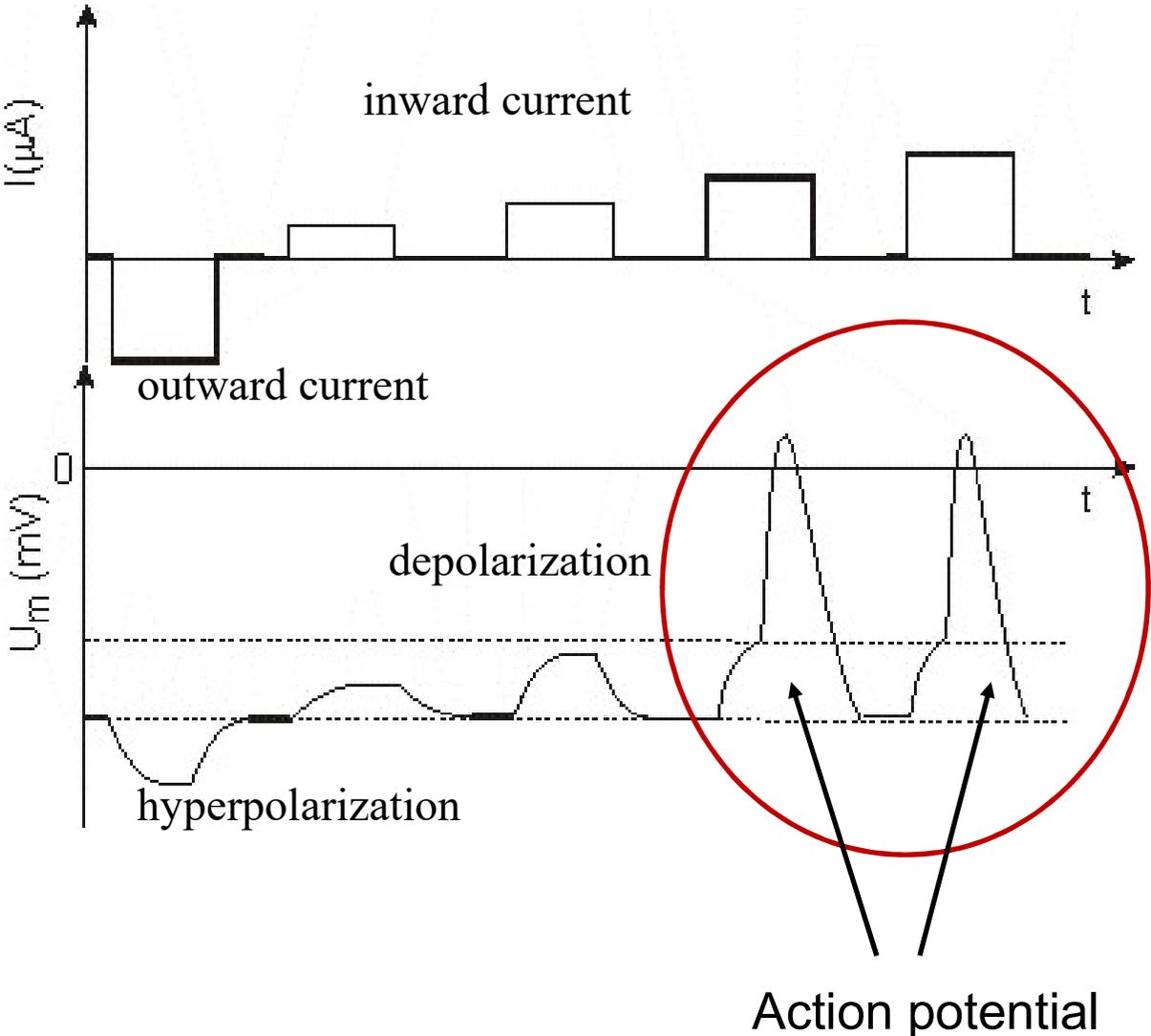


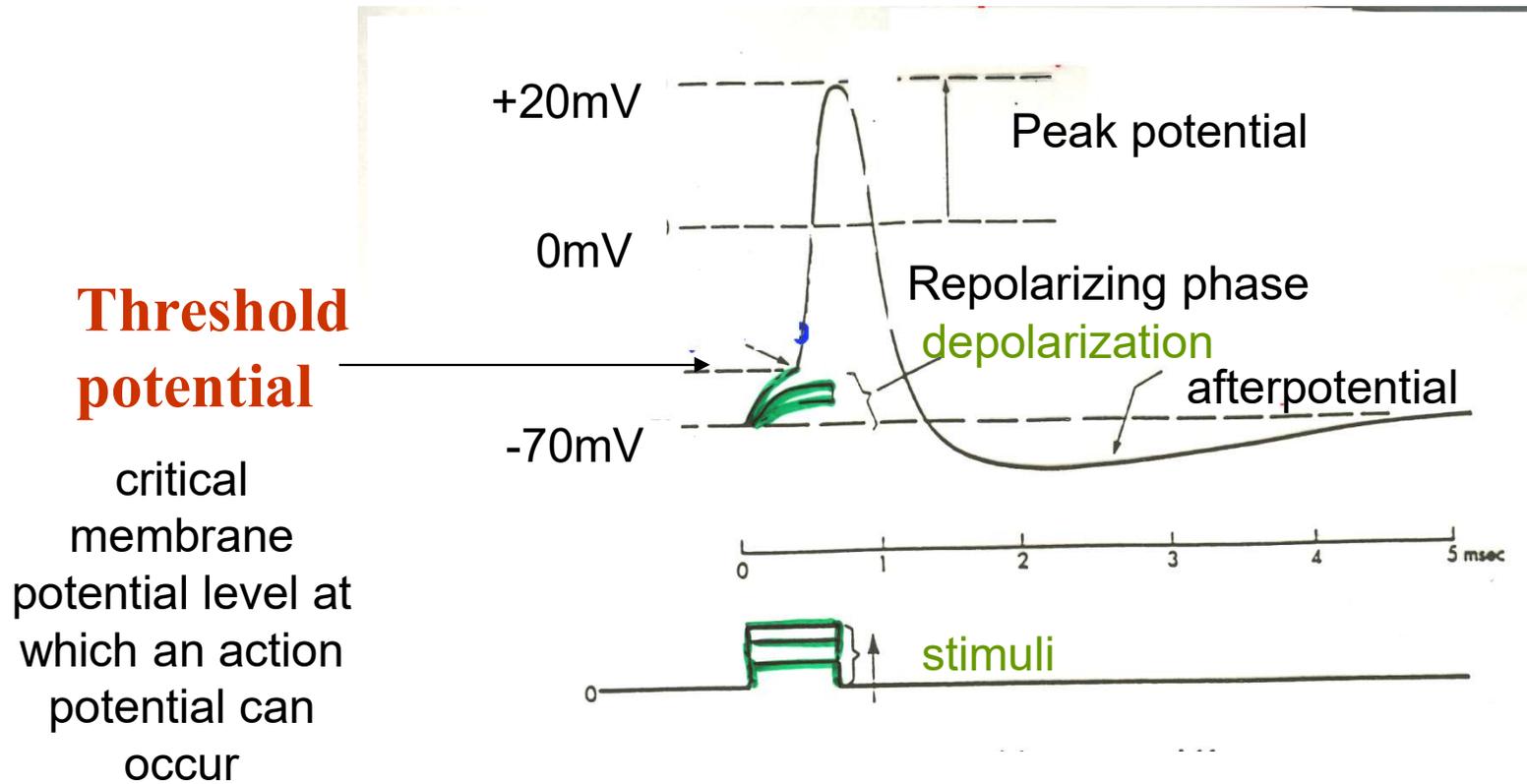
## *Alteration of resting membrane potential*

2. “active” electric properties of the membrane in excited state

# Observation



# *Phases and landmark of the action potential*



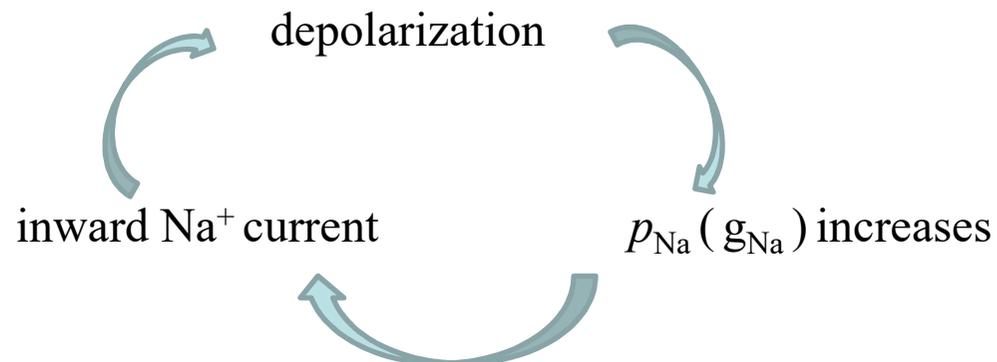
facultative

“All-or-none” amplitude  
conducted with constant amplitude

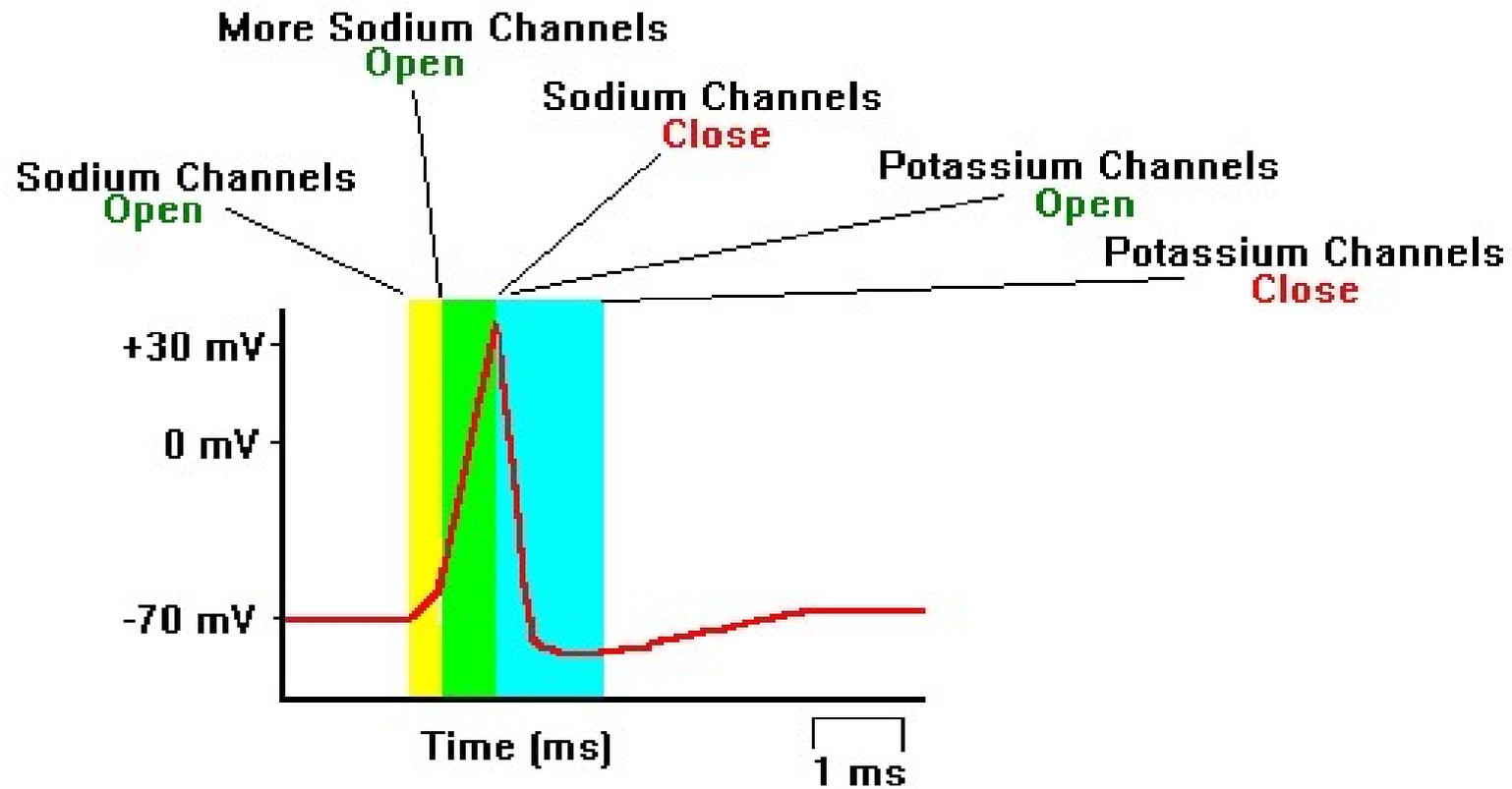
# Hodgkin-Katz hypothesis of action potential generation

Voltage-gated, potential sensitive ion channels


$$\varphi_e - \varphi_i = -\frac{RT}{F} \ln \frac{\sum p_k^+ c_{ke}^+ + \sum p_k^- c_{ki}^-}{\sum p_k^+ c_{ki}^+ + \sum p_k^- c_{ke}^-}$$

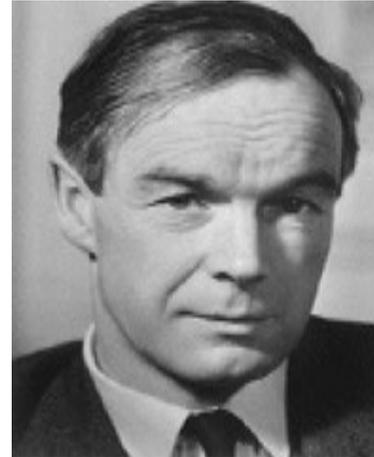


# Hodgkin-Katz hypothesis of action potential sequence





**Andrew Fielding Huxley  
(1917-2012 )**

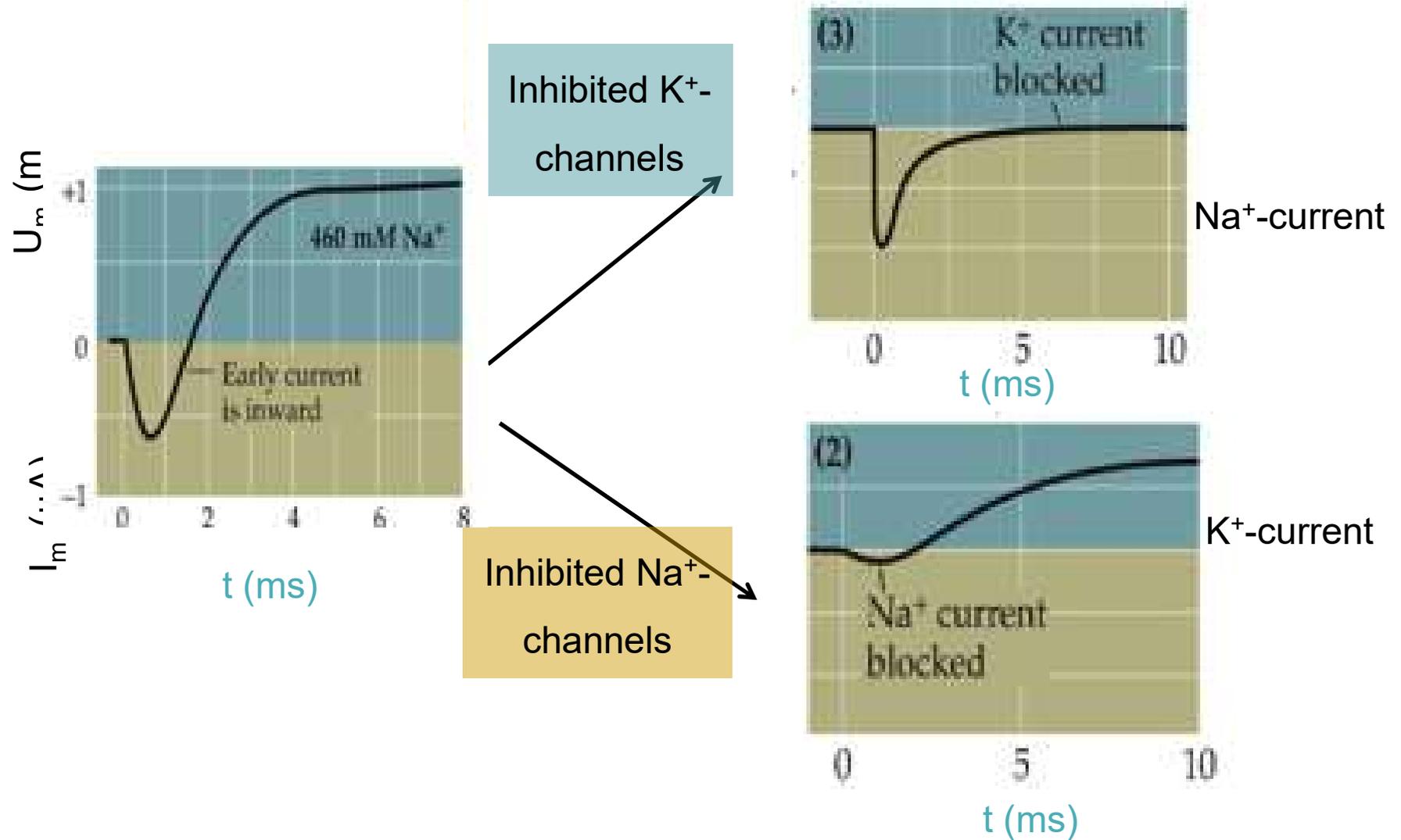


**Alan Loyd Hodgkin  
(1914-1998)**

The Nobel Prize in Physiology or Medicine  
1963

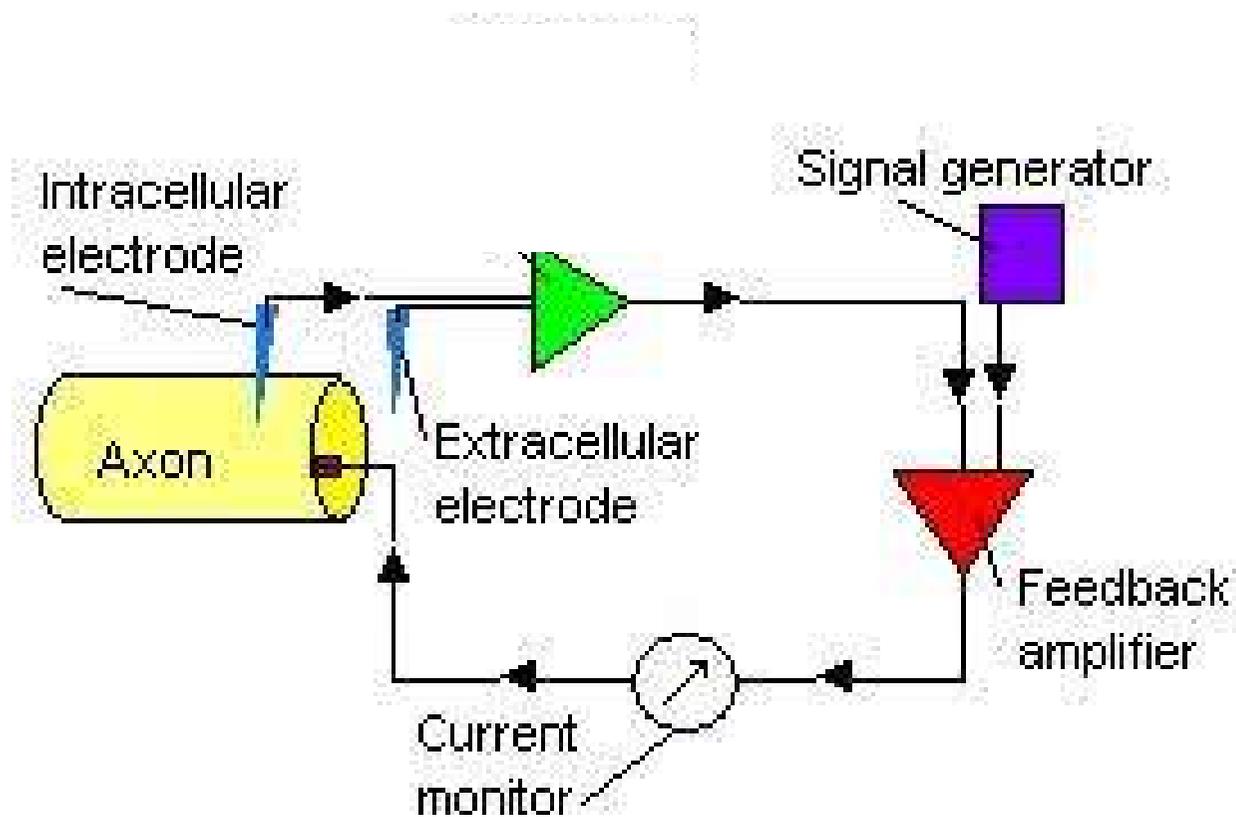
“for their discoveries concerning the ionic mechanisms involved in excitation and inhibition in the peripheral and central portions of the nerve cell membrane”

# Measurement of separated ionic currents



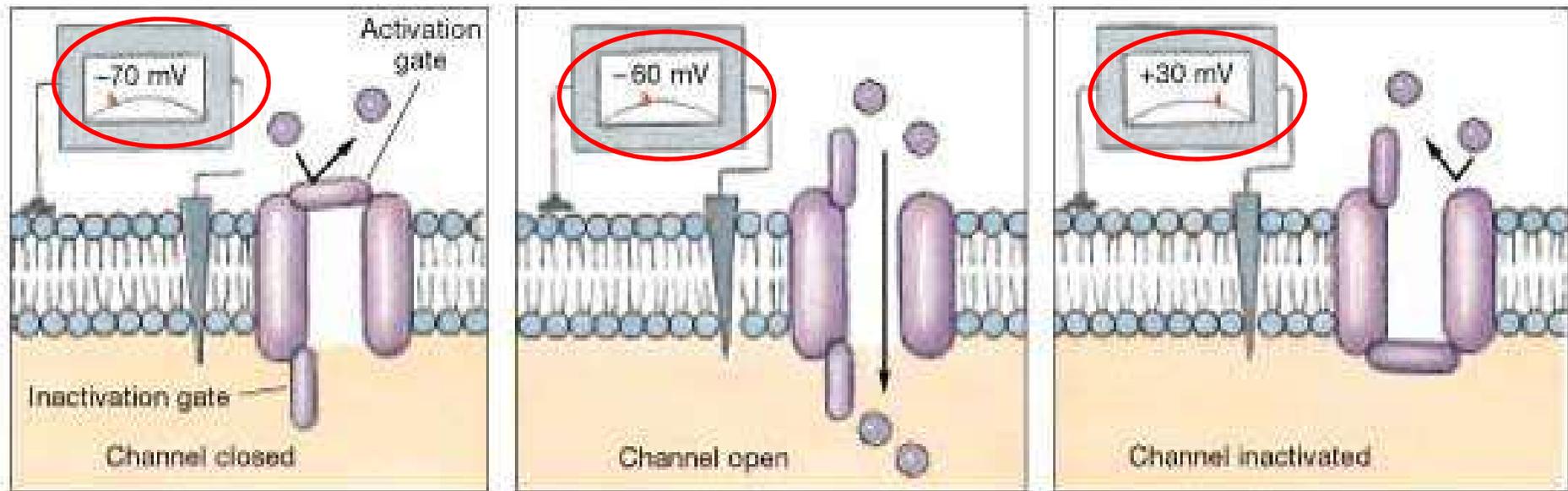
Voltage-Gated Na<sup>+</sup> and K<sup>+</sup> Channels

# Voltage Clamp



- Membrane potential is kept constant
- ion-current is monitored

# States of voltage-gated sodium channels

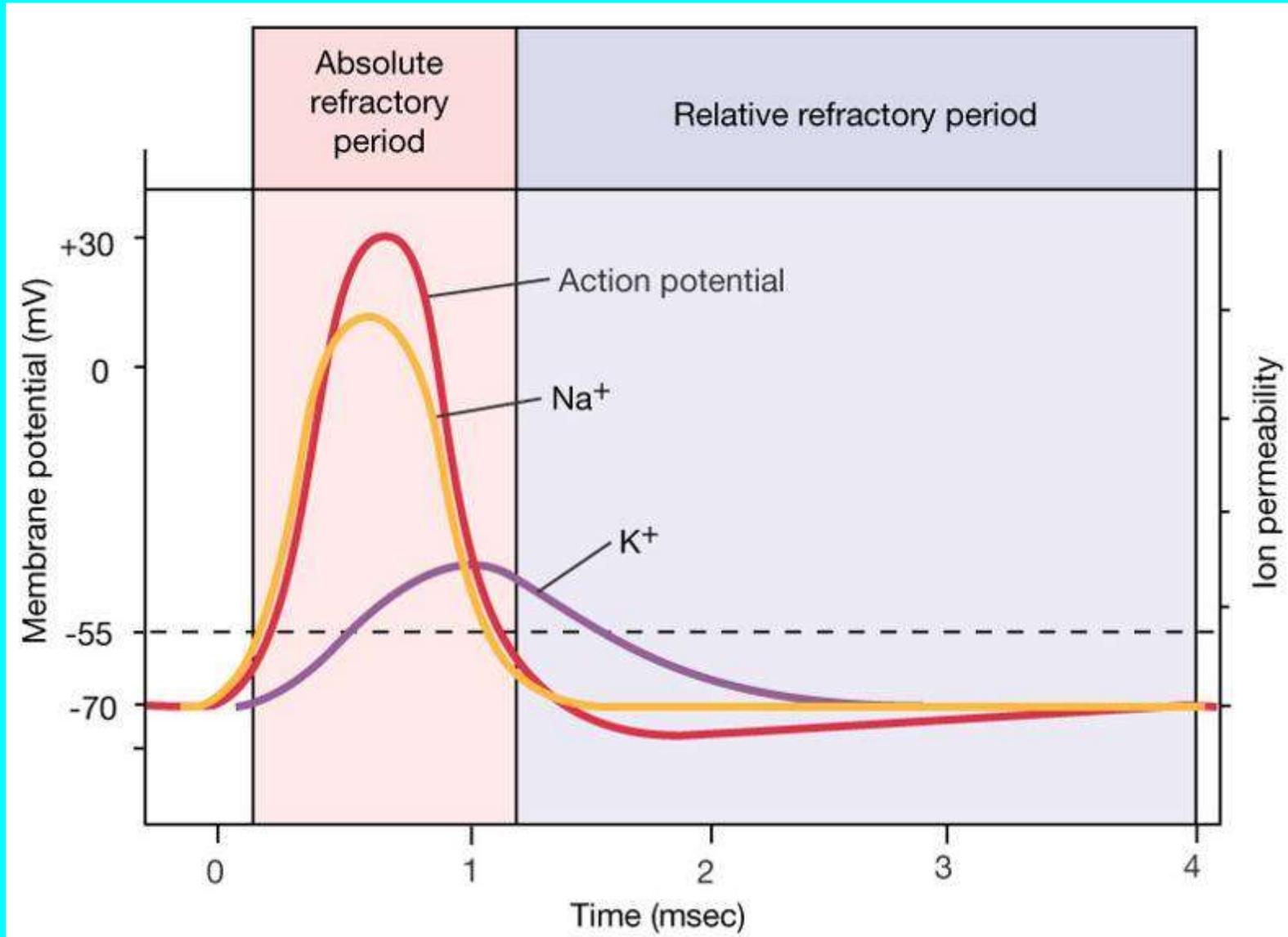


(c)



at depolarization threshold

# Conductivities during action potential

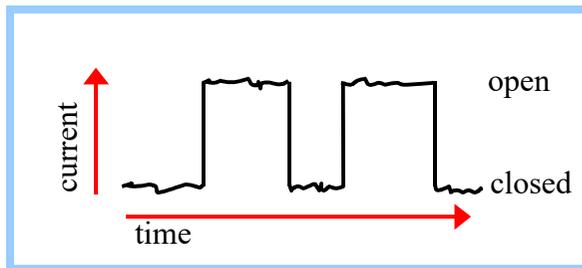


# Patch-Clamp technika

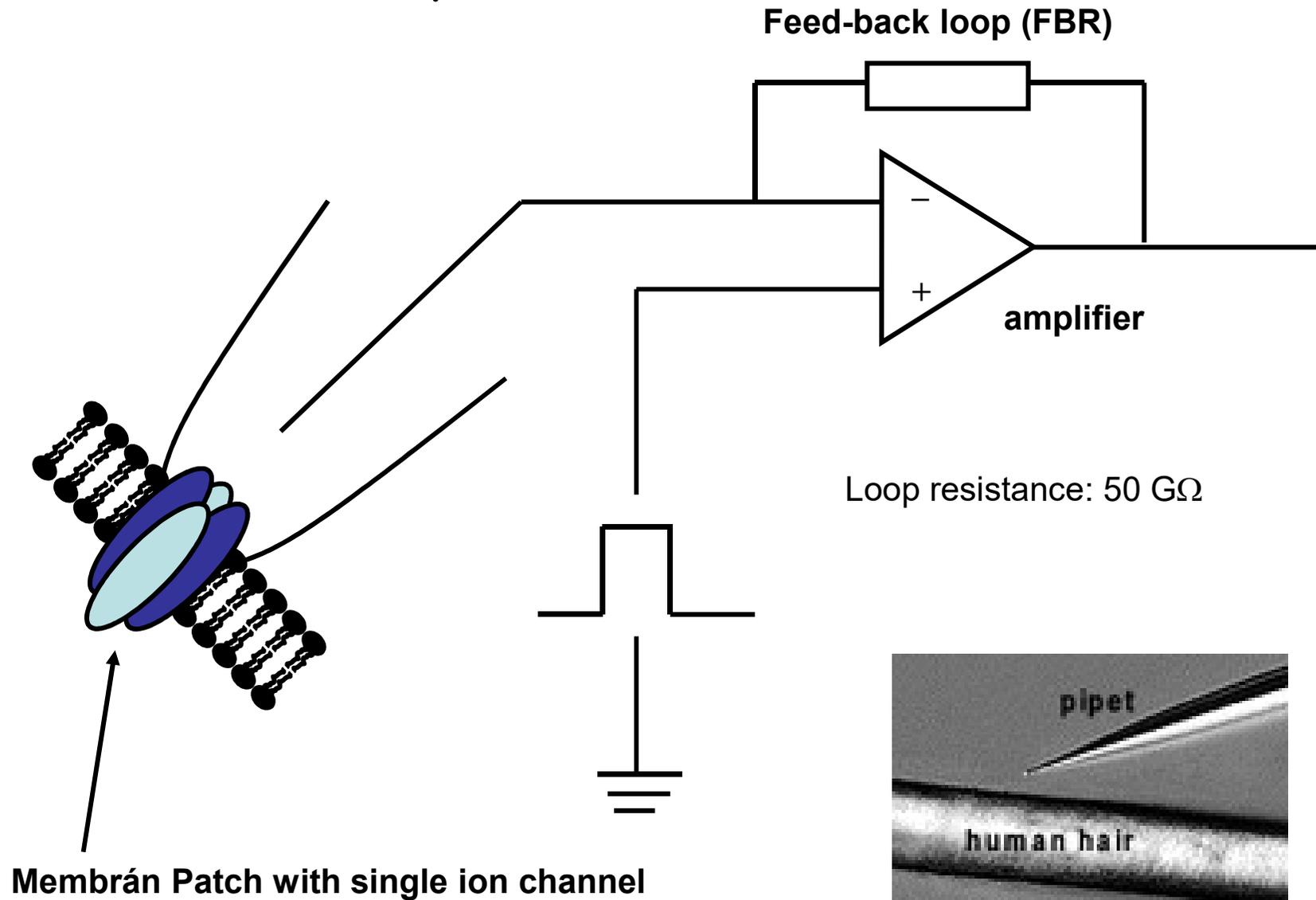
Ion current in single membrane channel ( $10^{-12}$  A)

The Nobel Prize in Physiology or Medicine  
1991 was awarded jointly to Erwin Neher  
(1944- ) and Bert Sakmann (1942- )

*"for their discoveries concerning the  
function of single ion channels in cells"*



# Patch-Clamp circuit



# Patch-Clamp device

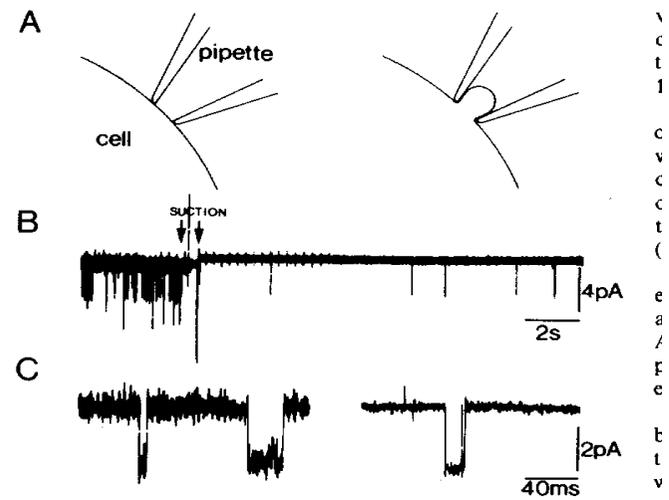
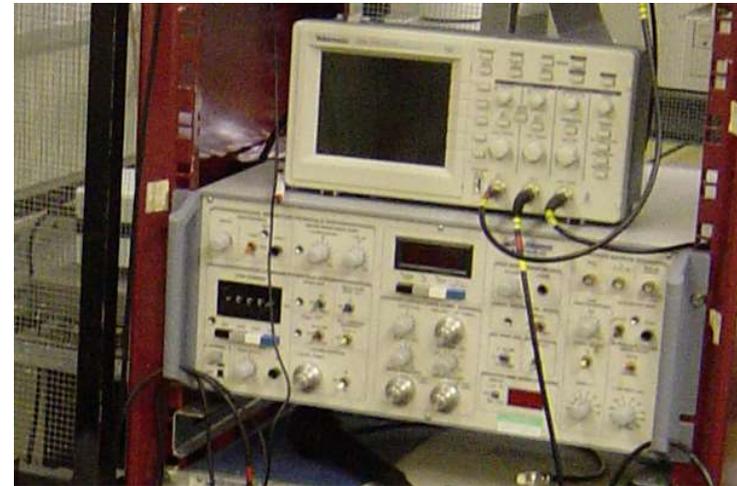
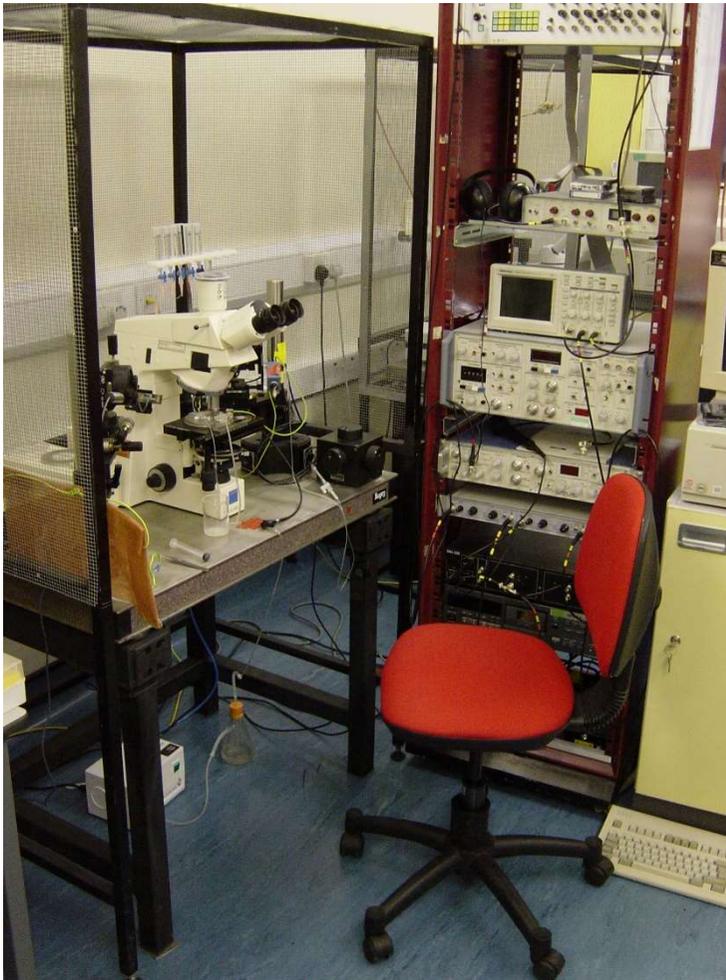
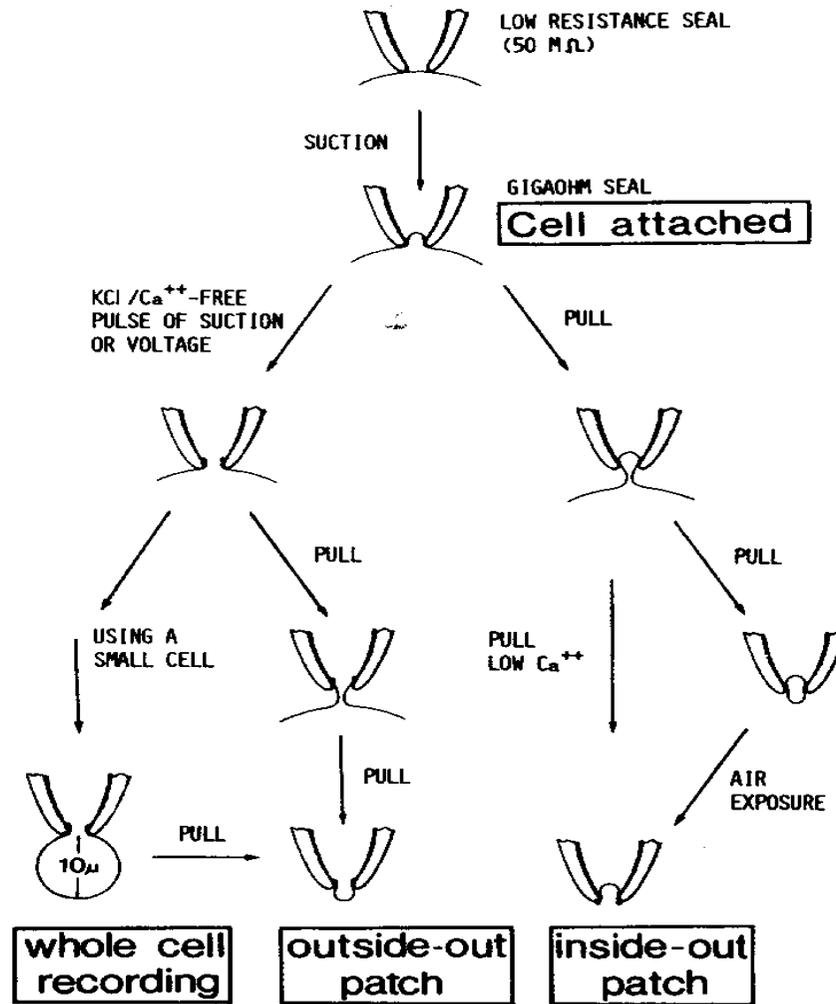


Fig. 6A—C. Giga-seal formation between pipette tip and sarcolemma of

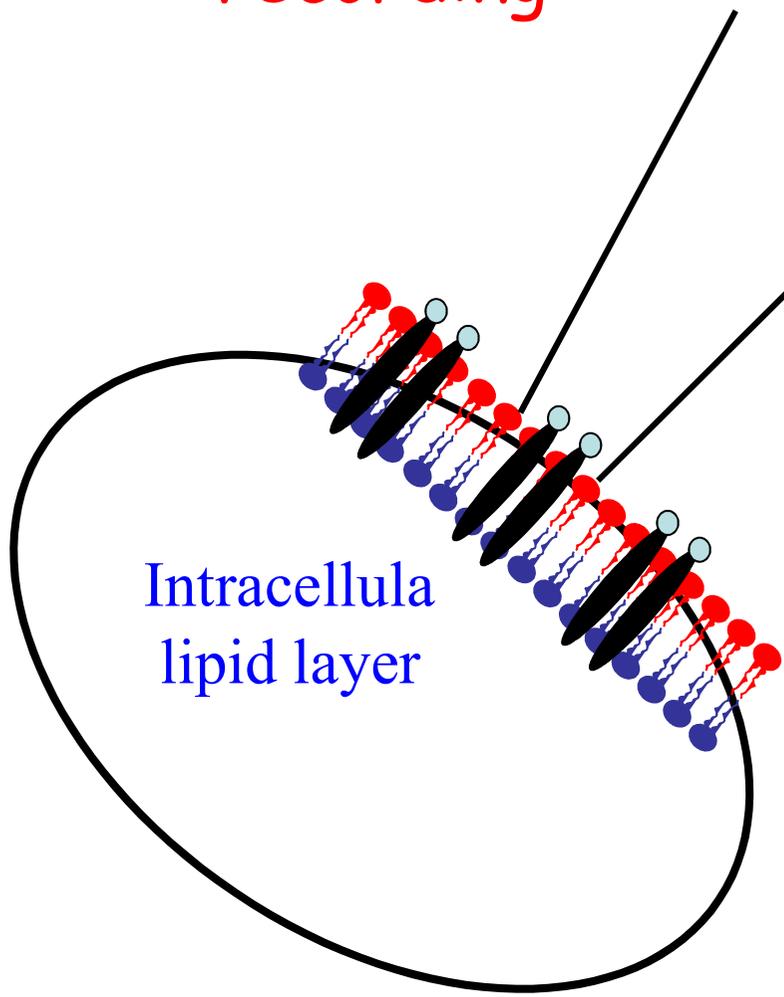
V  
c  
t  
1  
c  
v  
c  
t  
(  
e  
a  
/  
F  
e  
t  
v  
a

# Patch-Clamp arrangements



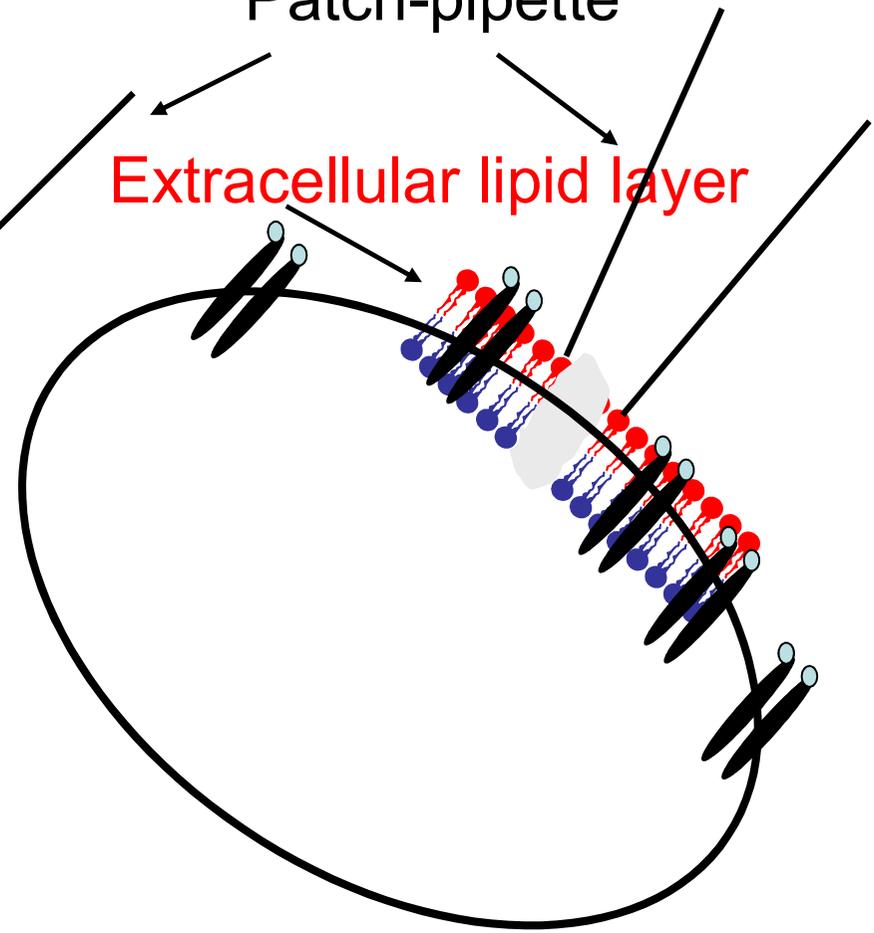
From Hamill *et al* 1981

Cell-attached recording

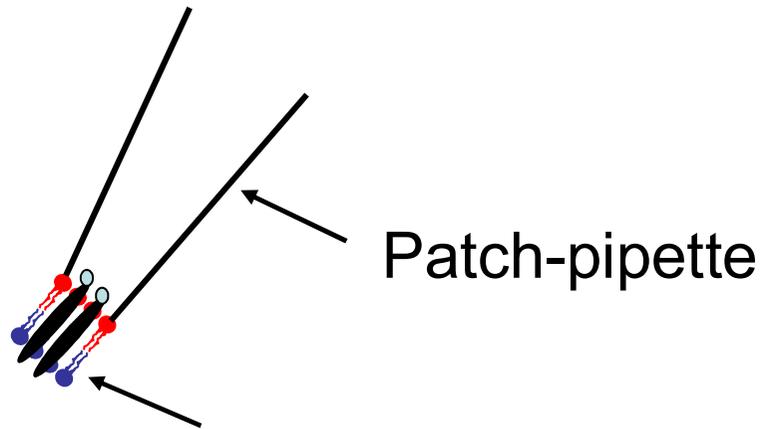


Patch-pipette

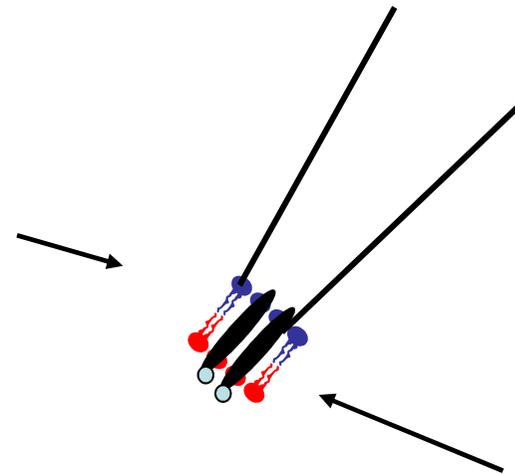
Extracellular lipid layer



## Inside-out recording



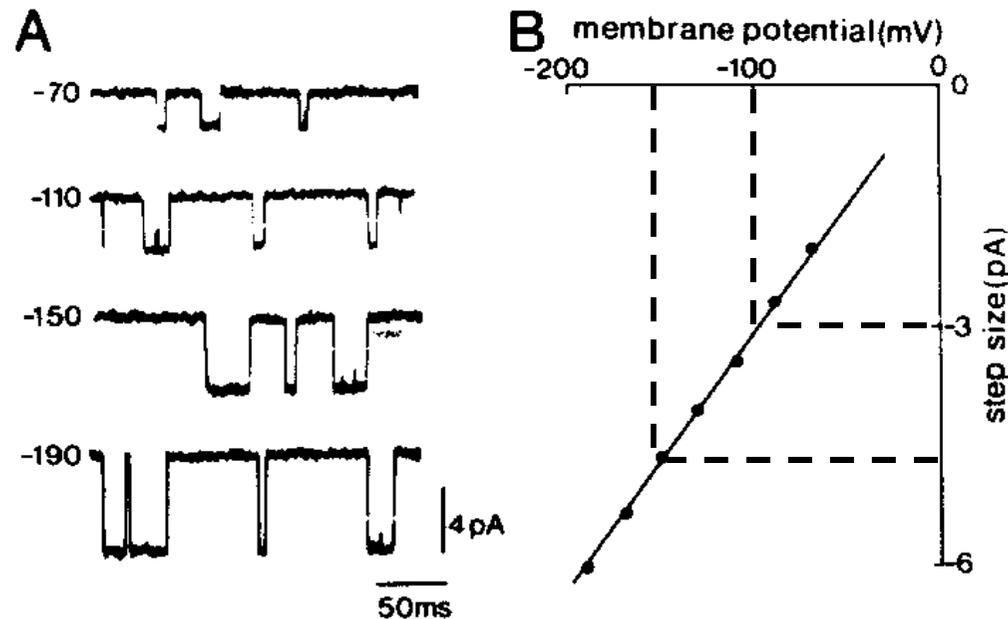
The **internal** face of the lipid bi-layer faces the bath solution



The **external** face of the lipid bi-layer faces the bath solution

Outside-out  
recording

# Single-channel I/V function

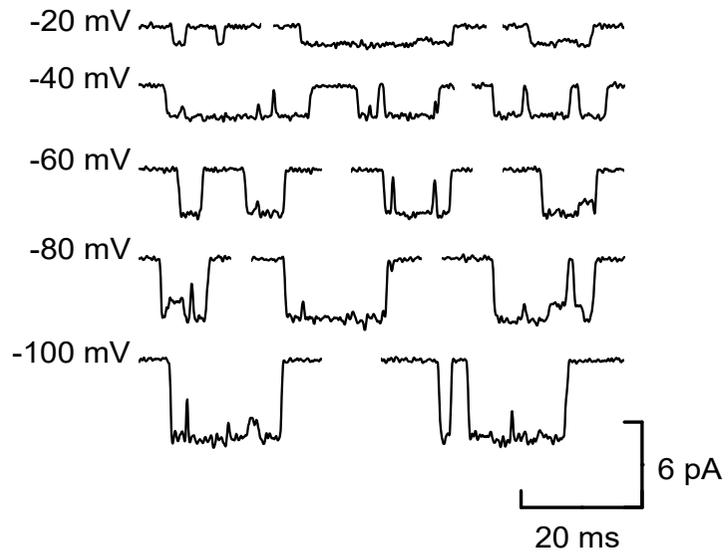


$$\begin{aligned}g_{\text{csatorna}} &= \Delta I \div \Delta V \\ &= 1.6 \times 10^{-12} \text{ A} \div 50 \times 10^{-3} \text{ V} \\ &= 32 \times 10^{-12} \text{ S} \\ &= 32 \text{ pS}\end{aligned}$$

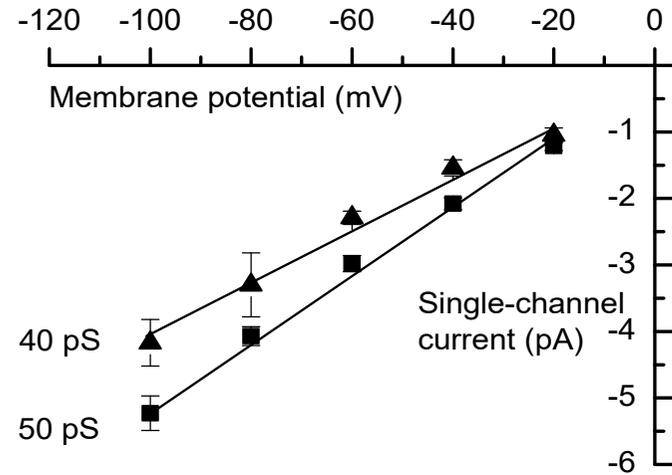
From Hamill *et al* 1981

# Multi-state single channel

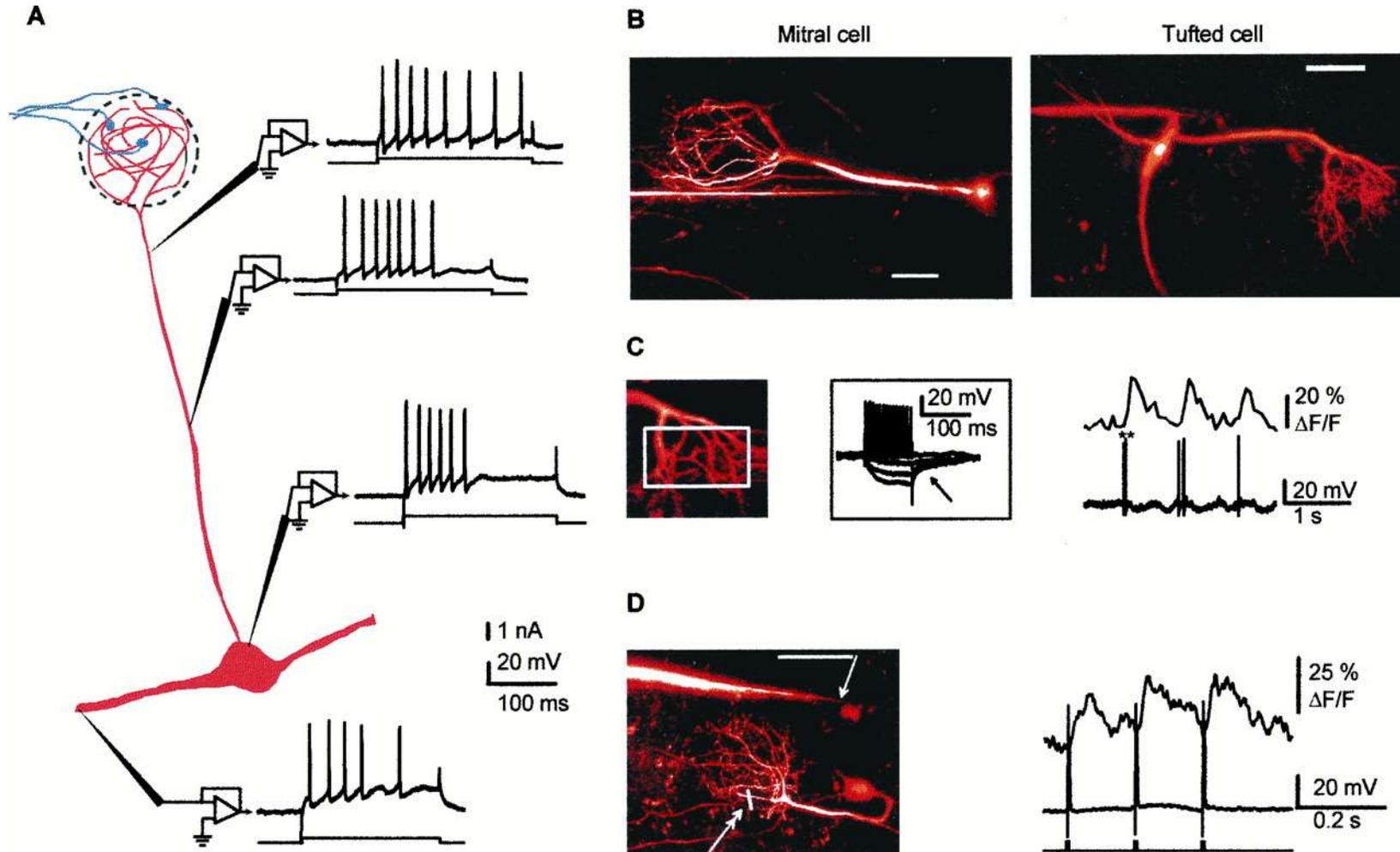
**A**



**B**



**Sodium action potentials synchronize  $[Ca^{2+}]$  transients in all dendritic compartments of mitral cells in the olfactory bulb of anesthetized rats.**

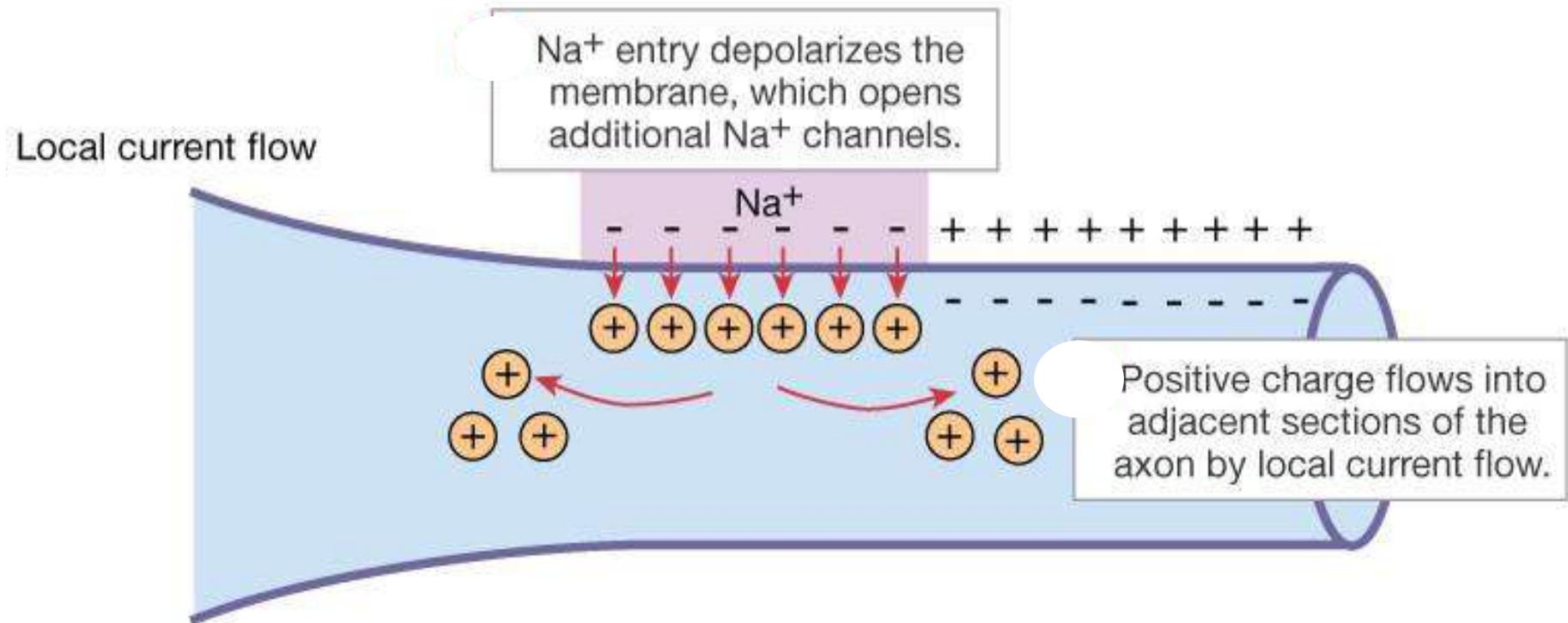


Charpak S et al. PNAS 2001;98:1230-1234

# Factors Influencing Conduction Direction and Velocity

The evolutionary need for the fast and efficient transduction of electrical signals

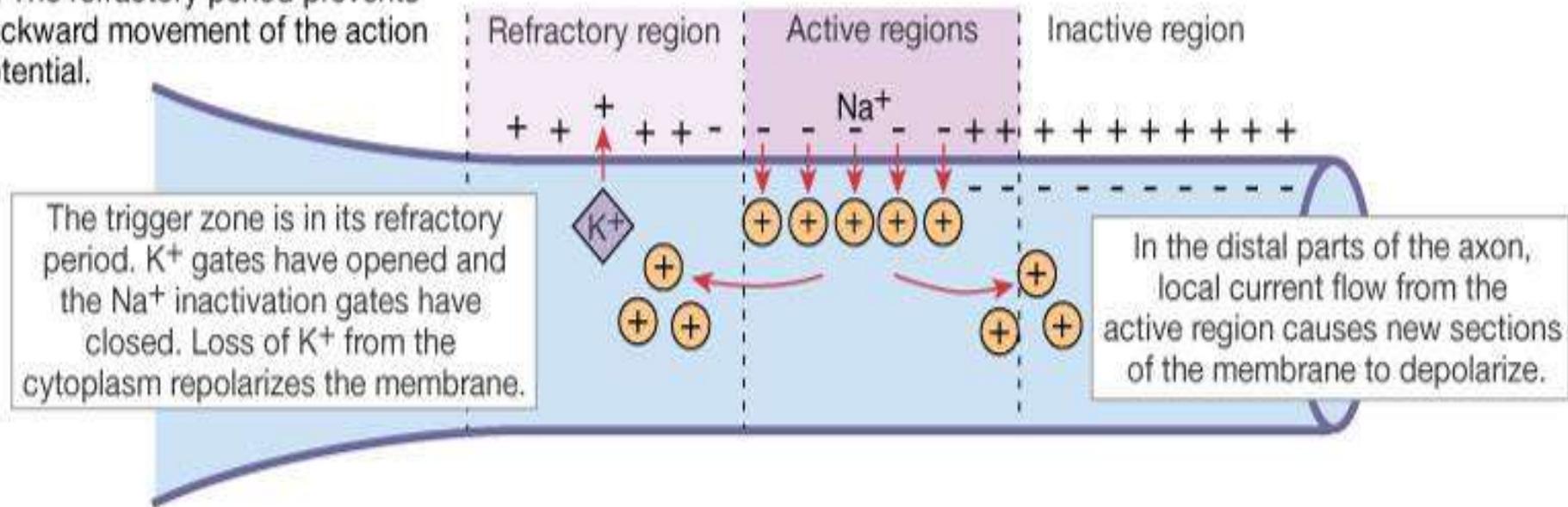
# Propagation of action potential (1)



based on local current flow and depolarization of adjacent membrane area

## Propagation of action potential (2)

(c) The refractory period prevents backward movement of the action potential.

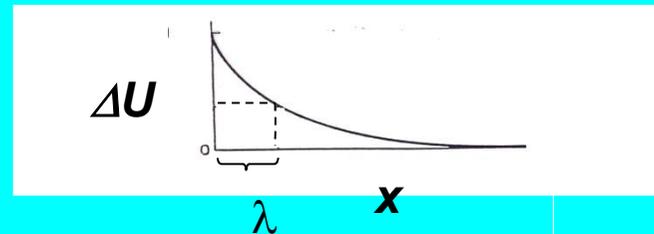


Speed and distance of propagation?

How are the *time constant* and the *space constant* related to propagation velocity of action potentials

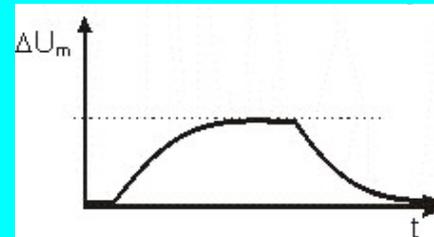
## Generation of the next peak potential

Where?



**The greater the space constant**, the more rapidly distant regions will be brought to threshold and the more rapid will be the propagation velocity

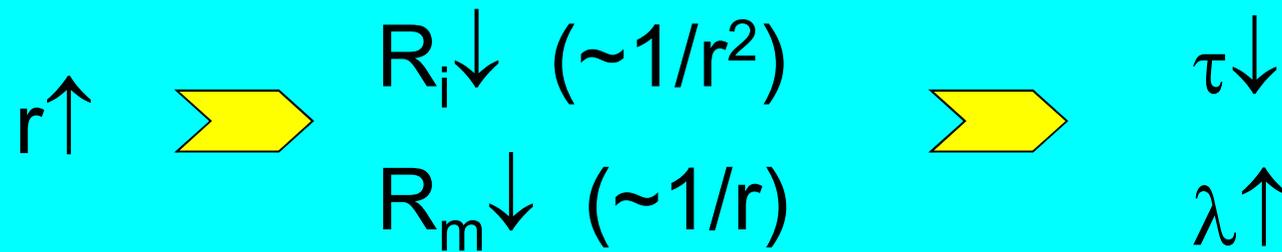
When?



**The smaller the time constant**, the more rapidly a depolarization will affect the adjacent region.

**Velocity is the function of passive properties –  $\tau$  and  $\lambda$  – of membranes**

## *Effect of axon diameter:*

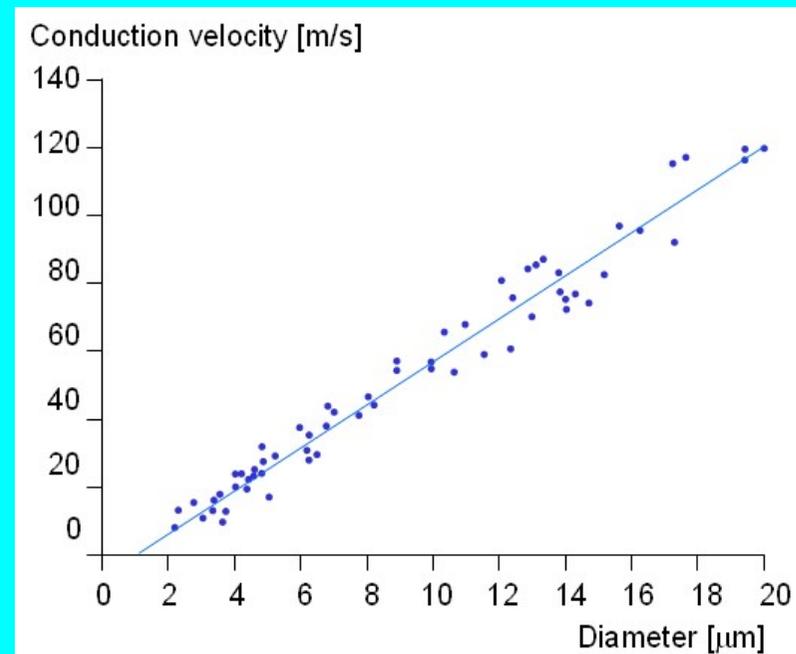


$$\tau = C_m R_m$$

$$\lambda \sim \sqrt{\frac{R_m}{R_i}}$$

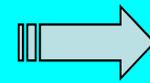
Squid giant axon  $r=250\mu\text{m}$   
 $v=25\text{m/s}$

human nerve cell  $r=10\mu\text{m}$   
 $v \neq 0.5\text{m/s}$  ?



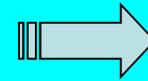
# Myelination!

$R_m$  – very high

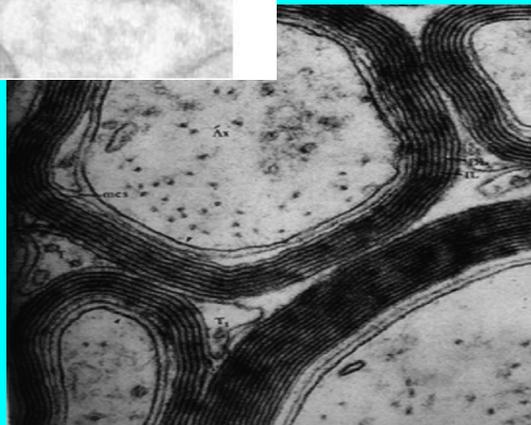
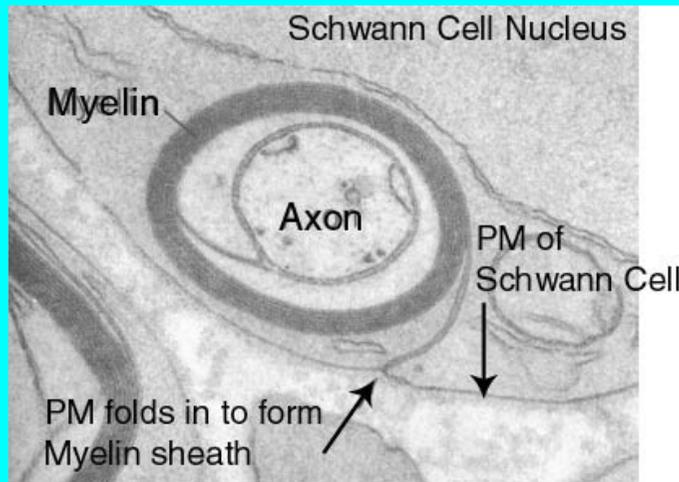


big space constant

$C_m$  – very small



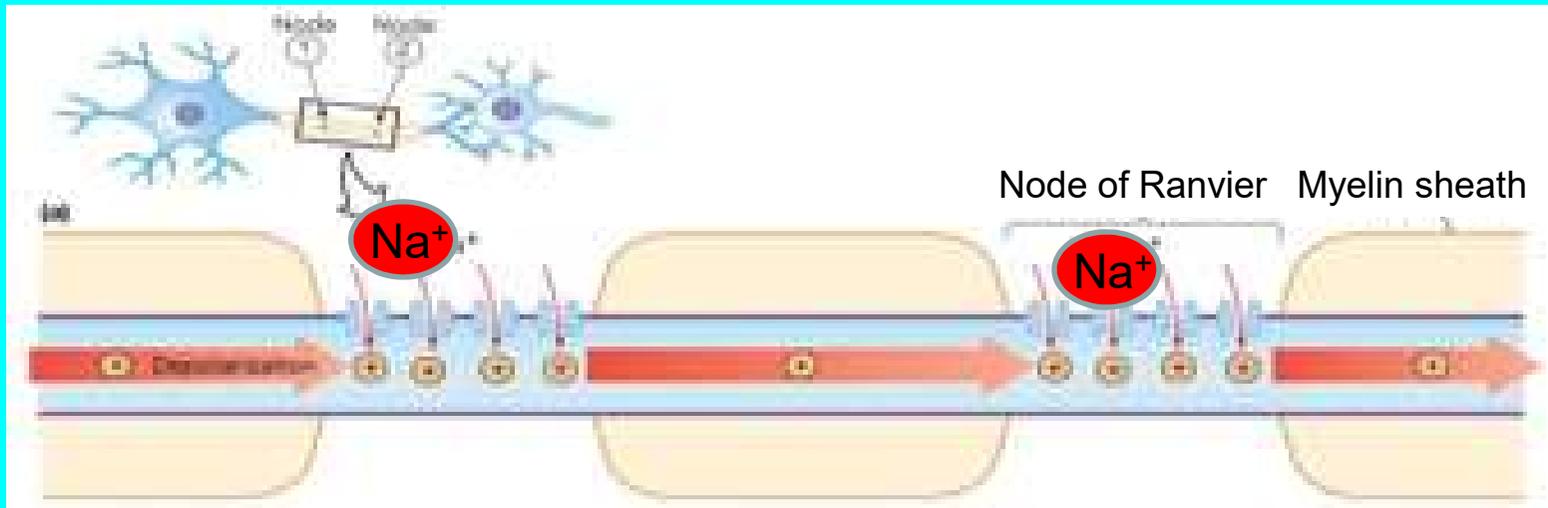
small time constant



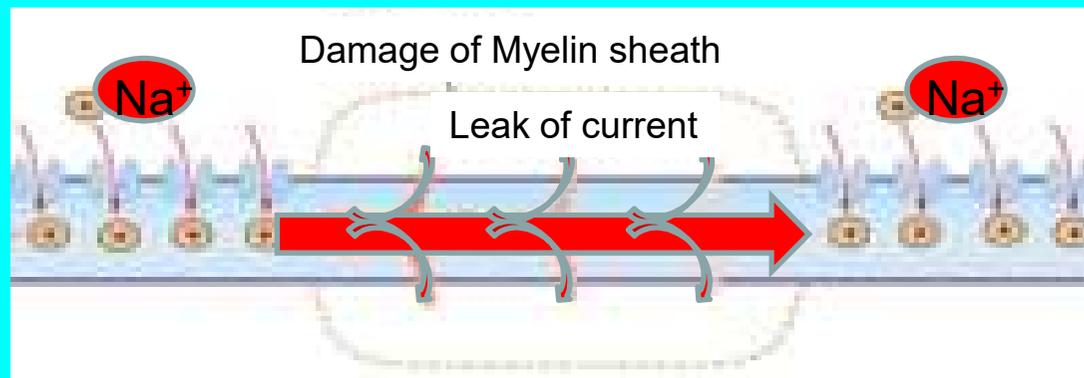
human nerve cell  $r = 10 \mu\text{m}$

$v \sim 100 \text{ m/s}$

# Saltatory conduction - quick, energy saving



Myelin prevents ions from entering or leaving the axon along myelinated segments.



## Effect of axon diameter and Myelination

The diameter of frog axons and the presence or absence of myelination control the conduction velocity.

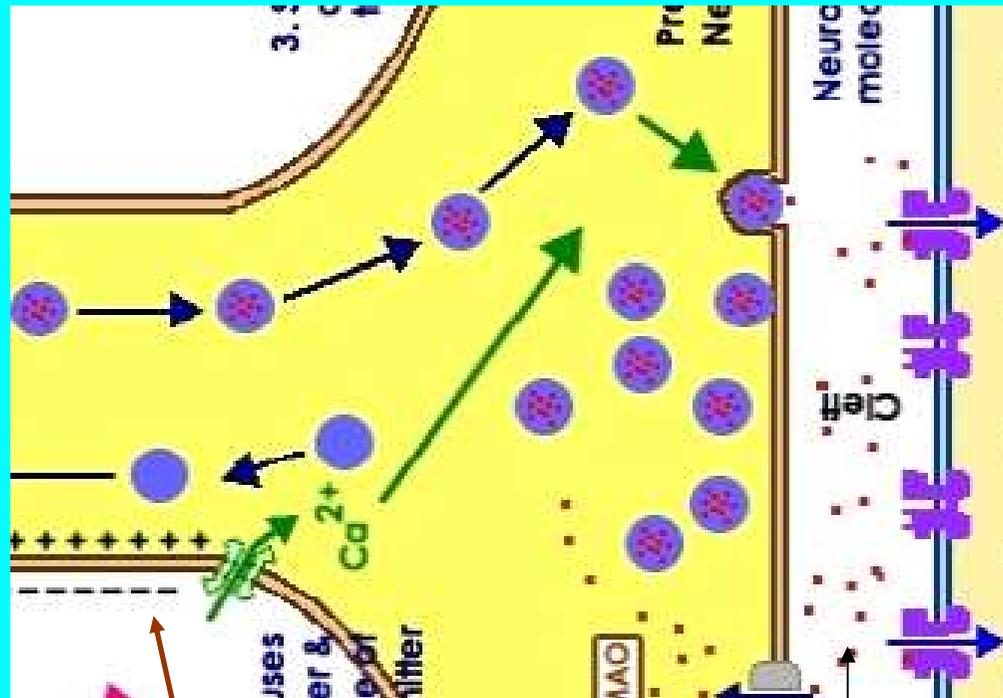
Fiber type	Average axon diameter ( $\mu\text{m}$ )	Conduction velocity ( $\text{m} \cdot \text{s}^{-1}$ )
<b>Myelinated fibers</b>		
A $\alpha$	18.5	42
A $\beta$	14.0	25
A $\gamma$	11.0	17
B	Approximately 3.0	4.2
<b>Unmyelinated fibers</b>		
C	2.5	0.4–0.5

Effect of passive electric properties on signal  
transduction in synapses

# Signal transmission in synapses

presynaptic terminal

postsynaptic terminal

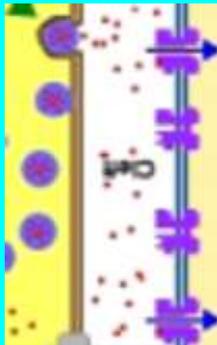


Action potential

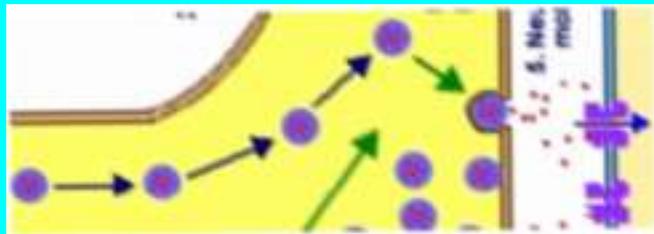
neurotransmitter

How can neurons transmit information from presynaptic to postsynaptic cells **if most synaptic effects are subthreshold?**

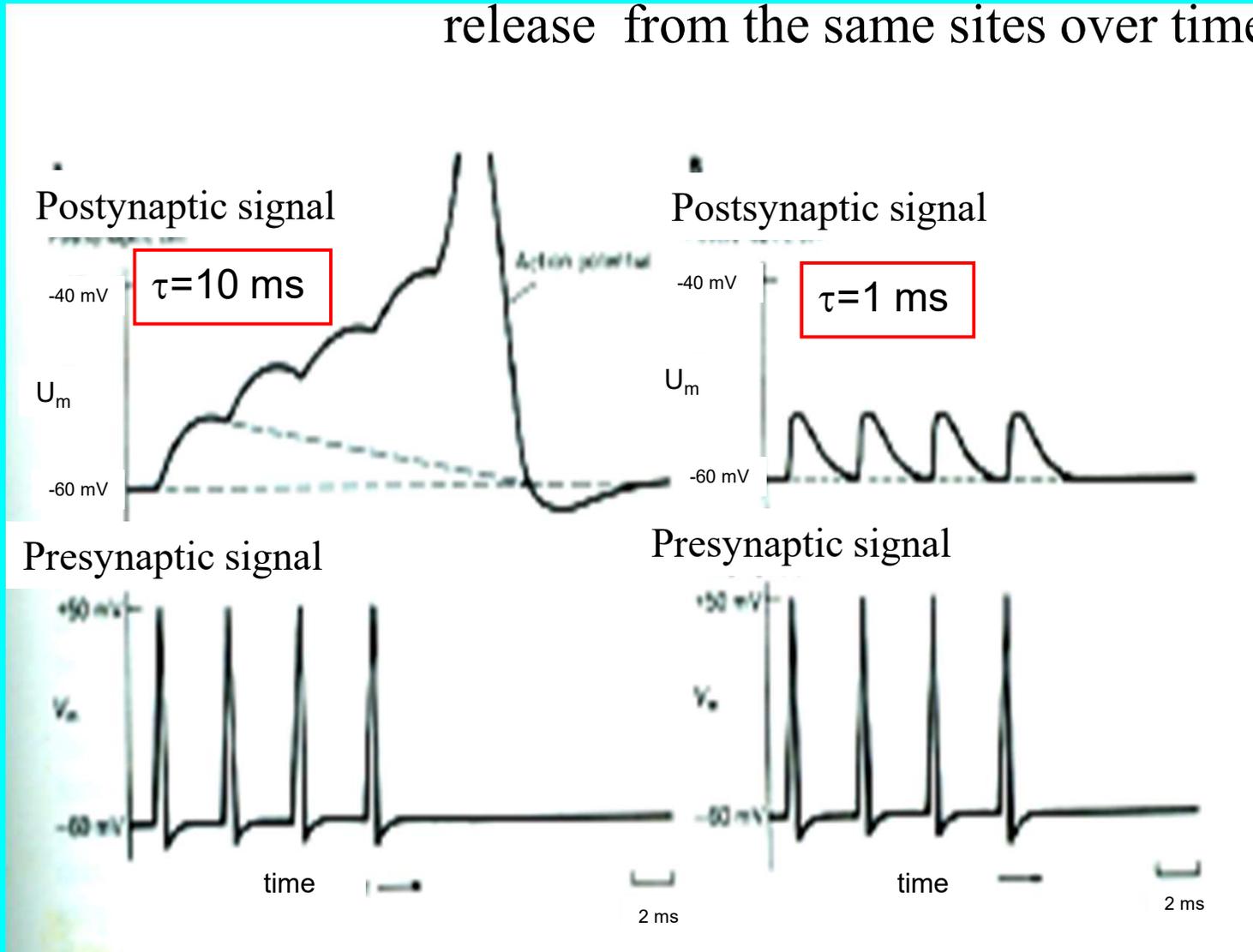
**Spatial Summation** : combined influences at the same cell at a particular moment in time



**Temporal Summation** : combined effects of neurotransmitter release from the same sites over time



**Temporal Summation** : combined effects of neurotransmitter release from the same sites over time



# Temporal and spatial summation

