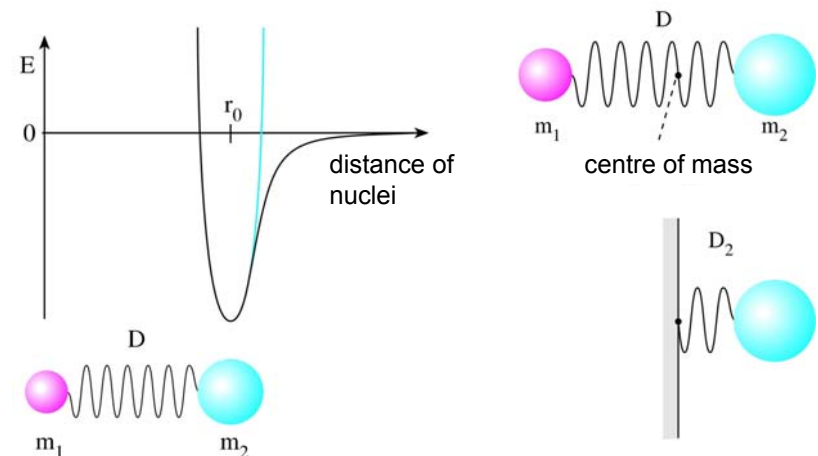
- Infrared light: $\lambda = 800 \text{ nm} - 1 \text{ mm}$
MIR (mid-infrared) : $2,5 - 50 \mu\text{m}$
- absorption spectroscopy
- the absorbed infrared radiation excite molecular vibrations
- very specific for the structure of the molecule
- special method for detection:
FT spectrometer

Molecular vibrations

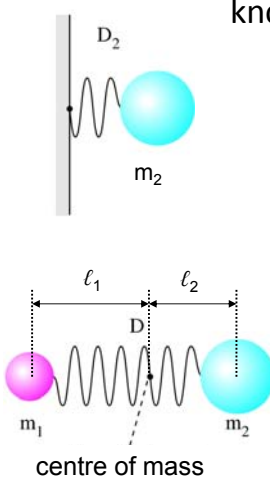
The electrons are light ($m_e \ll m_{\text{nucleus}}$), they can follow the movements of the nuclei easily, therefore the movements of the nuclei are independent of the movements of the electrons.

Classical physical description: the chemical bond is represented by a spring

Molecular vibrations:



known from elementary mechanics:



$$f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$$

$$\frac{m_2}{m_1} = \frac{\ell_1}{\ell_2}$$

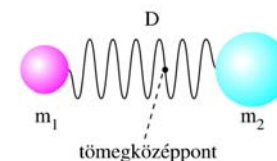
$$\frac{D_2}{D} = \frac{F/D}{F/D_2} = \frac{\Delta \ell}{\Delta \ell_2} = \frac{\ell}{\ell_2} =$$

$$= \frac{\ell_1 + \ell_2}{\ell_2} = \frac{\ell_1}{\ell_2} + 1 = \frac{m_2}{m_1} + 1 = \frac{m_1 + m_2}{m_1}$$

centre of mass

$F = D\Delta \ell$

substituting $\frac{m_1 + m_2}{m_1} = \frac{D_2}{D}$ into $f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$



frequency of the vibration:

$$f = \frac{1}{2\pi} \sqrt{\frac{D(m_1 + m_2)}{m_1 m_2}}$$

$m_{red} = \frac{m_1 m_2}{m_1 + m_2}$ is called as reduced mass

Frequency with the reduced mass: $f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$

The wavelength of an electromagnetic wave with frequency f is:

$$\lambda = \frac{c}{f} = 2\pi c \sqrt{\frac{m_{red}}{D}}$$

In the IR spectroscopy the wavenumber (ν) is used, which is the reciprocal of λ :

$$\nu = \frac{1}{\lambda} = \frac{1}{2\pi c} \sqrt{\frac{D}{m_{red}}}$$

ν : number of waves in a unit length [cm^{-1}]

Example: CO

The measured wavenumber: $\nu = 2143 \text{ cm}^{-1}$

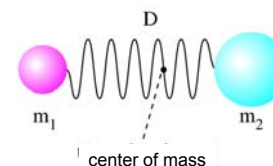
$$\left. \begin{aligned} \Rightarrow \lambda &= 4,67 \mu\text{m} \Rightarrow f = 6,43 \cdot 10^{13} \text{ Hz} \\ m_{\text{C}} &= 2 \cdot 10^{-26} \text{ kg}, m_{\text{O}} = 2,7 \cdot 10^{-26} \text{ kg} \end{aligned} \right\} \Rightarrow D = 1875 \text{ N/m}$$

if ν is known, D can be calculated

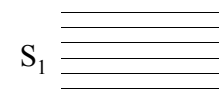
if D is known, ν can be calculated

Classical vs. quantum physics

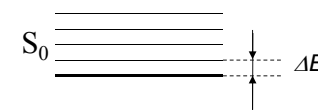
Classical physical picture



Quantum mechanical picture



$$f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$



resonance with the light with frequency f

$\Delta E = hf$

=

Vibrations of the large molecules

Molecule consisting of N atoms:

- $3N$ degree of freedom,
3-3 are the rotations and translations of the whole molecule
- $3N-6$ vibrational degree of freedom ($3N-5$ for the linear molecules)
- $3N-6$ independent normal vibrations

Normal vibrations

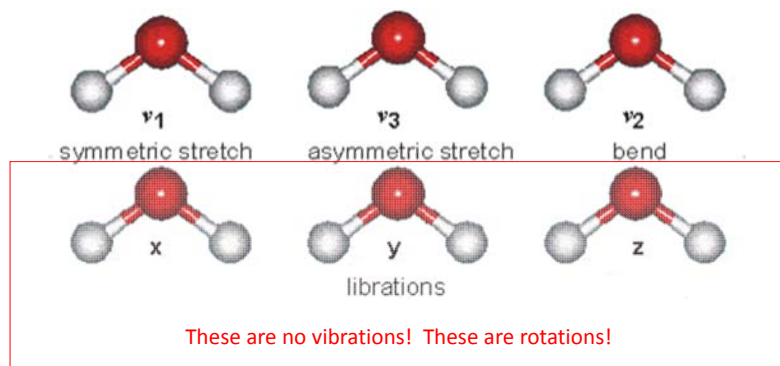
All the atoms vibrate

- with the same frequency but
- with different amplitude and
- in different direction.

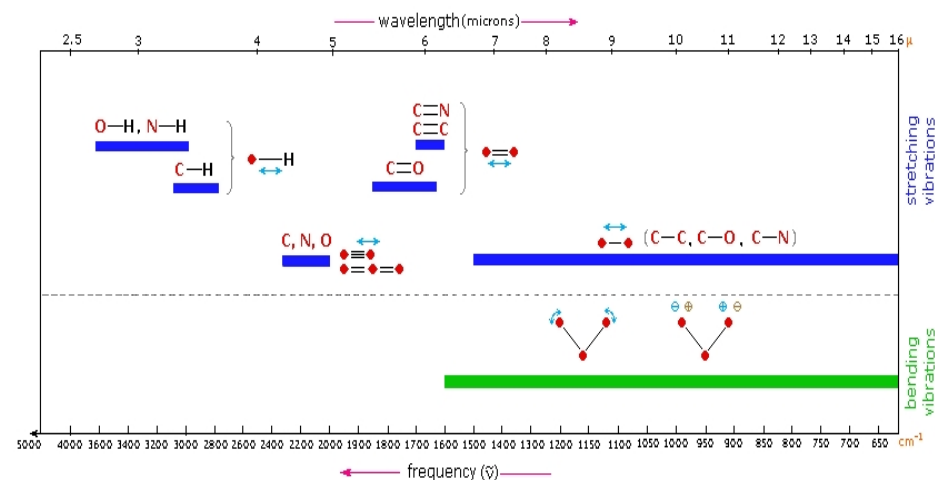
Example: water



Normal vibrations of water

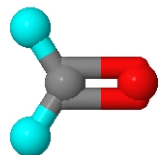
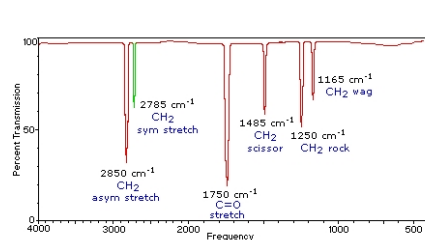


Typical vibrational frequencies (wavenumbers)



Example: Formaldehyde

Gas Phase Infrared Spectrum of Formaldehyde, $\text{H}_2\text{C}=\text{O}$

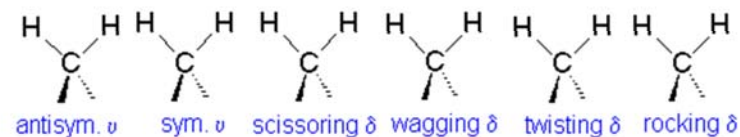
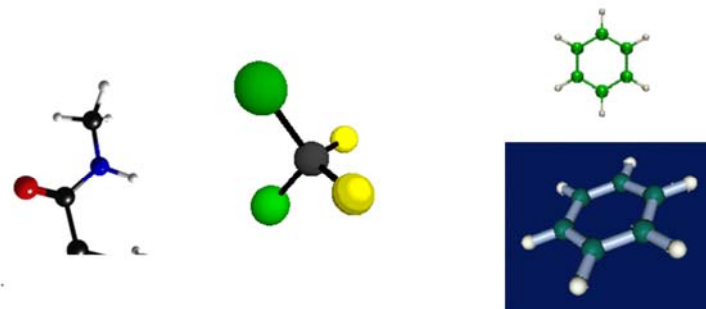


- ☐ View CH₂ Asymmetric Stretch
- ☐ View CH₂ Symmetric Stretch
- ☐ View C=O Stretch
- ☐ View CH₂ Scissoring
- ☐ View CH₂ Rocking
- ☒ View CH₂ Wagging

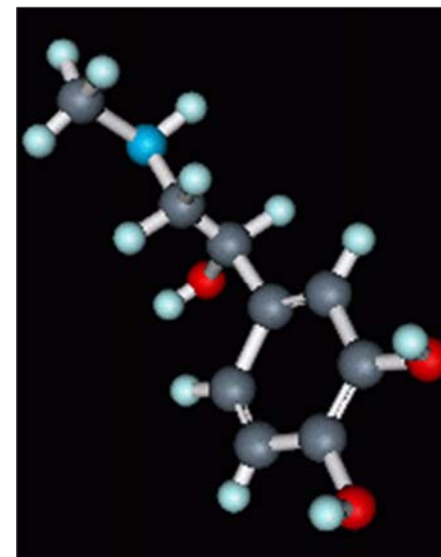
- ☐ Ball&Stick Model
- ☐ Spacefill Model
- ☐ Stick Model
- ☒ Motion Off

<http://www2.chemistry.msu.edu/faculty/reusch/VirtTxtJml/Spectrpy/InfraRed/infrared.htm>

A few vibrations for illustration



Flavin



Vibrations of the macromolecules

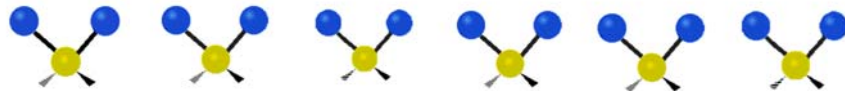
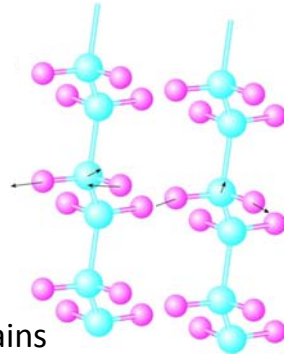
Complex global vibrations

Localised vibrations:

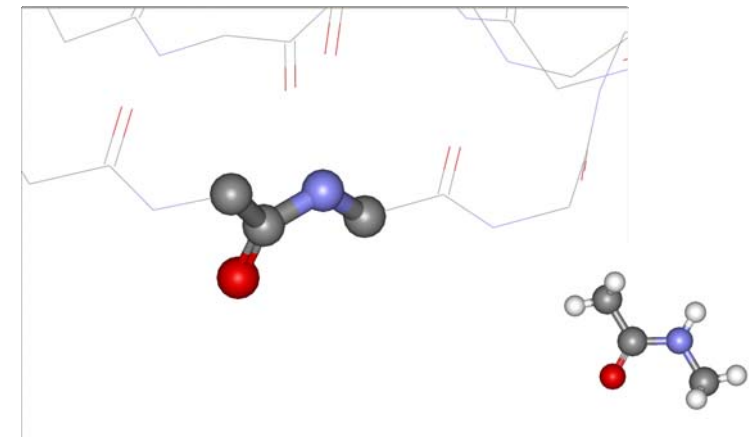
Information about the environment
of the vibrating group

e.g.:

CH₂ vibrations of the lipid fatty acid chains

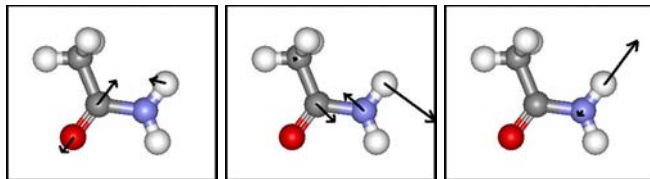


Infrared spectroscopy of the proteins



Conformation sensitive protein vibrations

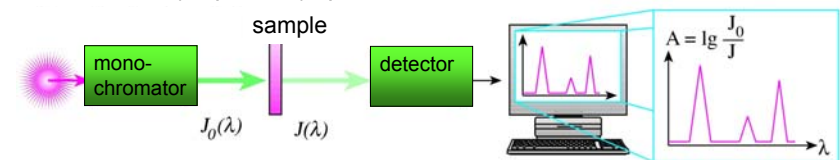
Amide vibrations



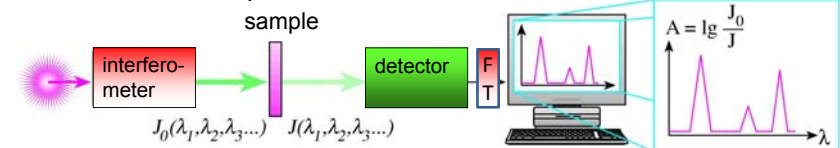
Sensitive to the
secondary and tertiary structures

The technique of the measurement : Fourier transform spectrometer (FTIR)

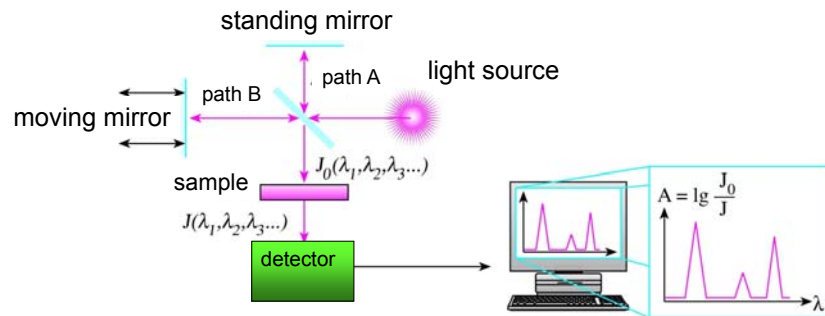
conventional (dispersion) spectrometer



Fourier transform spectrometer



The function of the interferometer

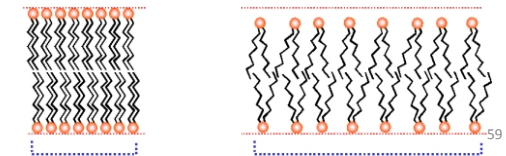
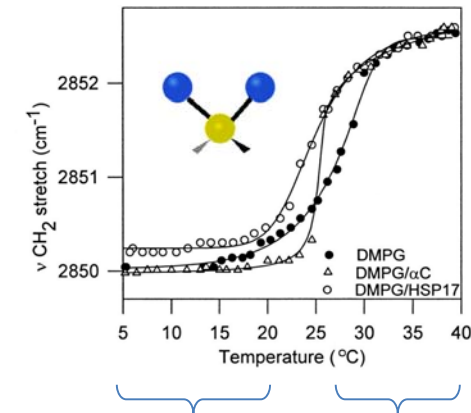


book 6.18

Applications

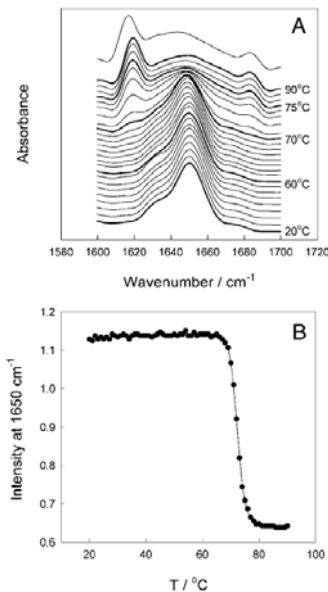
phase transitions
of a lipid bilayer

Interaction of
heat-shock proteins
with the lipid layer



Applications

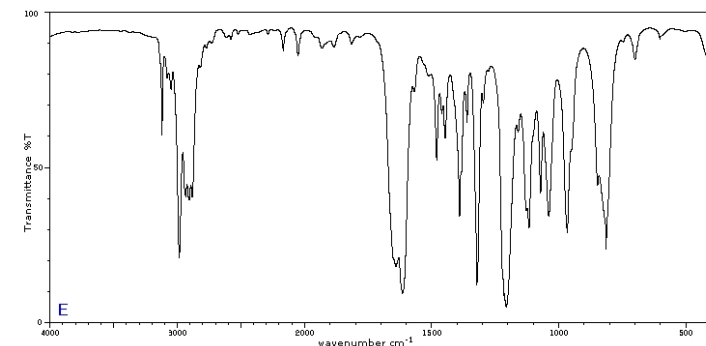
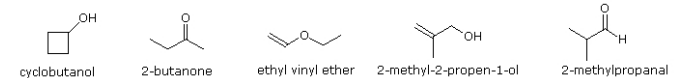
Protein denaturation



Meersman et al. Biophys J⁶⁰

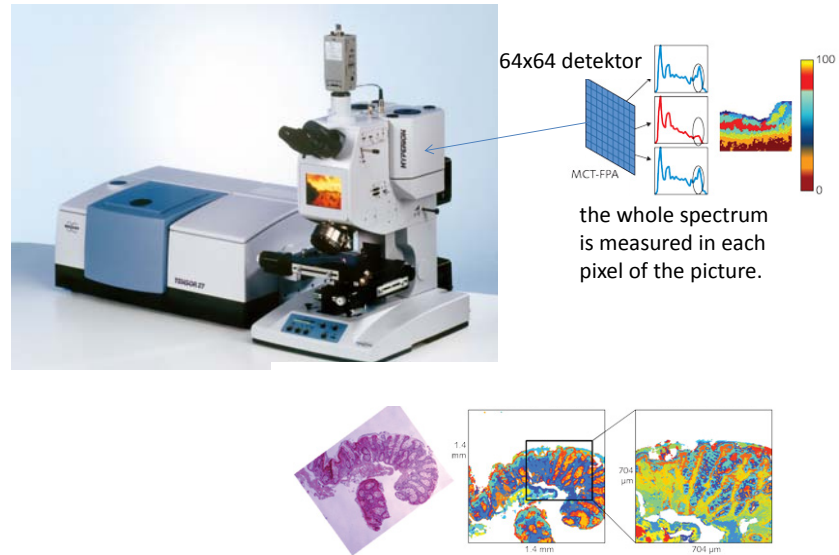
Identification of molecules

$\text{C}_4\text{H}_8\text{O}$



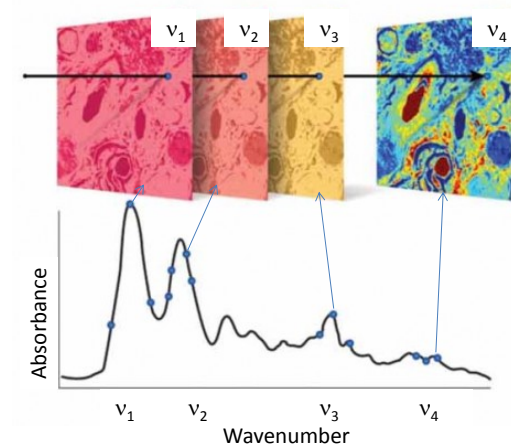
<http://www2.chemistry.msu.edu/faculty/reusch/VirtTxtJml/Spectry/InfraRed/infrared.htm>₅₁

Infrared microscopy

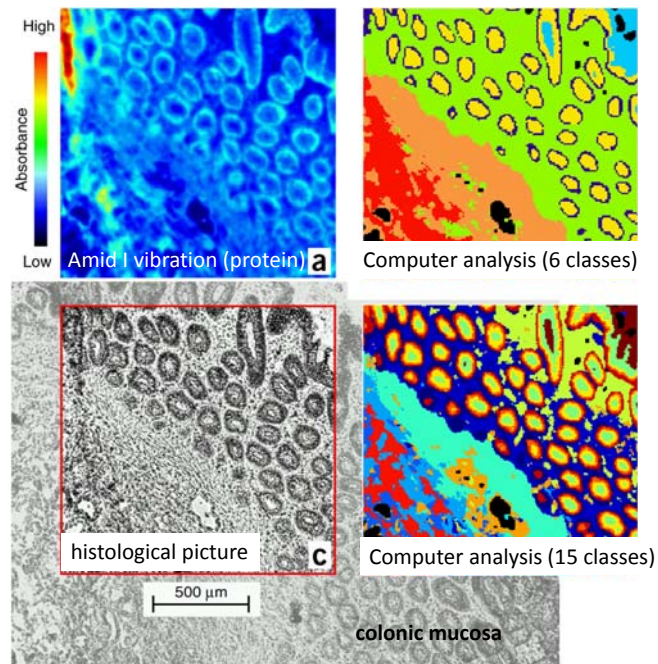


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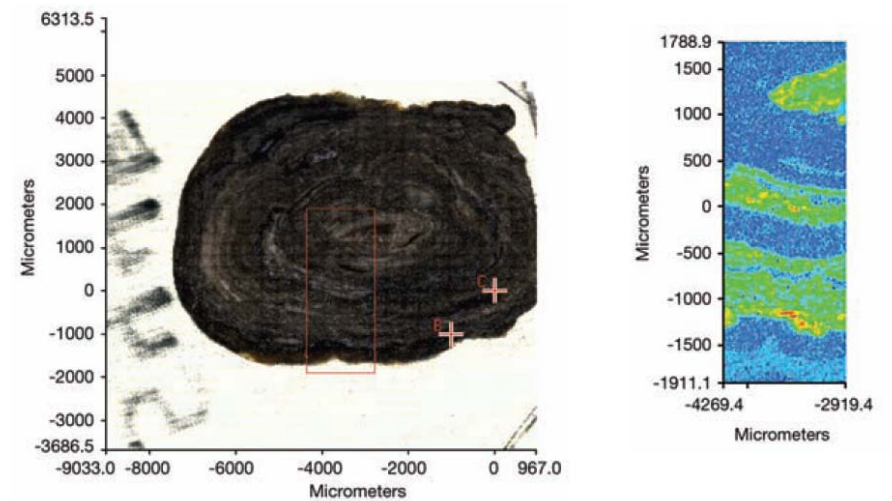
Infrared microscopy



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Classification of kidney stones using IR microspectroscopy



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