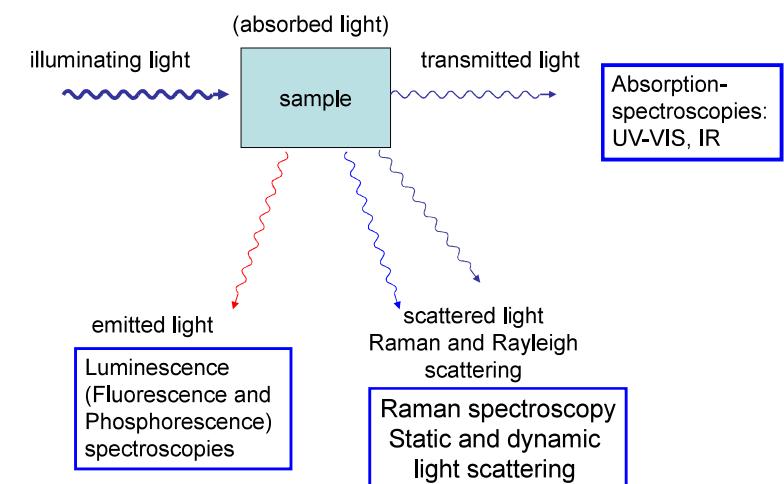


Methods for investigation of macromolecular structures: IR, CD

László Smeller

2019

What happens if a sample is illuminated by light?



Spectroscopy (Absorption and emission spectroscopy)

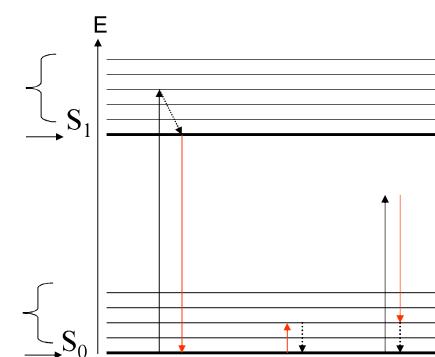
- Analysis of the wavelength dependence of the transmitted or emitted light.
- Information:
 - identification of atoms and molecules,
 - detection of changes in the molecular structure (conformation)
 - determination of the concentration

Why is light absorbed or emitted?

Jablonski diagram

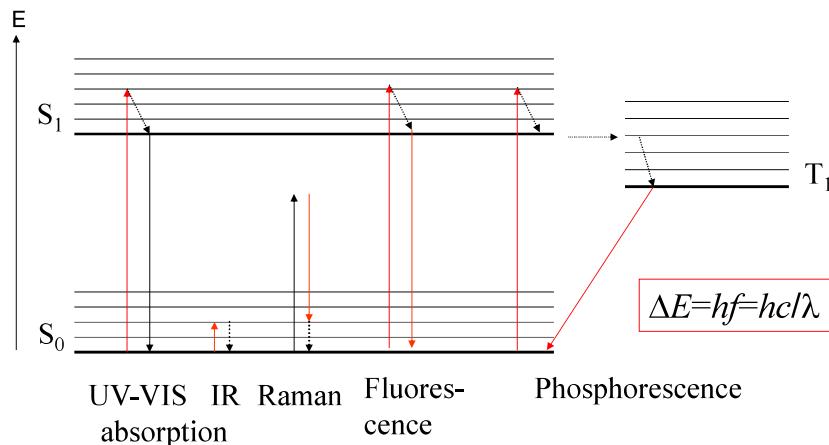
Excited electron and excited vibrational state*
Excited electron state

Vibrational excited state.*
Ground state



*only for molecules! (not for atoms)

Why is light absorbed or emitted?



Absorption spectroscopy The law of absorption

The diagram illustrates light intensity J passing through a medium of thickness x . A small differential element of width dx is shown at distance x from the entrance. The intensity changes from J to $J + dJ$ over this distance.

$$\begin{aligned} \frac{dJ}{J} &= -\mu dx \\ \int \frac{dJ}{J} &= \int -\mu dx \\ \ln J &= -\mu x + \text{const} \\ J &= J_0 e^{-\mu x} \end{aligned}$$

Absorption spectroscopy Lambert-Beer law

Law of absorption: $J = J_0 \cdot e^{-\mu x}$ where μ (material, c , λ)

Lambert-Beer law:

$$A = \lg \frac{J_0}{J} = \varepsilon(\lambda) c x$$

spectrum: $A(\lambda)$

measurement: spectrophotometer

(details: see pract. exc.)

reference solution (J_0)

information: identification (λ_{\max})

concentration (A)

UV-VIS absorption spectroscopy

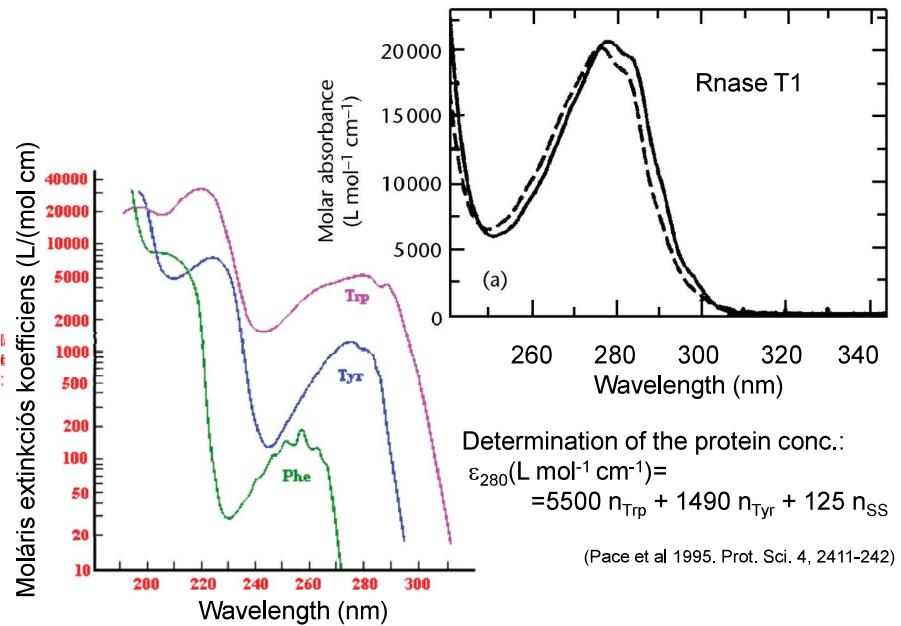
Proteins:

Absorbing site	λ_{\max} (nm)	ε (L/cm mol)
Trp	280	5600
Tyr	274	1400
Phe	257	200
Disulfide bridge	250-270	300
Peptide bond	190-230	

Determination of the protein concentration:

$$\varepsilon_{280} (\text{L mol}^{-1}\text{cm}^{-1}) = 5500 n_{\text{Trp}} + 1490 n_{\text{Tyr}} + 125 n_{\text{SS}}$$

(Pace et al 1995. Prot. Sci. 4, 2411-242)

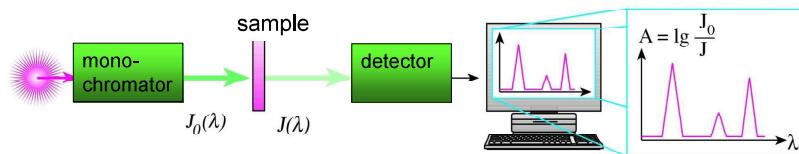


Infrared spectroscopy

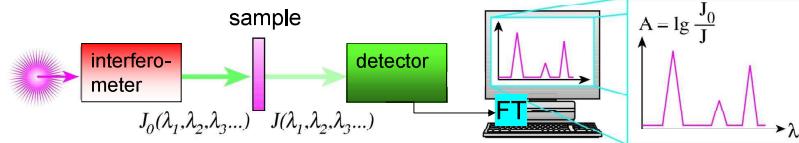
- Infrared light: $\lambda=800 \text{ nm} - 1 \text{ mm}$
MIR (mid-infrared) : $2.5-50 \mu\text{m}$
- absorption spectroscopy
- the absorbed infrared radiation excite molecular vibrations
- very specific for the structure of the molecule
- special method for detection:
FT spectrometer

The technique of the measurement : Fourier transform spectrometer (FTIR)

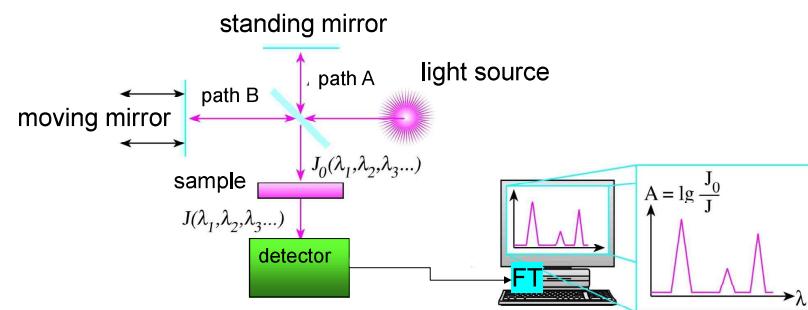
conventional (dispersion) spectrometer



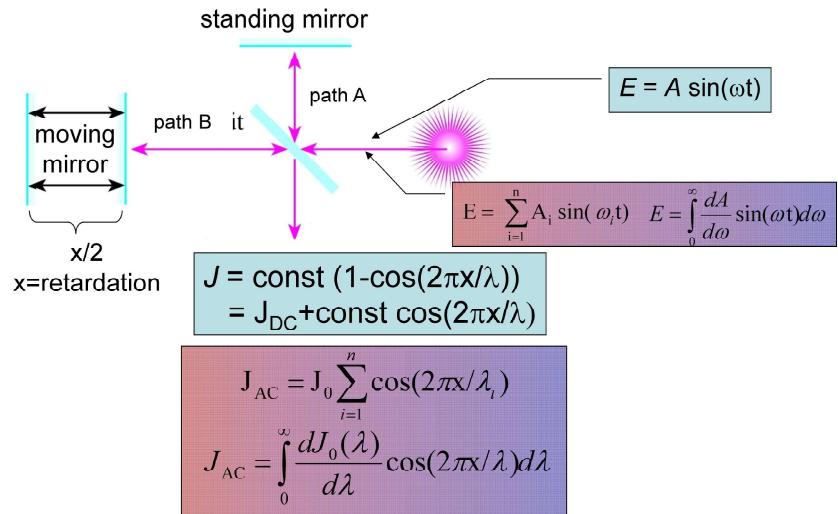
Fourier transform spectrometer



The Fourier transform spectrometer



Principle of the FTIR technique



Fourier transformation

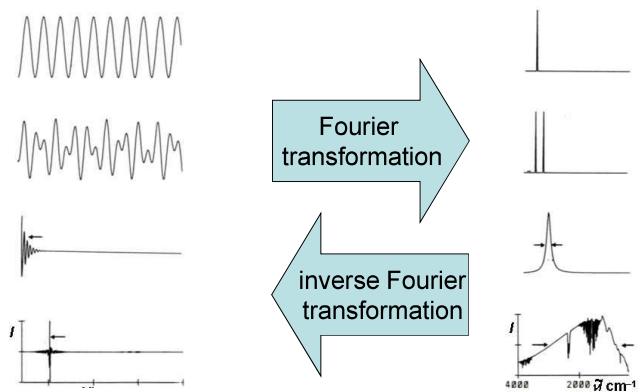
$g(x)$ is the Fourier transform of the function $f(x)$:

$$F(f(t)) = \int_{-\infty}^{\infty} f(t) e^{-2\pi i \nu t} dt = g(\nu)$$

Inverse Fourier transformation:

$$F^{-1}(g(\nu)) = \int_{-\infty}^{\infty} g(\nu) e^{2\pi i \nu t} d\nu = f(t)$$

Fourier transformation



How to obtain the spectrum in an FT spectrometer?

The intensity of the radiation passed through the interferometer:

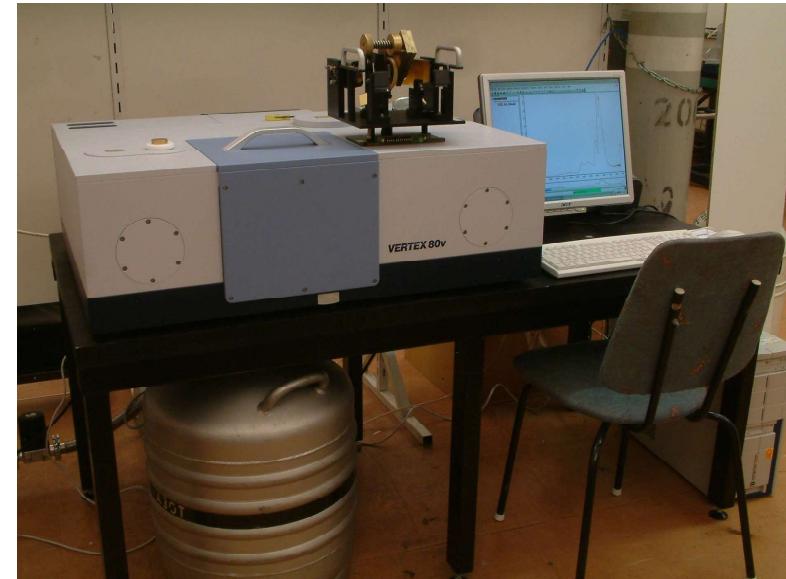
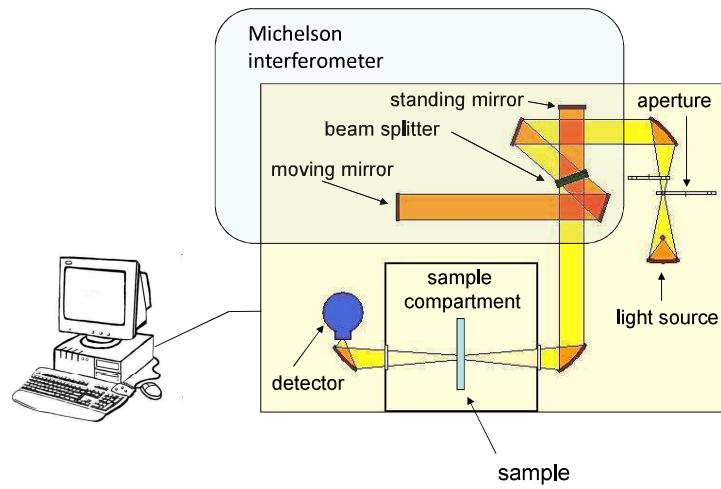
$$J_{AC} = \int_0^\infty \frac{dJ_0(\lambda)}{d\lambda} \cos(2\pi x/\lambda) d\lambda$$

is the Cosine transform of $\frac{dJ_0(\lambda)}{d\lambda}$

After the sample it reduces to $\frac{dJ(\lambda)}{d\lambda}$

$$\text{The transmission spectrum is: } T(\lambda) = \frac{J(\lambda)}{J_0(\lambda)} = \frac{\frac{dJ(\lambda)}{d\lambda}}{\frac{dJ_0(\lambda)}{d\lambda}}$$

The absorption spectrum is: $A(\lambda) = \lg(1/T(\lambda))$

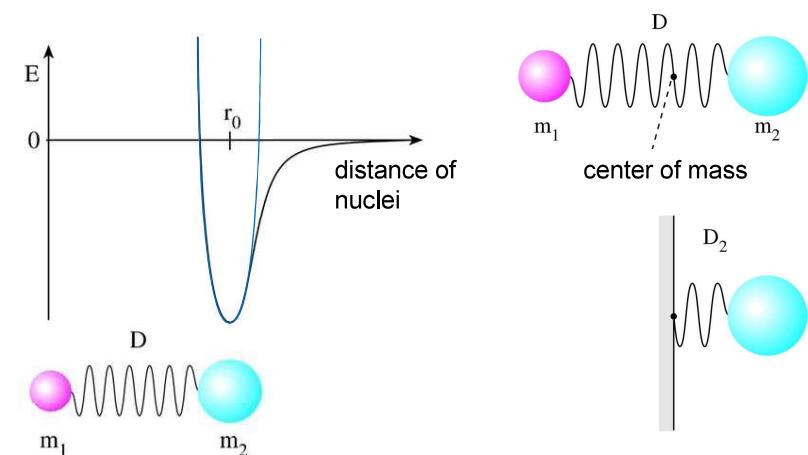


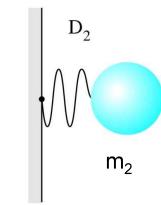
Molecular vibrations

The electrons are light ($m_e \ll m_{\text{nucleus}}$), they can follow the movements of the nuclei easily, therefore the movements of the nuclei are independent of the movements of the electrons.

Classical physical description: the chemical bond is represented by a spring.

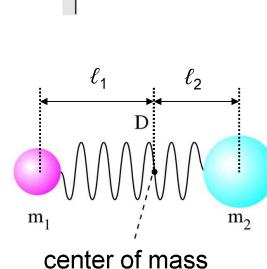
Molecular vibrations:





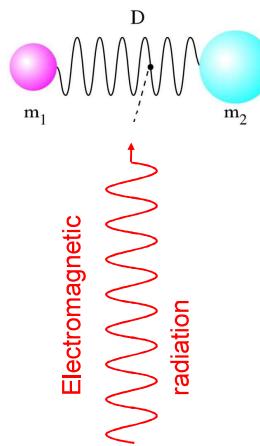
known from elementary mechanics:

$$f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$$



$$\begin{aligned} \frac{m_2}{m_1} &= \frac{\ell_1}{\ell_2} & F &= D\Delta\ell \\ \frac{D_2}{D} &= \frac{F/D}{F/D_2} = \frac{\Delta\ell}{\Delta\ell_2} = \frac{\ell}{\ell_2} = \\ &= \frac{\ell_1 + \ell_2}{\ell_2} = \frac{\ell_1}{\ell_2} + 1 = \frac{m_2}{m_1} + 1 = \frac{m_1 + m_2}{m_1} \end{aligned}$$

How can we induce molecular vibrations?



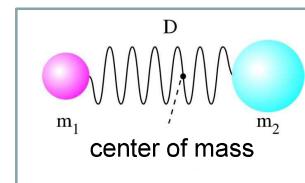
$$f_{E.M.rad} = f_{vibration} = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

Resonance



absorption of the
energy of the radiation

substituting $\frac{m_1 + m_2}{m_1} = \frac{D_2}{D}$ into $f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$



The frequency of the vibration:

$$f = \frac{1}{2\pi} \sqrt{\frac{D(m_1 + m_2)}{m_1 m_2}}$$

$m_{red} = \frac{m_1 m_2}{m_1 + m_2}$ is called as reduced mass

Frequency with the reduced mass: $f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$

The wavelength of this radiation is:

$$\lambda = \frac{c}{f} = 2\pi c \sqrt{\frac{m_{red}}{D}}$$

In the IR spectroscopy the wavenumber (ν) is used, which is the reciprocal of λ :

$$\nu = \frac{1}{\lambda} = \frac{1}{2\pi c} \sqrt{\frac{D}{m_{red}}}$$

ν : number of waves in a unit length [cm^{-1}]

Example: CO

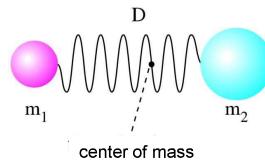
The measured wavenumber: $\nu = 2143 \text{ cm}^{-1}$

$$\left. \begin{aligned} \Rightarrow \lambda &= 4,67 \mu\text{m} \Rightarrow f = 6,43 \cdot 10^{13} \text{ Hz} \\ m_C &= 2 \cdot 10^{-26} \text{ kg}, m_O = 2,7 \cdot 10^{-26} \text{ kg} \end{aligned} \right\} \Rightarrow D = 1875 \text{ N/m}$$

if ν is known, D can be calculated
if D is known, ν can be calculated

Classical vs. quantum physics

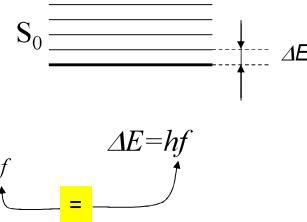
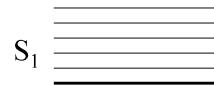
Classical physical picture



$$f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

resonance with the light with frequency f

Quantum mechanical picture

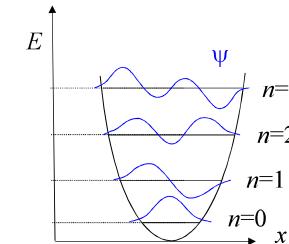


$$\Delta E = hf$$

Quantummechanical description

Quantum harmonic oscillator

Particle in a harmonic potential



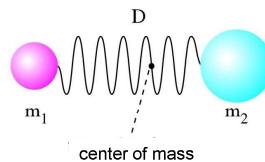
Energy eigenvalues

$$E_n = hf(n + \frac{1}{2})$$

$n = 0, 1, 2, \dots$

Classical vs. quantum physics

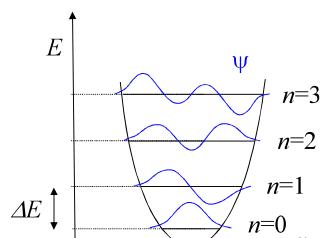
Classical physical picture



$$f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

resonance with the light with frequency f

Quantum mechanical picture



$$\Delta E = hf$$

Dependence of the vibrational frequency on the mass of the atoms and on the strength of the chemical bond.

mass: ↓				
Infravörös rezgési frekvenciák (cm⁻¹)				
B-H 2400	C-H 3000	N-H 3400	O-H 3600	F-H 4000
Al-H 1750	Si-H 2150	P-H 2350	S-H 2570	Cl-H 2890
	Ge-H 2070	As-H 2150	Se-H 2300	Br-H 2650

Water (O-H): 3600 => D₂O: 2600 cm⁻¹

Bond strength:

C-N: 1100 cm⁻¹,
C=N: 1660 cm⁻¹,
C≡N: 2220 cm⁻¹.

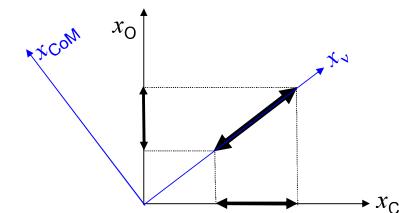
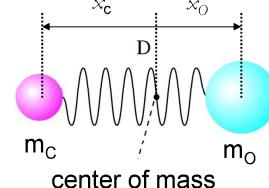
Vibrations of the large molecules

Molecule consisting of N atoms:

- $3N$ degree of freedom,
3-3 are the rotations and translations
of the whole molecule
- $3N-6$ vibrational degree of freedom
($3N-5$ for the linear molecules)
- $3N-6$ independent normal vibrations

Normal coordinates

Shown in case of a simple molecule of two atoms:



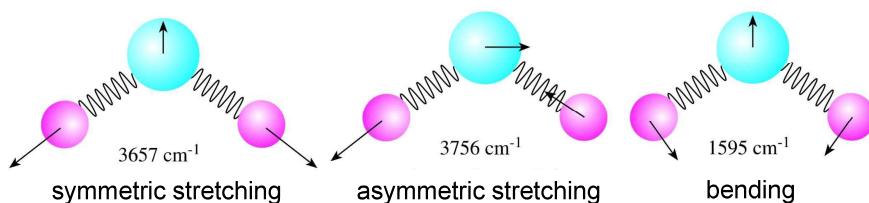
In general case one has to rotate a $3N$ dimensional coordinate system.
Linear transformation (matrix operation)

Normal vibrations

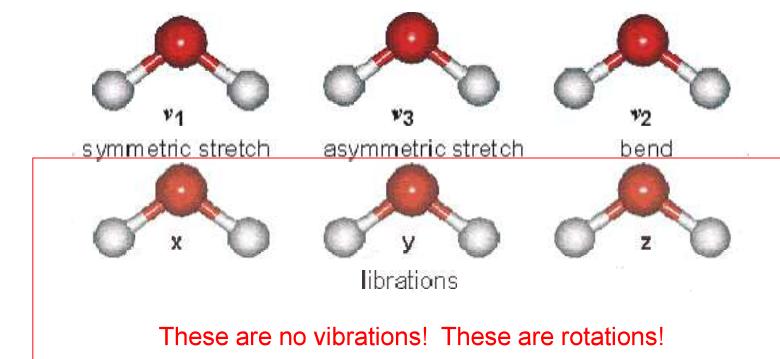
All the atoms vibrate

- with the same frequency but
- with different amplitude and
- in different direction.

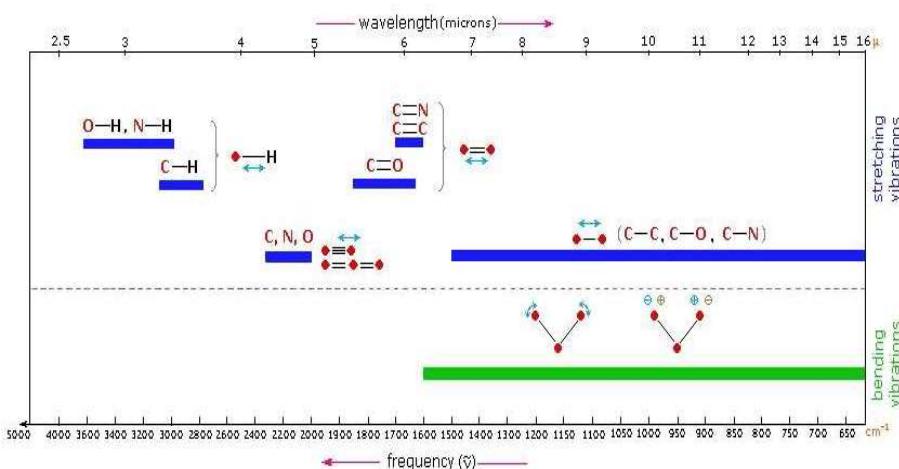
Example: water



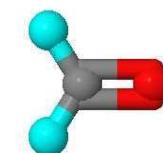
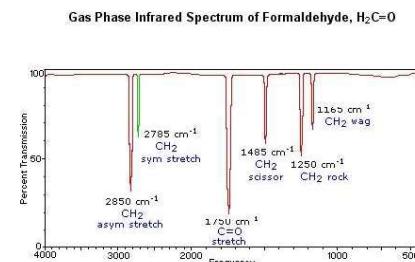
Normal vibrations of water



Typical vibrational frequencies (wavenumbers)



Example: Formaldehyde



- View CH₂ Asymmetric Stretch
- View CH₂ Symmetric Stretch
- View C=O Stretch
- View CH₂ Scissoring
- View CH₂ Rocking
- View CH₂ Wagging

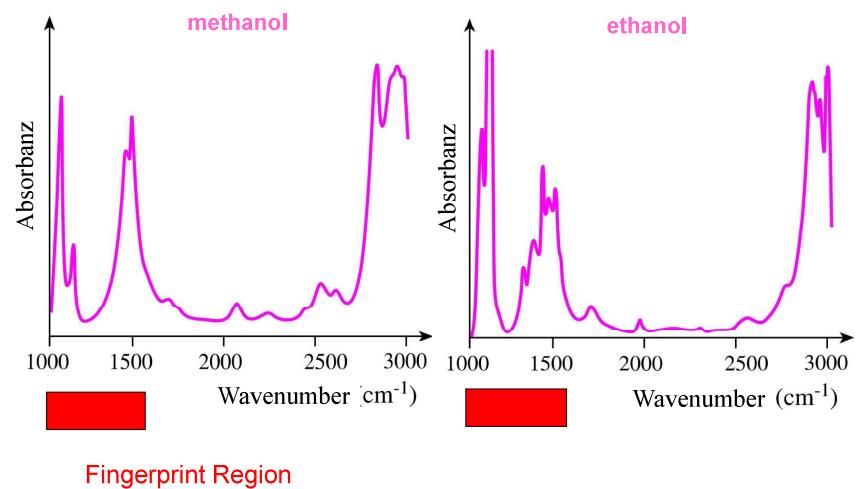
- Ball&Stick Model
- Spacefill Model
- Stick Model
- Motion Off

<http://www2.chemistry.msu.edu/faculty/reusch/VirtTxtJml/Spectpy/Infrared.htm>

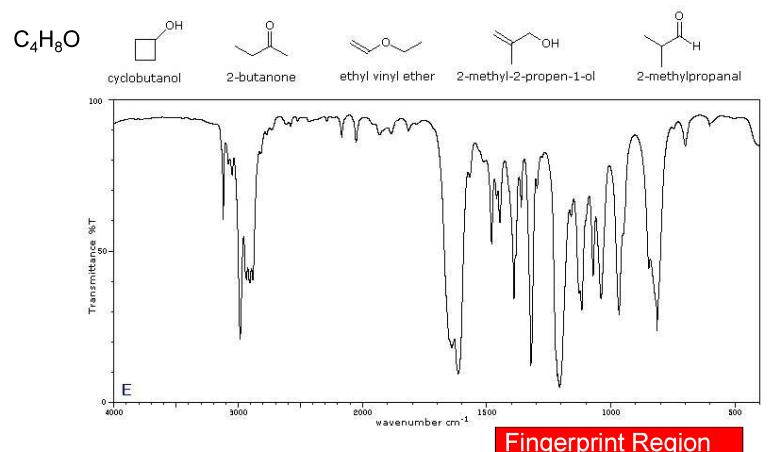
Analytical applications

- synthesis: identification of the intermediate and the end product
- determination and justification of the molecular structure
- detection of the metabolites
- quality control (purity)
- Remark.: Lambert-Beer law is valid, determination of concentration is possible

Identification of the Molecules



Example: Identification of molecules



<http://www2.chemistry.msu.edu/faculty/reusch/VirtTxtJml/Spectry/InfraRed/infrared.htm>

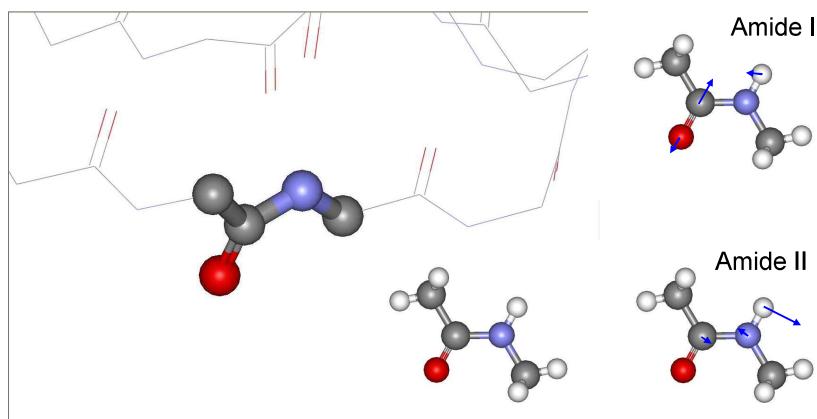
Vibrations of the macromolecules

Complex global vibrations

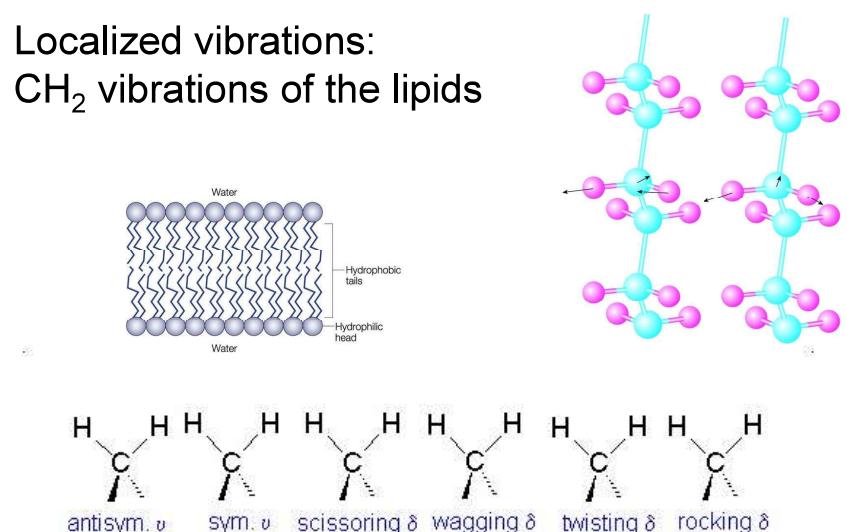
Localized vibrations, e.g.:

- amid vibrations of proteins
(obtained from N-methylacetamide)
- CH₂ vibrations of the lipids

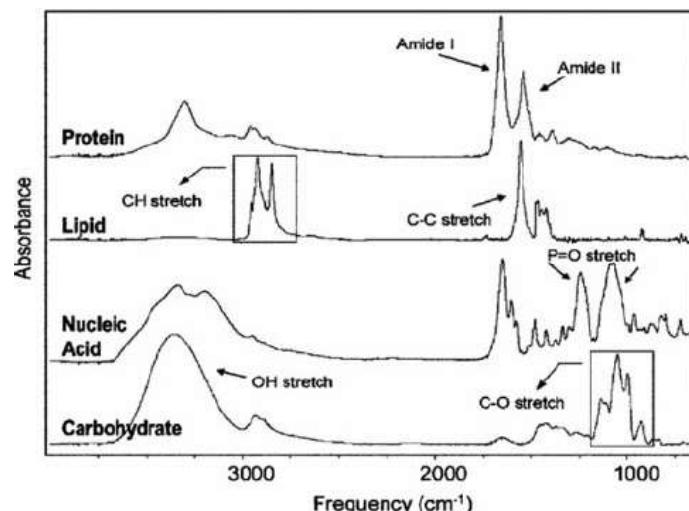
Proteins: model of the polypeptide backbone



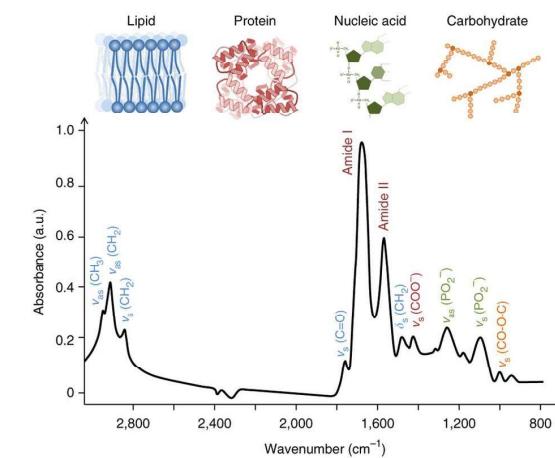
Localized vibrations:
CH₂ vibrations of the lipids



Typical infrared spectra of biological macromolecules



Infrared spectrum of the cell

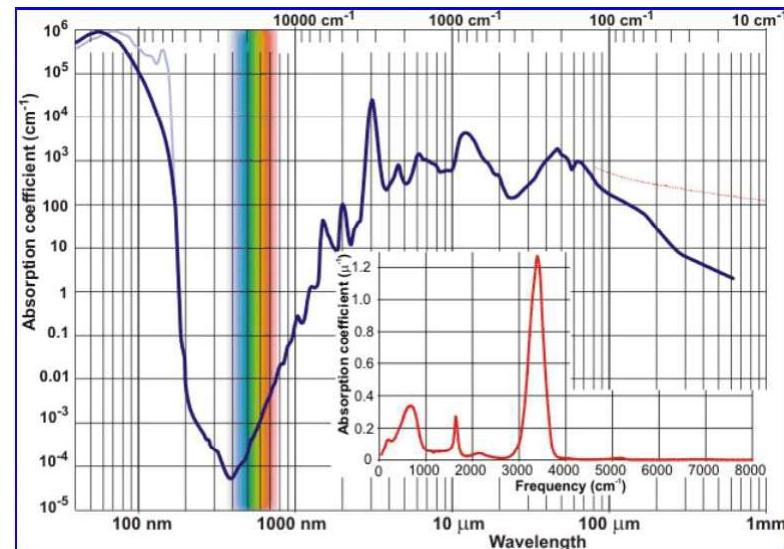


IR spectroscopy of proteins

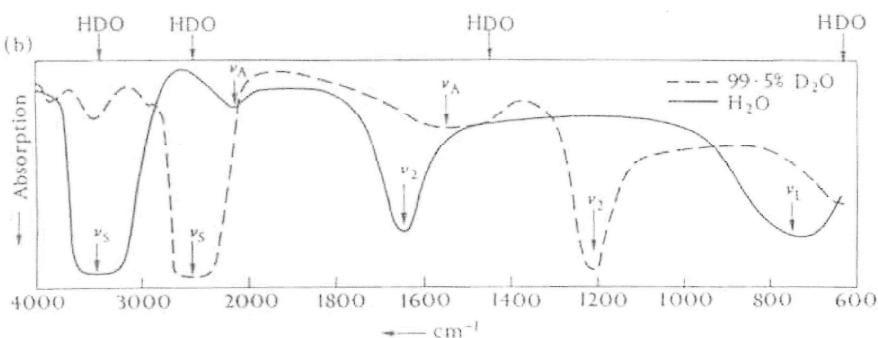
- Backbone: amide vibrations
 - conformation (secondary structure)
 - H/D exchange, (tertiary structure)
- Side chains
 - interaction with other molecules
 - Ca²⁺ binding

Technical point: heavy water solutions (D₂O)

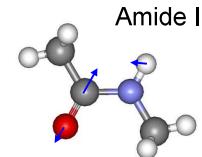
Absorption spectrum of the water



Absorption spectra of water and heavy water

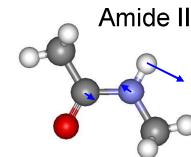


Amide vibrations of proteins



amide I

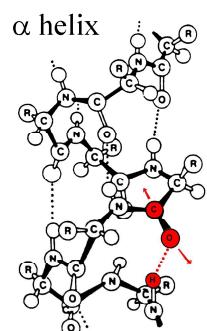
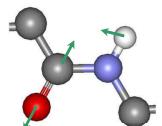
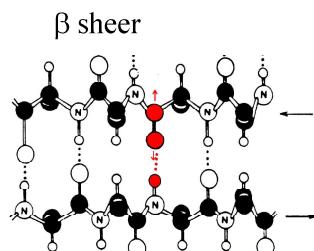
C=O vibration.
Sensitive to the secondary structure because of the hydrogen bond.



amide II

N-H in plane bending vibration
Sensitive to the H-D exchange i.e. to the compactness of the molecule.
(tertiary structure)

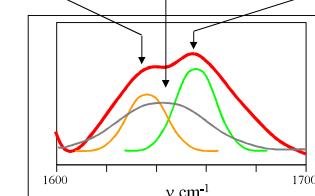
The amide I vibration and the secondary structure



Position of the amide I band in different secondary structures

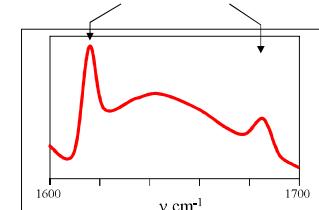
Intramolecular structure

β -sheet unordered α -helix

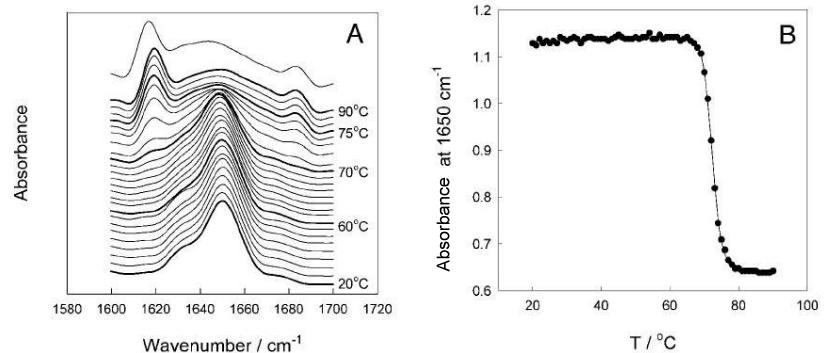


Intermolecular interaction

Intermolecular antiparallel β -sheet

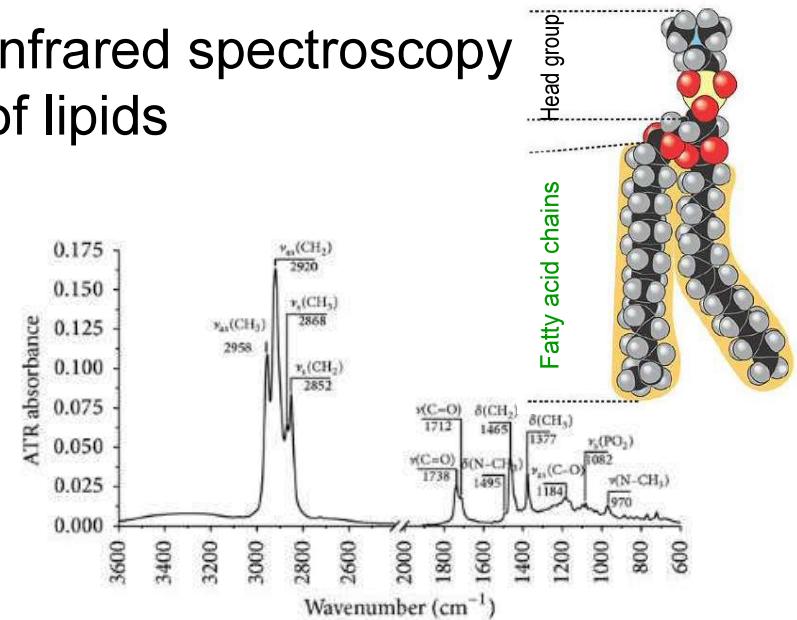


Applications: Denaturation

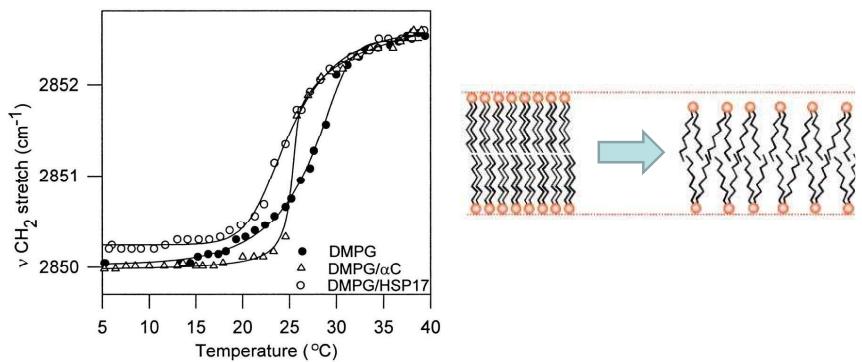


Meersman et al. Biophys J.

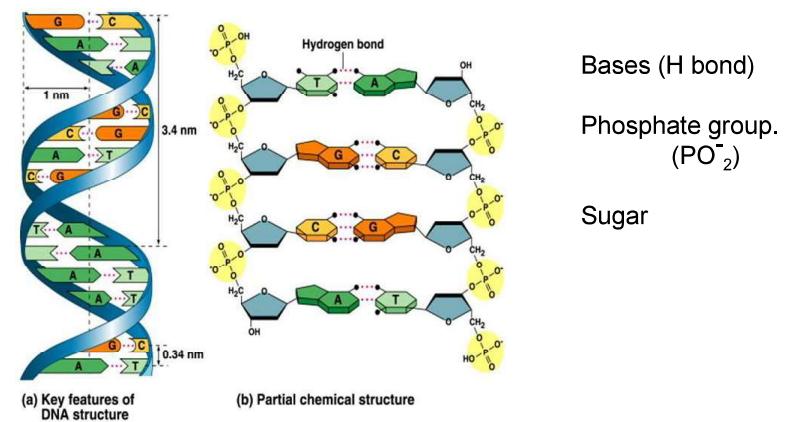
Infrared spectroscopy of lipids



Applications: lipid phase transitions



Nucleic acids



DNA

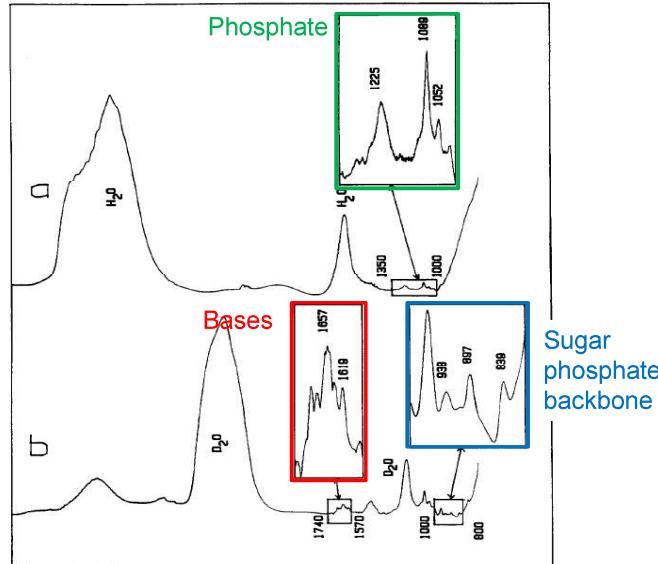


FIG. 1. FT-IR spectra of DNA in solution. (a) H_2O solution; (b) D_2O solution. Enlarged parts of the spectra present the absorptions involving mainly the vibrations of the phosphate groups (top), the double bonds of the bases in their plane (bottom left), and the sugar-phosphate backbone (bottom right).

DNA, RNA A and B structures

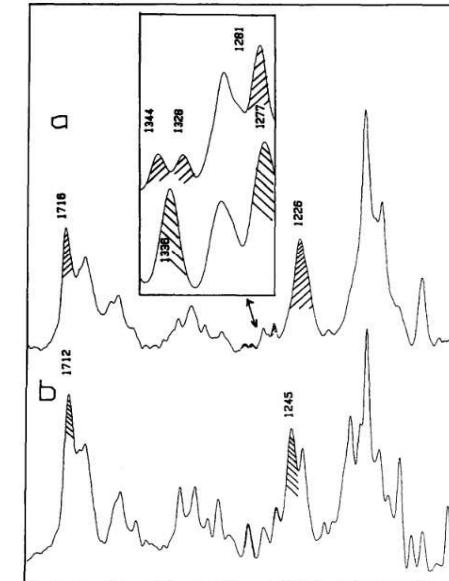
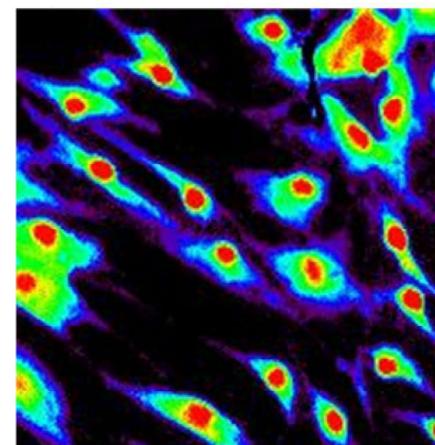
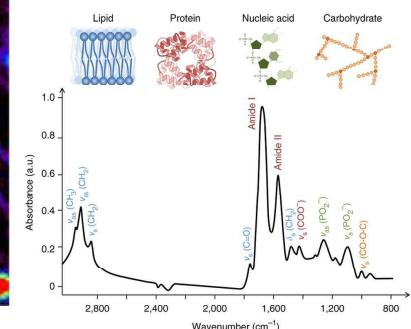


FIG. 3. FT-IR spectra of H_2O solutions of $d(\text{A}-\text{U})_n$ (a) and $r(\text{A}-\text{U})_n$ (b). The enlarged area between 1350 and 1270 cm^{-1} shows absorptions characteristic of A (\\\\\\\\) and B (\\\\//) geometries.

Special IR methods: IR Microscope



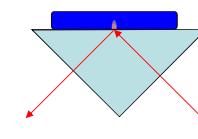
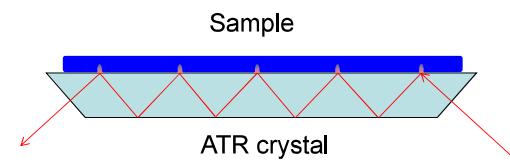
dermal fibroblasts imagined at
1224 cm^{-1}



Portable FTIR spectrometer



ATR technique (Attenuated Total Reflection)

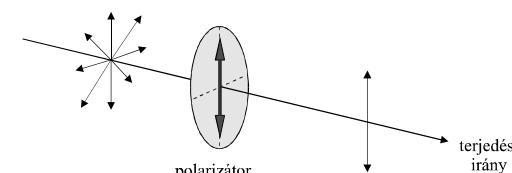


CD

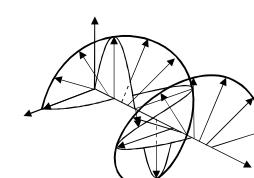
Circular dichroism spectroscopy

Polarized light

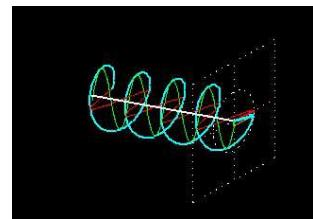
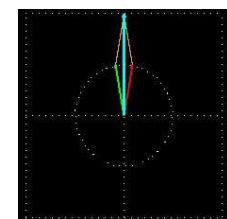
Linear polarized:



Circular polarized



lin. pol light= right + left circ. pol.



Animations can be found here: <http://www.enzim.hu/~szia/cddemo/demo0.htm>

Interaction of chiral molecules with left and right polarized light is different.
The absorbance difference is:

$$\Delta A = A_L - A_R = \Delta \varepsilon c x$$

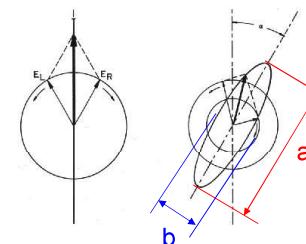
$$\Delta \varepsilon = \varepsilon_L - \varepsilon_R$$

$$\text{Ellipticity: } \theta \quad \tan \theta = b/a$$

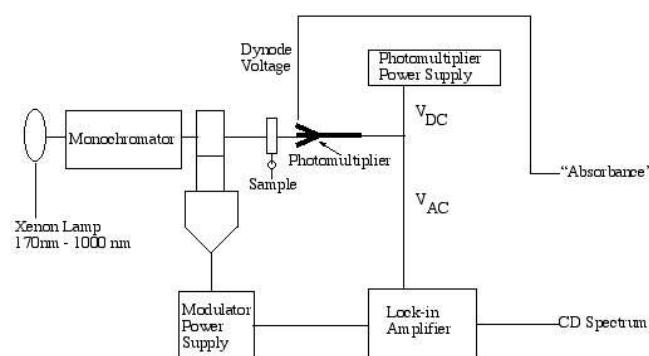
$$\theta = \frac{2.303}{4} \cdot (A_L - A_R) \cdot \frac{180}{\pi} \quad [\text{deg}]$$

$$\text{Lambert-Beer like law: } \theta = c \cdot l \cdot \theta_m$$

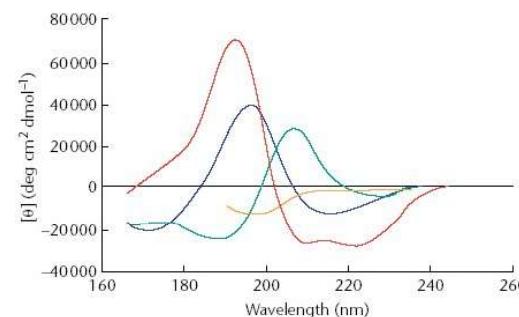
(θ_m : molar ellipticity)



The CD spectrometer

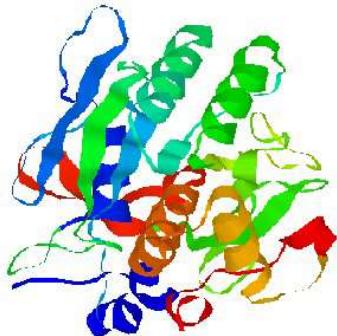


CD and protein structure

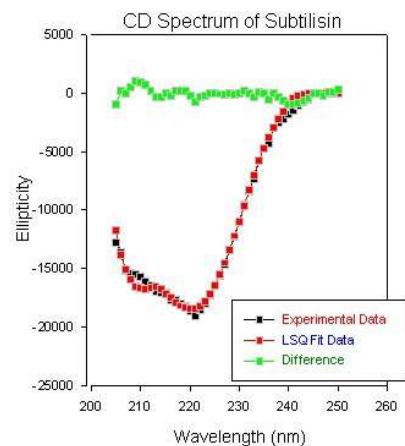


The far-UV CD spectra associated with various types of secondary structure elements in proteins. Red: α -helix; blue: antiparallel β -sheet; green: type I β -turn; orange: irregular structure.
(Data taken from the Encyclopedia of Life Sciences)

The Structure and CD spectrum of Subtilisin



helix sheet coil
57.92 26.22 15.85



A) triosephosphate isomerase

B) hen egg lysozyme

C) myoglobin

D) chymotrypsin

red: helix.

green: strand,

yellow: other.

