Bioelectric phenomena.

- **Related practices: Amplifier, Pulse generator, ECG**
- **Related book chapters: III/4** ***Textbook: pp. 276-300.***

Balázs Kiss

kissb3@gmail.com

Nanobiotechnology and Single Molecule Research Group and

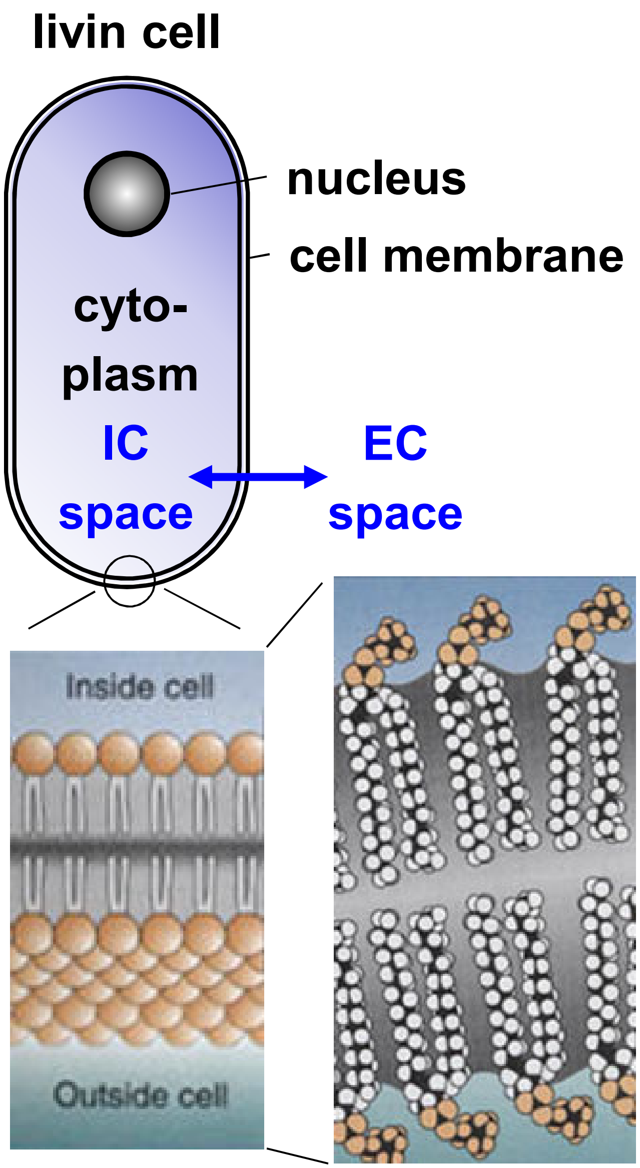
Muscle Mechanobiophysics Group

Department of Biophysics and Radiation Biology,

Semmelweis University

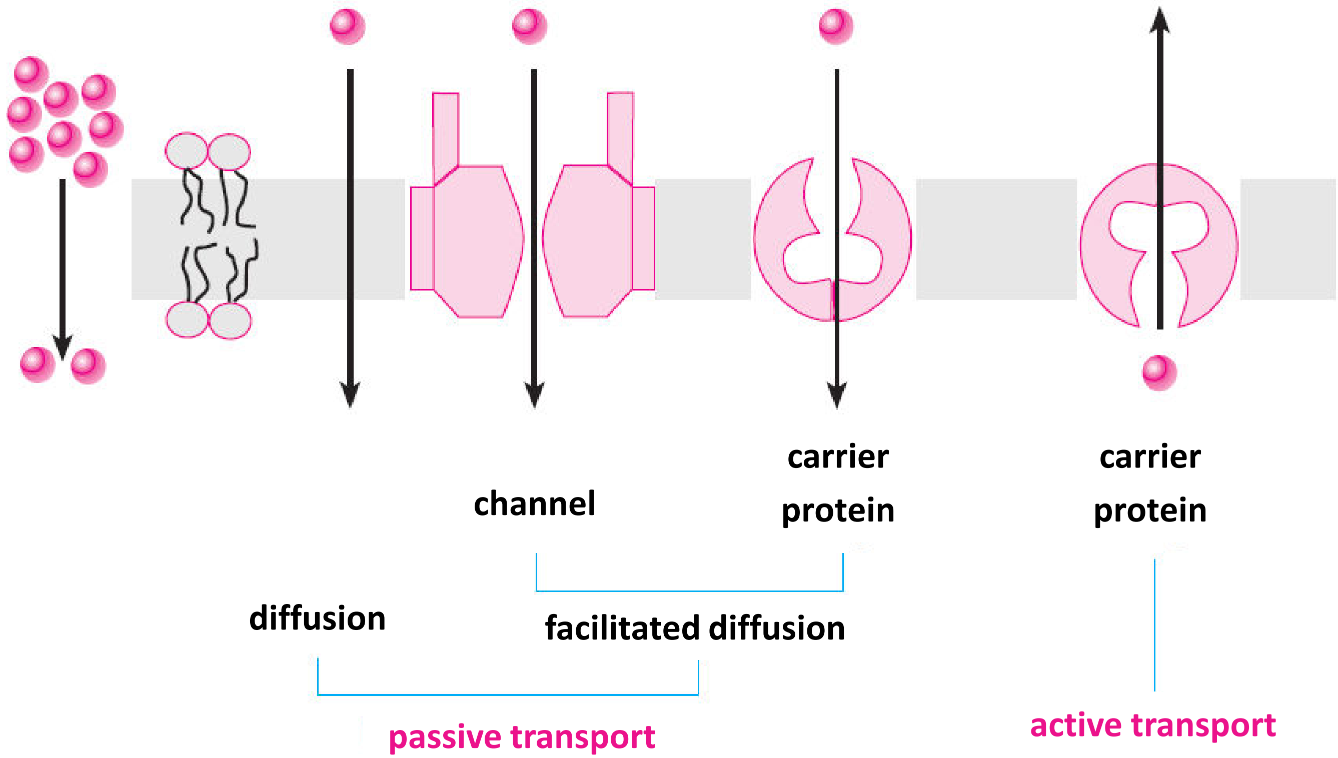
1. April 2020.

Physical properties of the cell membrane



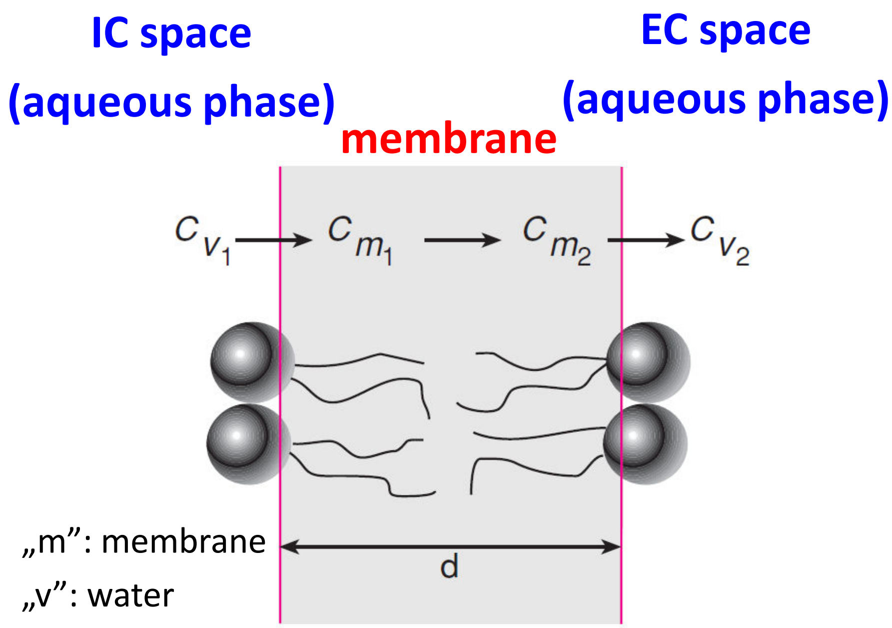
- **Noncovalent, cooperative structure:** phospholipid bilayer, vesicle formation, additional components (e.g. cholesterol, proteins)
- **Thin, layered**: d ~ 5 nm
- **Asymmetric:** the two sides of the membrane are different
- **Permeability:** impermeable to ions, permeable to water
- **Fluidity:** melting temperature (T_m)
- **Lateral diffusion:** lateral movement of lipid-, and protein molecules
- **Flip-flop:** phospholipid translocation between the two layers (low probability)
- **Flexibility, elasicity:** distorsion of erythrocytes in the capillary

Transport across biological membranes



- **Passive diffusion:** „real", classical diffusion (Fick's first law)
- **Facilitated or mediated diffusion:** through biological membranes, through/with protein(like) mediator molecules
- **Active transport:** the particle is transported against a gradient (chemical/electrochem.)

Passive diffusion across the membrane



Fick's first law:

$$J_m = -D \cdot \frac{\Delta c}{\Delta x} = -D_m \frac{c_{m2} - c_{m1}}{d}$$

D_m: diffusion coefficient within the membrane

Permeability constant: p_m, [m/s]

$$p_m = \frac{D_m}{d}$$

Partition coefficient: K

(between the membrane and aqueous phases)

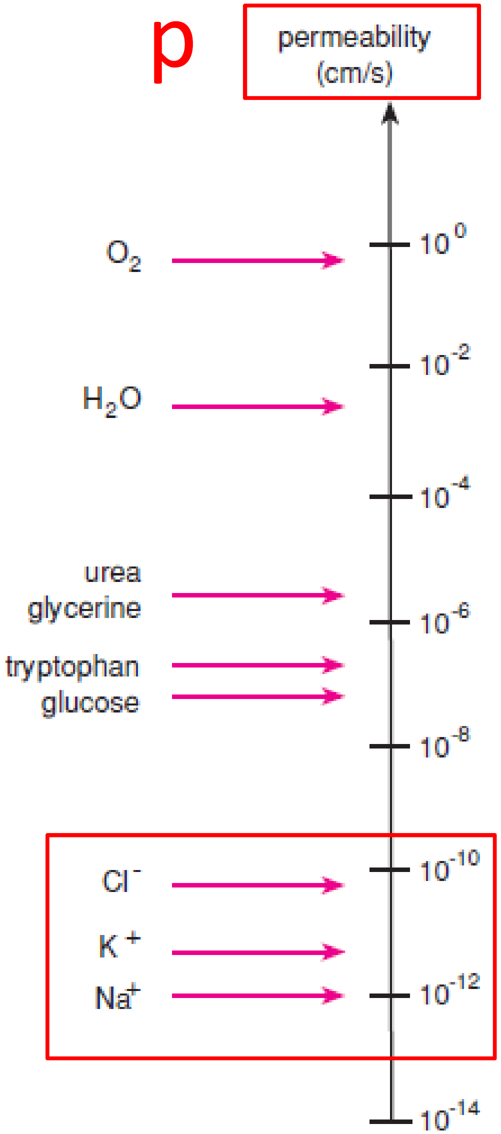
$$\frac{c_{m1}}{c_{v1}} = \frac{c_{m2}}{c_{v2}} = \text{const.} = K$$

$$J_m = -p_m \cdot K(c_{v2} - c_{v1}) = -p(c_{v2} - c_{v1})$$

Aggregated permeability constant: p, [m/s]

$$p = K \cdot p_m$$

Passive diffusion of particles



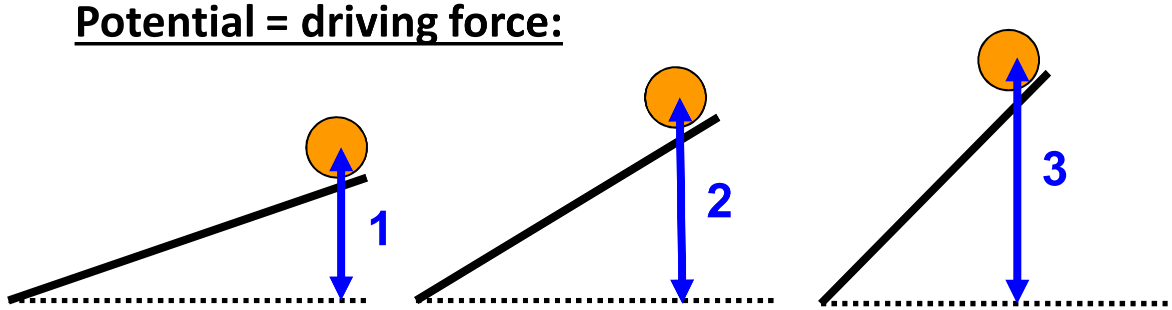
Onsanger equation:

$$J = L \cdot X$$

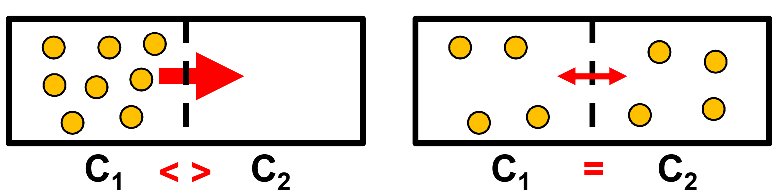
J	L	X
matter flow density	conductivity coefficient	gradient of an intensive quantity

The driving force of transport is the chemical potential gradient.

Potential = driving force:

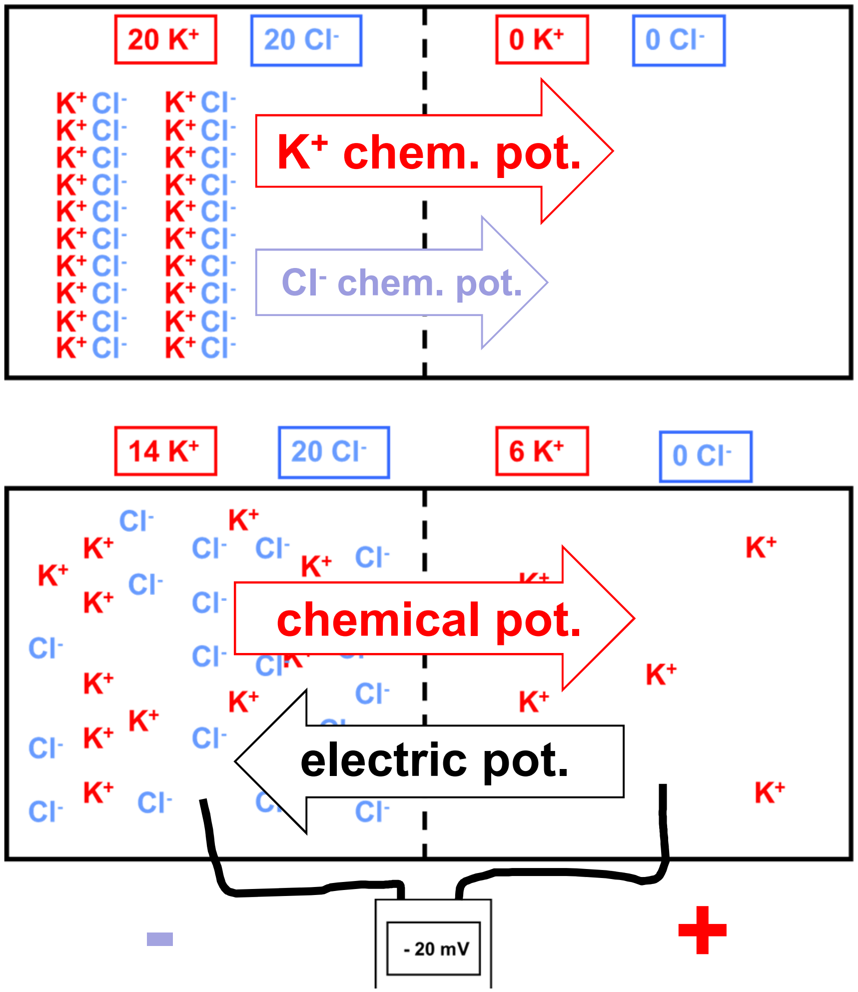


Chemical potential: μ $\mu = \mu_0 + RT \cdot ln(c)$



μ_0: standard chemical potential

Passive diffusion of ions: electrochemical potential



$$J_k = L_k \cdot X_k = -D_k\left(\frac{\Delta c_k}{\Delta x} + c_k \frac{z_k F}{RT}\frac{\Delta \varphi}{\Delta x}\right)$$

Assume that the membrane is only permeable to K^+ ($p_{Cl^-}=0$).

In equilibrium:

- concentration difference
- electric potential difference

exist between the two compartments.

- **the chemical and electric potentials are of the same magnitude but oppositely directed.**

Electrochemical potential: μ_e, [J/mol]

$$\mu_e = \underbrace{\mu}_{\text{chemical}} + \underbrace{zF\varphi}_{\text{electric}}$$

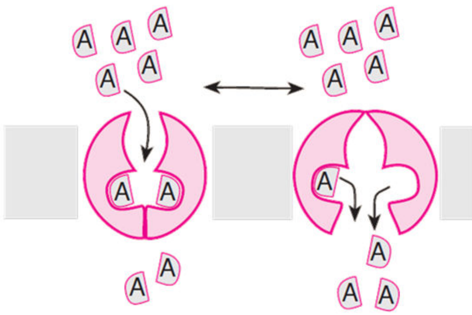
z: charge

F: Faraday constant

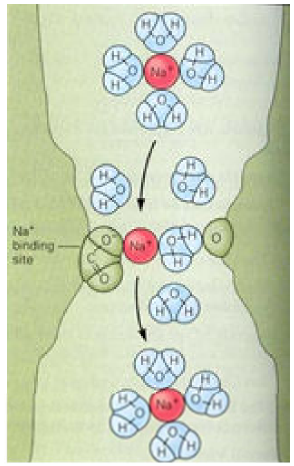
φ: electric potential

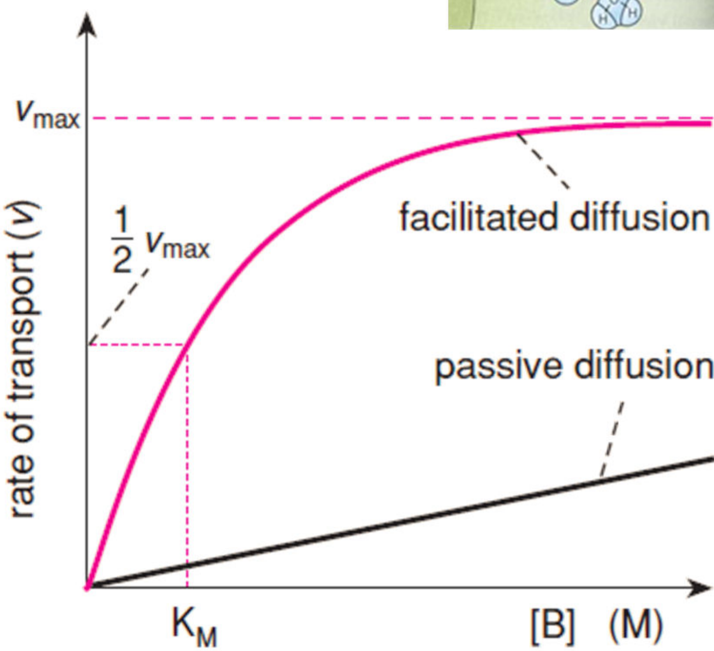
Properties of the facilitated diffusion

carrier



channel



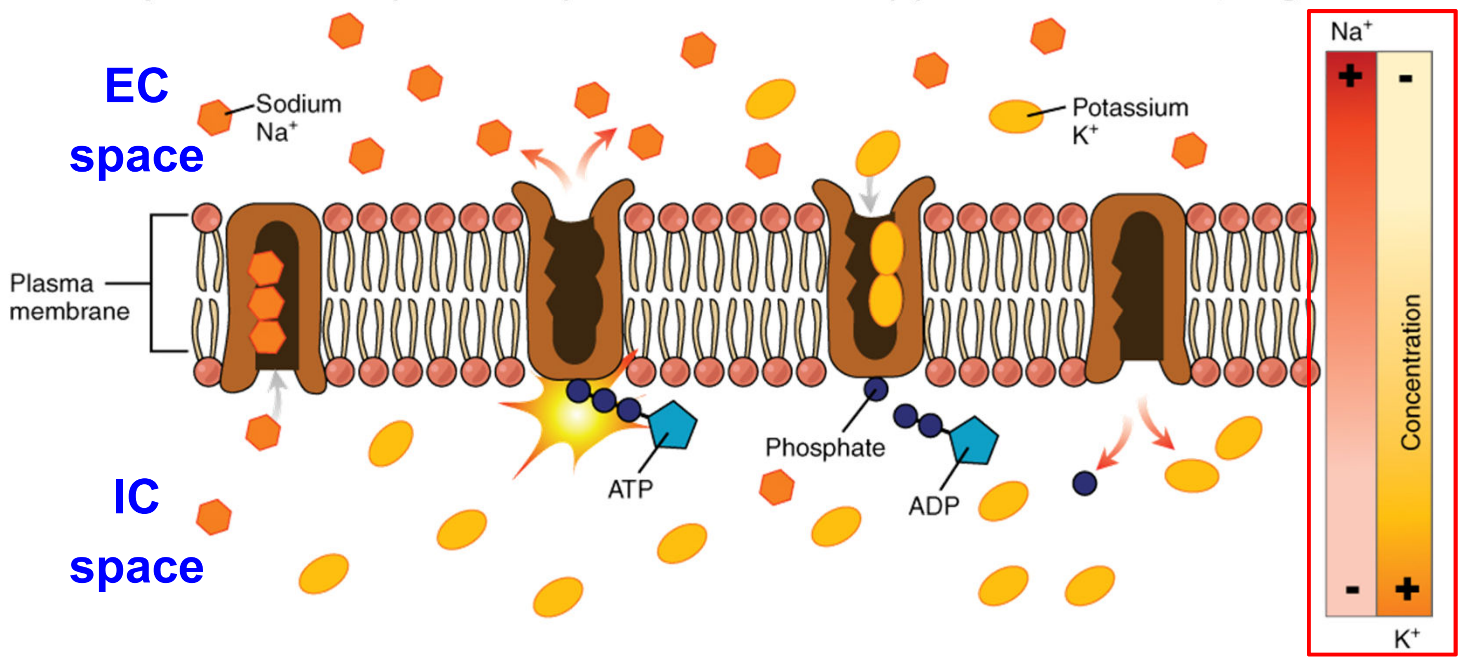


- **Faster, than the passive diffusion** (what is expected based on Fick's first law)
- **Selective**: works only for a given particle or for molecules sharing structural similarity
- **Can be saturated:** is realized through a limited number of mediator molecules (carrier or channel)
- **Can theoretically work in both directions:** the direction is determined by the sign or direction of the (electro)chemical potential gradient of the transported molecule
- **Can be selectively inhibited:** with inhibitors targeting the mediator molecules
- Ionophores: mobile ioncarriers or channel-forming molecules. Application: antibiotics

Active transport

- **Transpost of molecules against their (electro)chemical gradient, energetically:**
  - **ATP-driven**: ATP-ases, they hydrolyze ATP
  - **light-gated** (e.g. channelrhodopsin-2: non-selective cation channel)
  - **coupled transporter:** couples the transport of a substance with sufficient electrochemical gradient to the <u>transport of another molecule</u> against its gradient

- **According to the numbers of the transported molecules:**
  - **uniporter**: translocates only one molecule across the membrane
  - **symporter:** transport the particles in the same direction
  - **antiporter:** transport the particles in the opposite direction, e.g. **Na^+-K^+ ATP-ase:**



ATP!

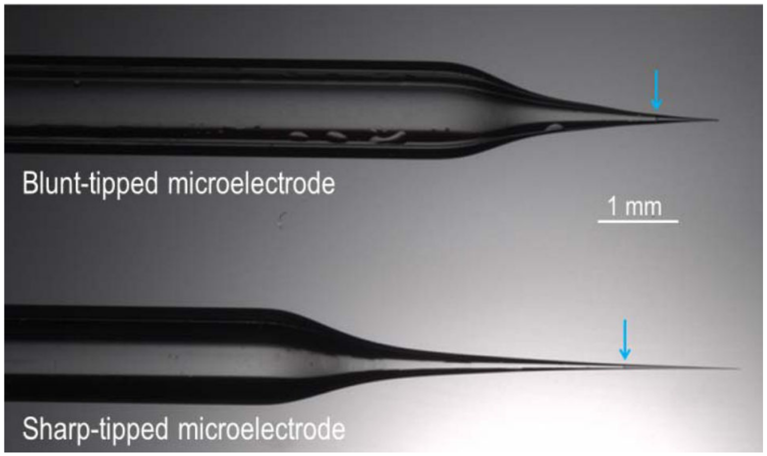
3 Na^+ out

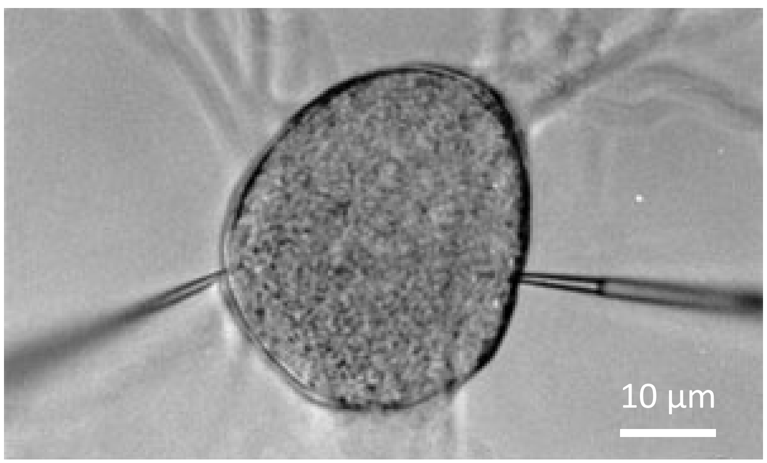
2 K^+ in

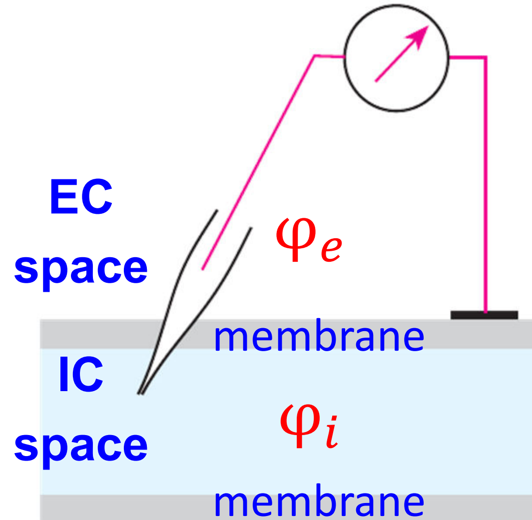
Resting membrane potential

Measurement: with microelectrodes

- active
- reference







Observation: $\Delta\varphi = \varphi_i - \varphi_e < 0$

Cell	Δφ (mV)
squid giant axon	-62
frog muscle	-92
rat muscle	-92

The intracellular space is more negative.

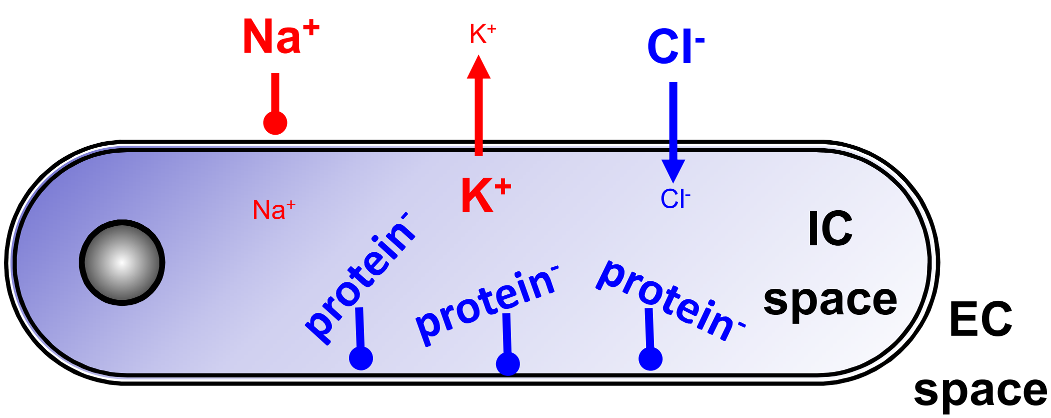
Resting membrane potential

Further observation: different ion concentrations on both sides of the membrane

	intracellular concentration (mmol/l)			extracellular concentration (mmol/l)		
cell	Na^+	K^+	Cl^-	Na^+	K^+	Cl^-
squid giant axon	72	345	61	455	10	540
frog muscle	20	139	3,8	120	2,5	120
rat muscle	12	180	3,8	150	4,5	110

Considering the ion distribution shown in the table above which physical model gives the best approximation of the resting membrane potential?

Model #1: Donnan-model: equilibrium ion distribution, additional prot. anions in the cell



- The membrane is impermeable to certain ions ($p_{prot^-}=0$).
- Electrochemical equilibrium is assumed.

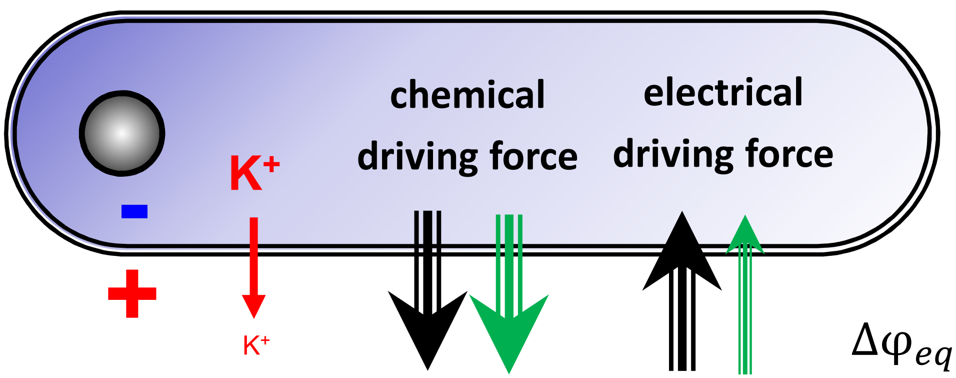
Resting membrane potential

Equilibrium potential: calculated based on the Donnan-model... (book p. 287.)

Nernst equation: $\Delta\varphi = \varphi_2 - \varphi_1 = -\frac{RT}{F}\ln\frac{c_2}{c_1}$

- let's calculate it for the K^+ ion...

	intracellular concentration (mmol/l)			extracellular concentration (mmol/l)		
cell	Na^+	K^+	Cl^-	Na^+	K^+	Cl^-
squid giant axon	72	345	61	455	10	540



$$\Delta\varphi_{eq} = -\frac{RT}{F}\ln\frac{c_i}{c_e}$$

$$\Delta\varphi_{eq} = -\frac{8{,}31*293}{96500}\ln\frac{345}{10} = \text{-0,089 V} = \boxed{\text{-89 mV}}$$

Measured membrane potential: -62 mV

The equlibrium model does not correctly describe the real situation!

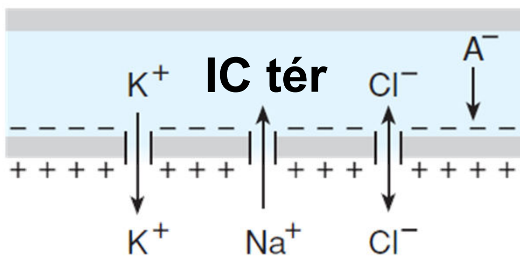
Outward flux of K^+ at -62 mV.

The Goldman-Hodgkin-Katz equation

Cell	$\Delta\varphi_{eq}$ (mV) using the Nernst equation			$\Delta\varphi_m$ (mV)
	Na^+	K^+	Cl^-	
squid giant axon	+46	-89	-55	-62
frog muscle	+45	-101	-87	-92
rat muslce	+64	-93	-85	-92

<u>No equilibrium at rest but the transport processes contiune:</u>

- outward flux of K^+
- inward flux of Na^+
- minor outward flux of Cl^-



- active transport: requires energy (ATP)

<u>Transport model #2:</u> continuous diffusion of <u>different ions</u> with <u>different permeability</u>

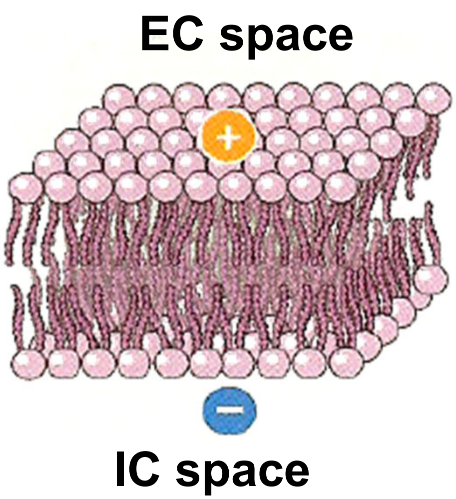
$$\Delta\varphi = \varphi_i - \varphi_e = -\frac{RT}{F} ln \frac{p_{Na}c_{Na}^{i} + p_K c_K^{i} + p_{Cl}c_{Cl}^{e}}{p_{Na}c_{Na}^{e} + p_K c_K^{e} + p_{Cl}c_{Cl}^{i}} = -91 \text{ mV}$$

in frog muslce

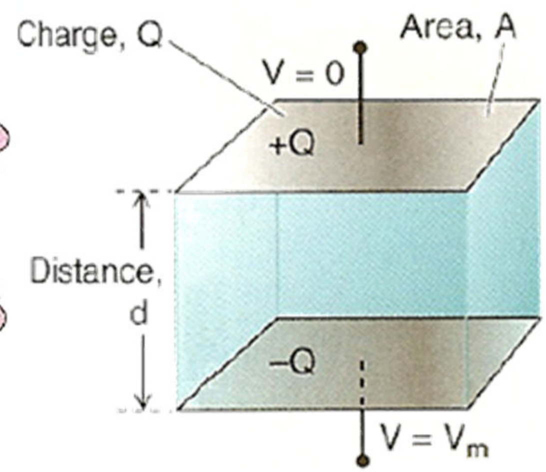
The calculation using the GHK equation is in agreement with the measurements.

The electric model of the cell membrane

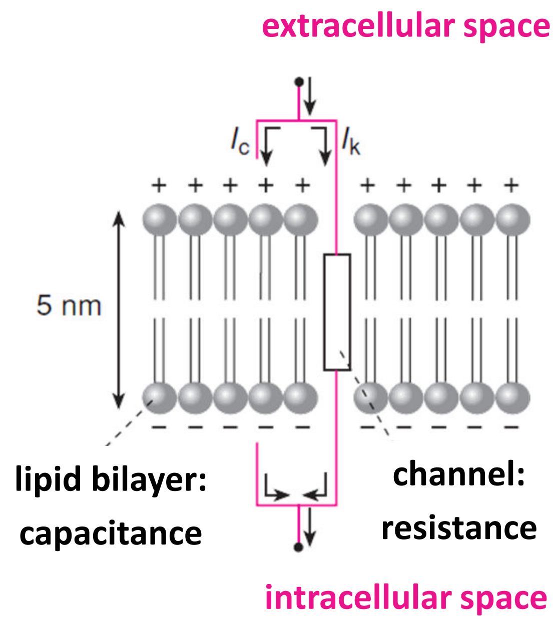
cell membrane



capacitor



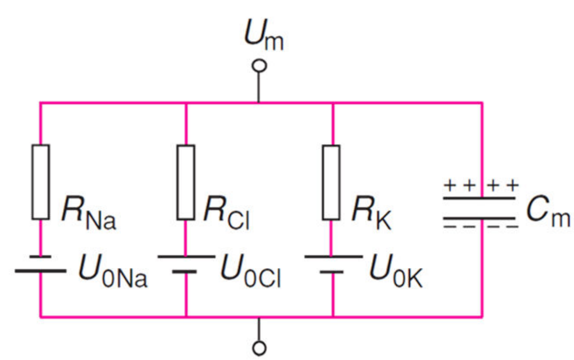
electric model



- different transmembrane resistance in the case of the different ion channels
- electric conductivity: $$G = \frac{1}{R}$$

  proportional to the permeability

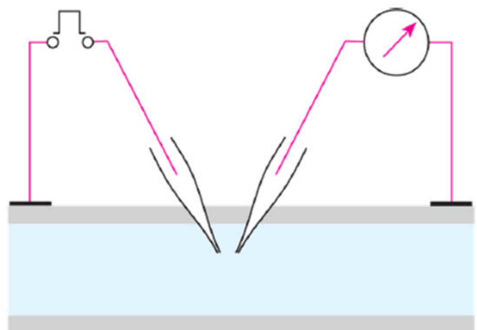
- specific condictivity: $$\sigma = \frac{1}{RA}$$

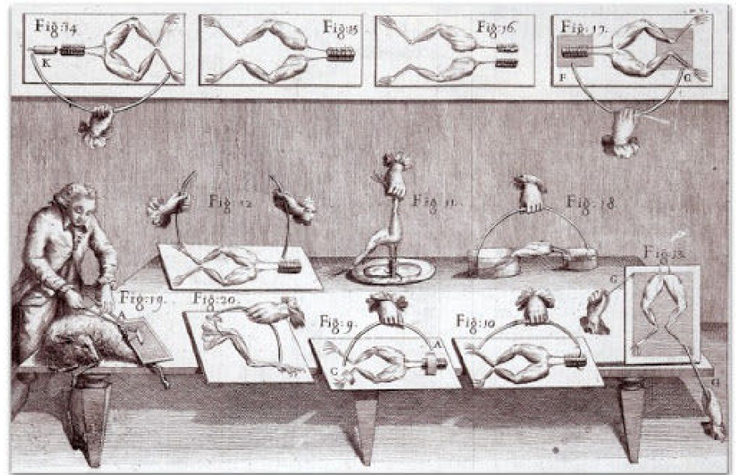


The change of the resting potential

stimulator electrodes

registering electrodes





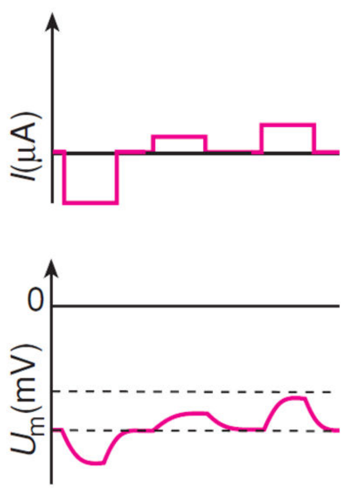
Local (electrotonic) changes of the membrane potential:

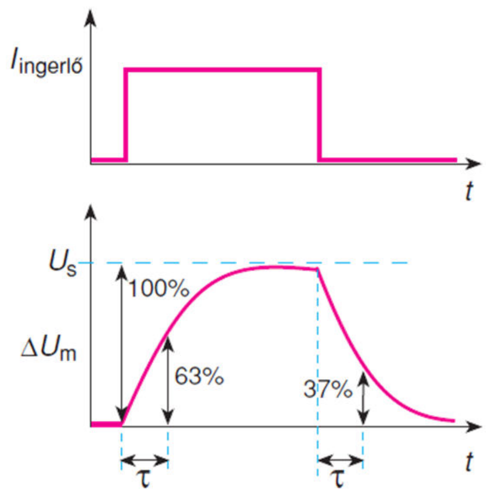
Stimulation

$\mathbf{I_{stim}}$

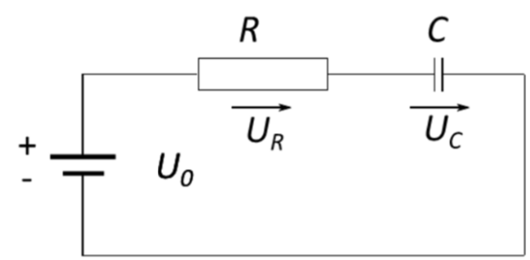
Response

$\mathbf{\Delta\varphi_m}$





Model: RC-circuit

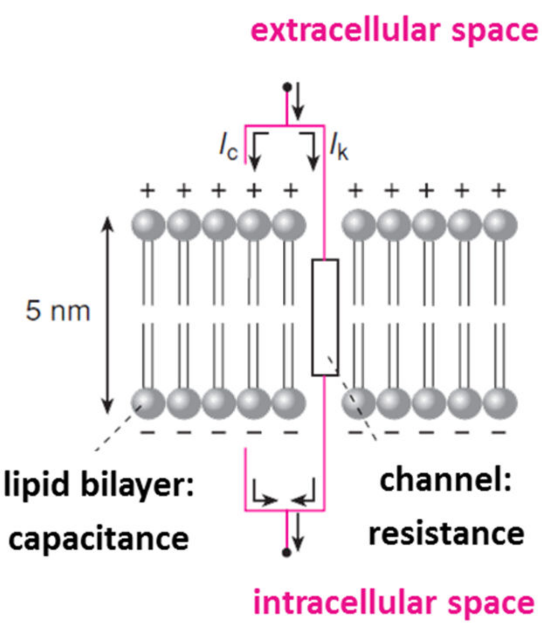


Time-constant: τ, [s]

$\tau = R_m \cdot C_m$

The amplitude of the response is proportional to the stimulating current, but shows a characteristic delay.

Electric properties of the membrane



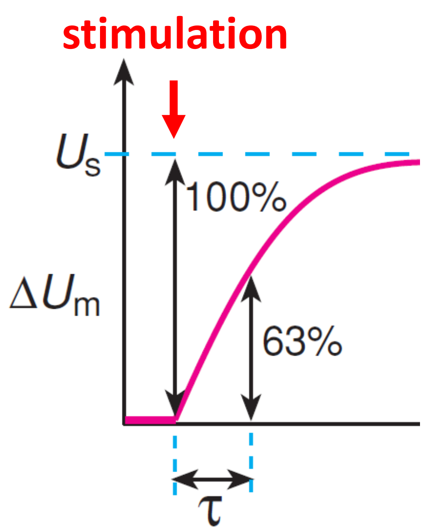
Currents across the membrane:

- stimulating: I_{stim} negative with the influx of + charges
- conductive: I_k
- capacitive: I_C

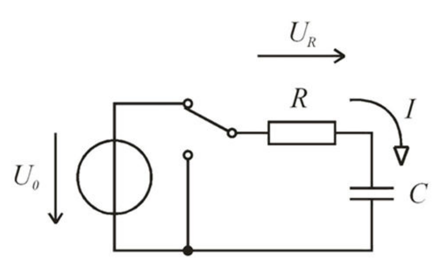
Based on the transport model for the resting state:

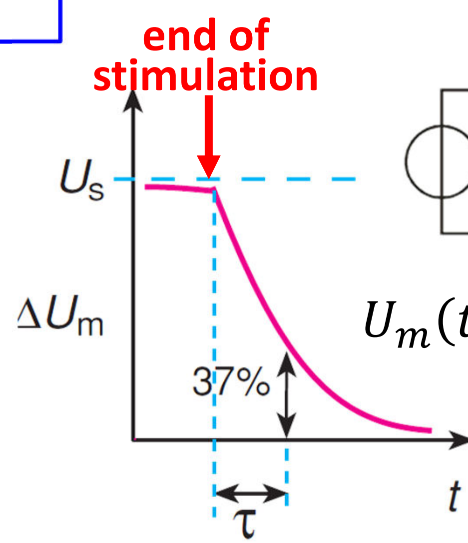
$$I_C + I_k - I_{stim} = 0$$

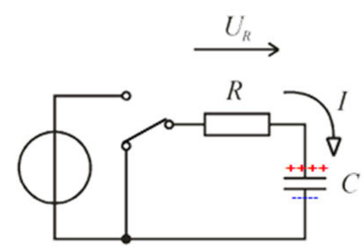
$$\underbrace{C_m \frac{\Delta U_m}{\Delta t}}_{I_C} + \underbrace{\frac{U_m - U_0}{R_m}}_{I_k} - I_{stim} = 0$$



$$U_m(t) = U_t\left(1 - e^{-\frac{t}{\tau}}\right)$$



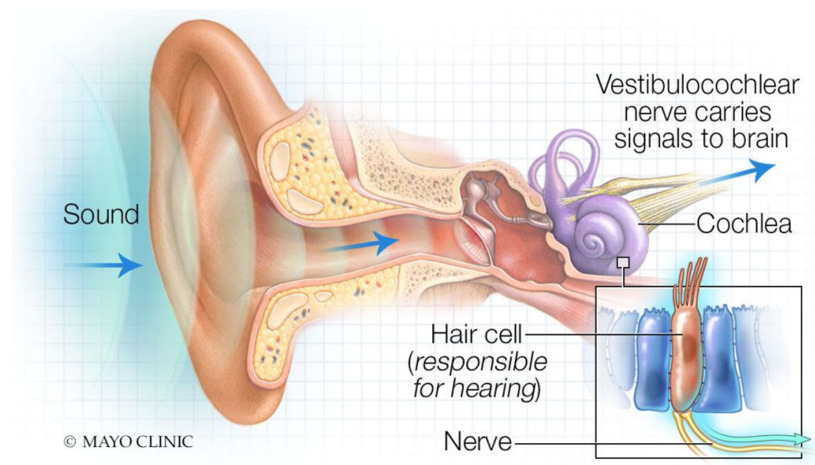


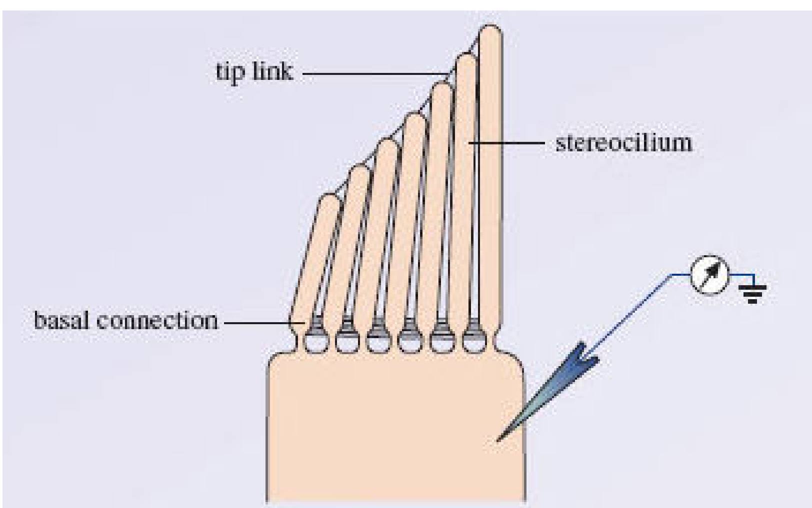


$$U_m(t) = U_t e^{-\frac{t}{\tau}}$$

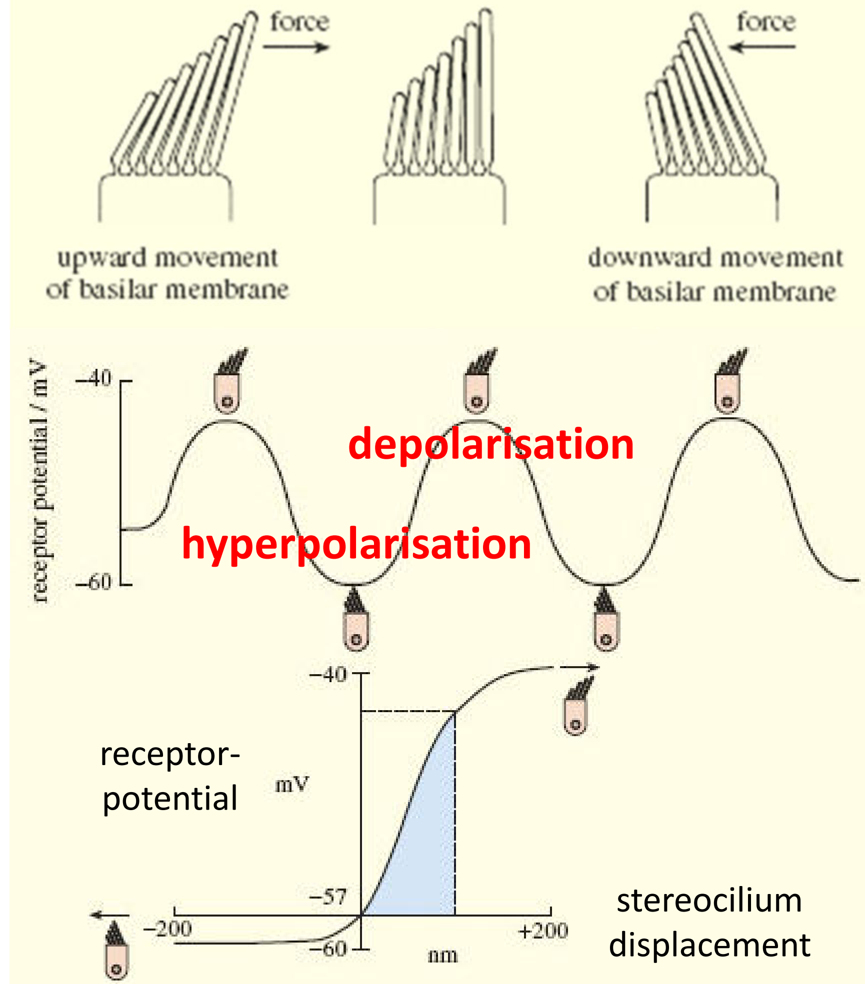
Example: receptor potential

<u>**Example for the local change of the membrane potential:**</u> hair cells as mechanoreceptors



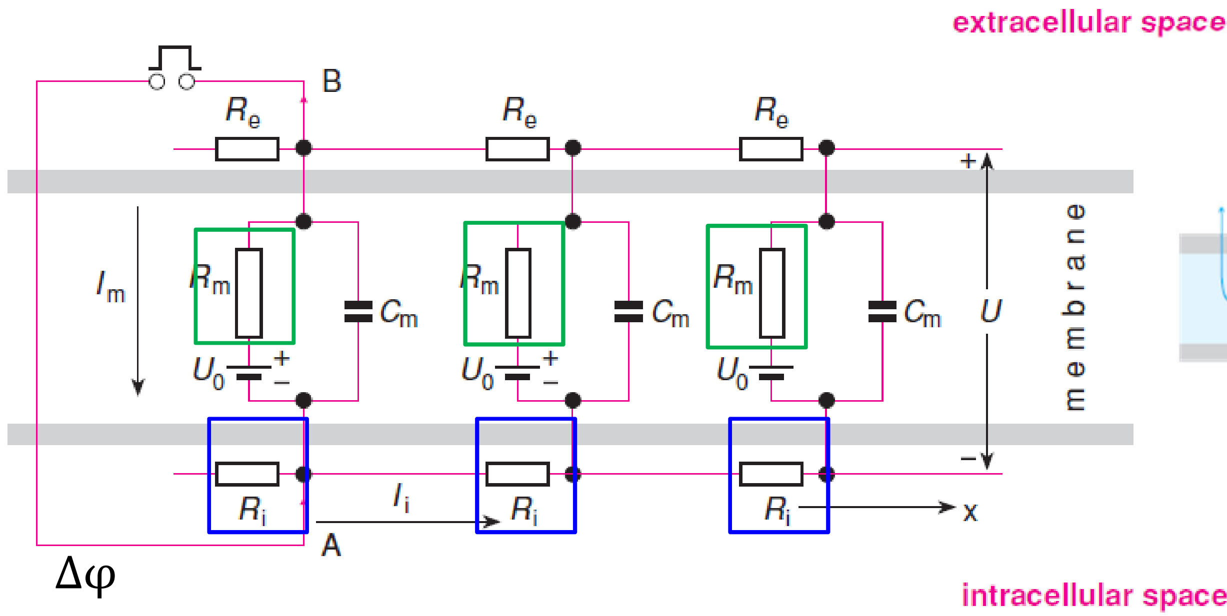


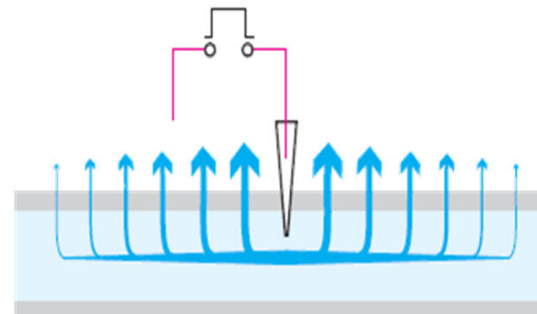
mechanosensitive K^+-channel: K^+ in

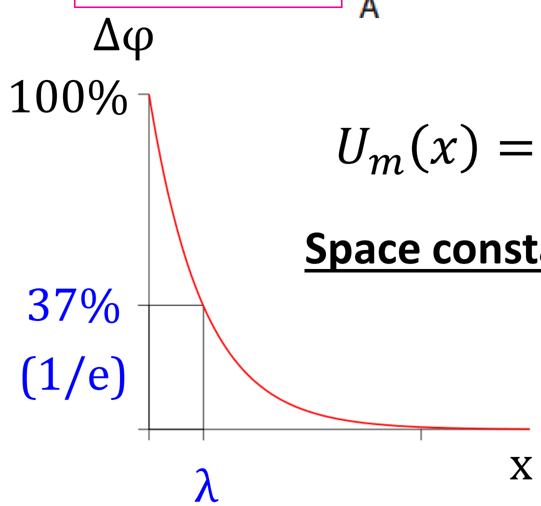


The propagation of a potential change

Model of a larger membrane section:







$$U_m(x) = U_0 e^{-\frac{x}{\lambda}}$$

Space constant: λ, [cm] $\lambda \sim \sqrt{\frac{R_m}{R_i}}$

R_m: transmembrane resistance

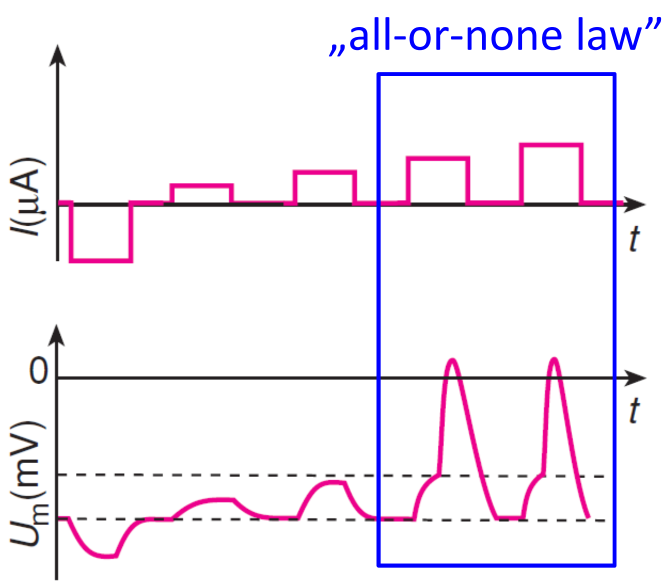
R_i: intracellular resistance

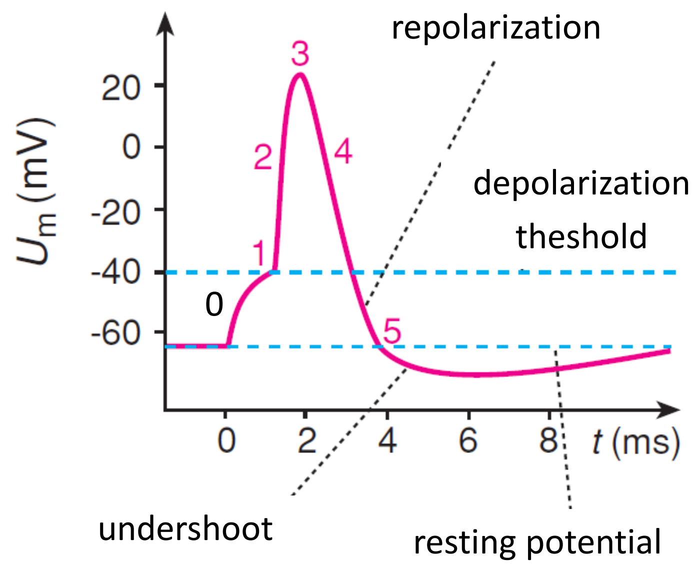
When R_m ↑ or R_i ↓: improved potential propagation.

Example: myelin sheath.

Action potential

For stimuli above threshold: generalized change of the membrane potential





t ~ 1-5 ms

(muscle and neuron)

U ~ 100 mV

0: local change of membrane potential

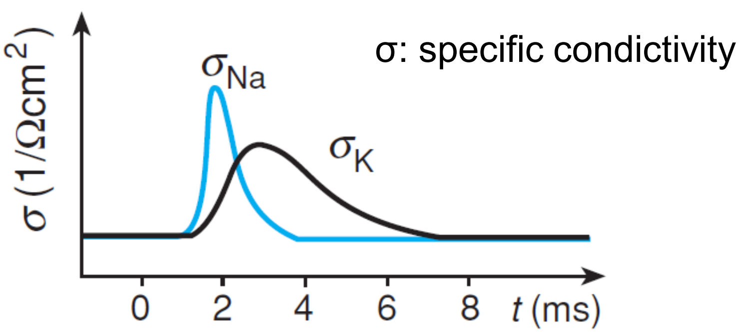
1: **volt. gated Na^+** ch. open (Na^+: **in**)

2: **volt. gated K^+** ch. open (K^+: **out**)

3: **Na^+** ch. **inactivation** (partial)

4: Na^+ channel closure

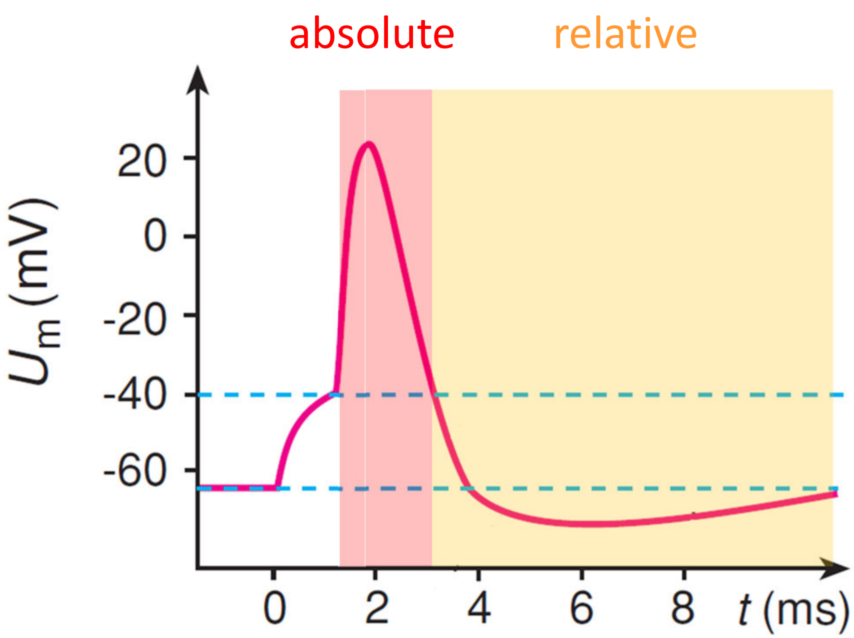
5: **K^+** channel **closure** (delayed)



Properties of the action potential

Unaltered ion concentration: the transported ions diffuse away far from the membrane. During the AP only the permeability changes (GHK).

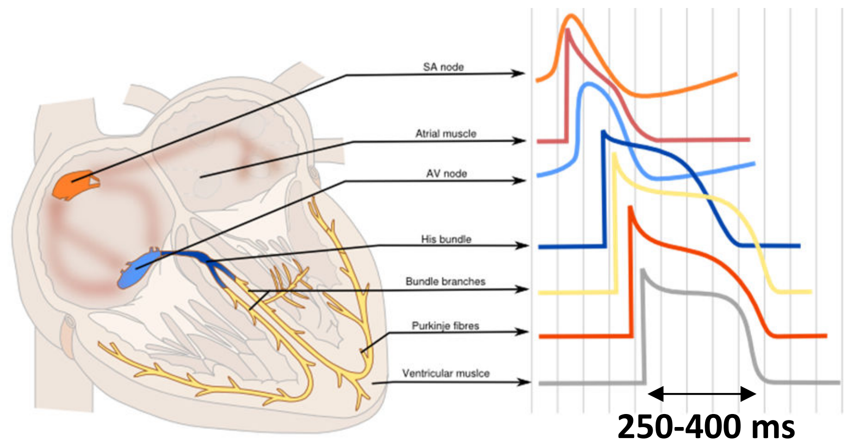
Refractory period: the cell is not excitable

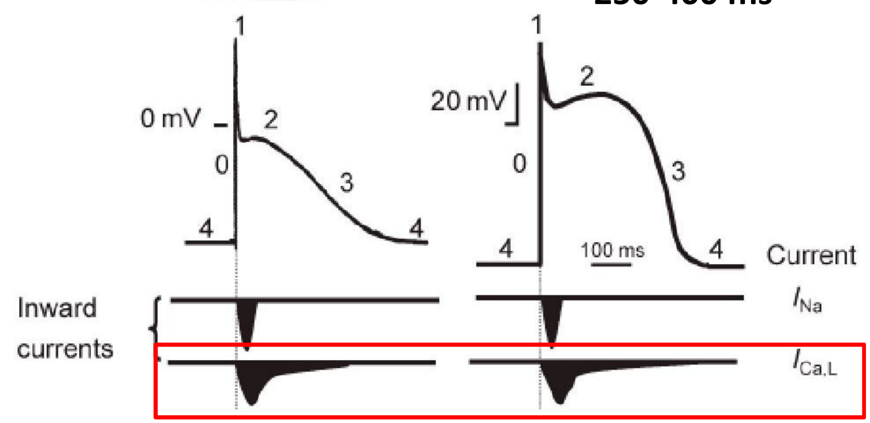


- **absolute:** voltage-gated Na^+ channels are inactivated
- **relative:** AP with supra-threshold stimulus

prevents the backpropagation

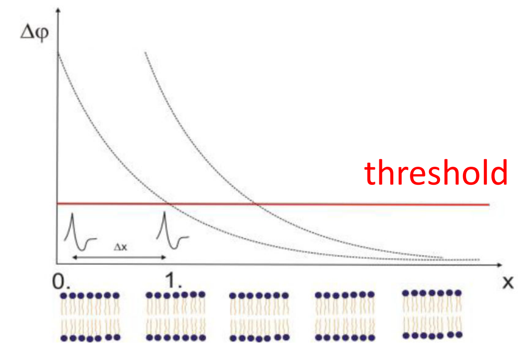
Special AP: e.g. ventricular cardiomyocytes





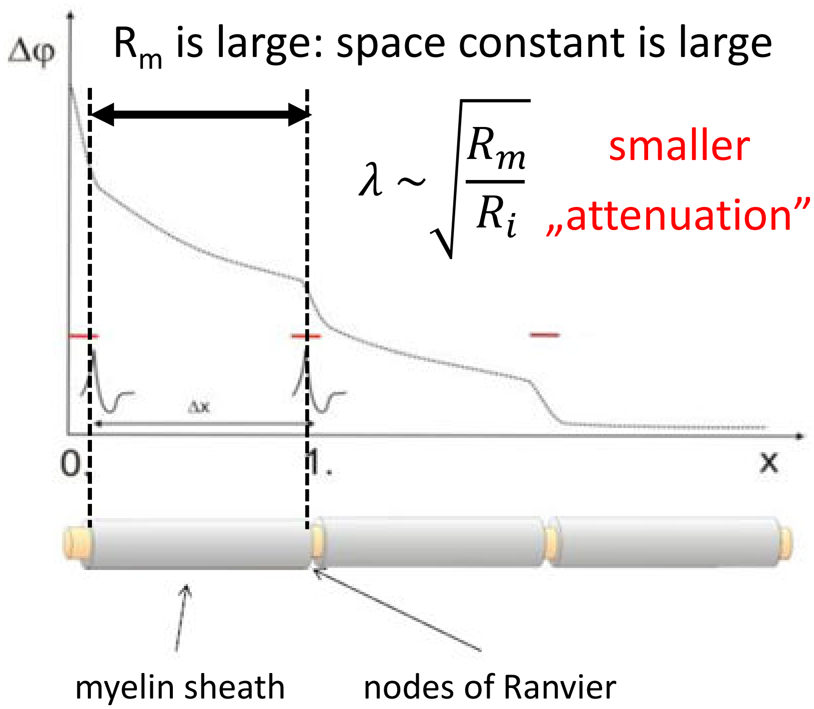
- **voltage-gated Ca^{2+} channels**

The propagation of the action potential



Properties:

- shape is independent from stimulus
- propagates far without attenuation
- much faster than hormonal response



fiber	diameter (μm)	Speed (m/s)
α	15	70-120
β	8	30-70
γ	5	15-30
δ	<3	12-30
No sheath	<1	0.5-2

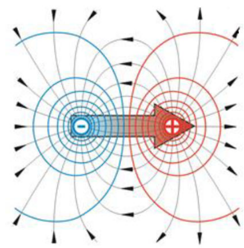
Medical application of bioelectric phenomena

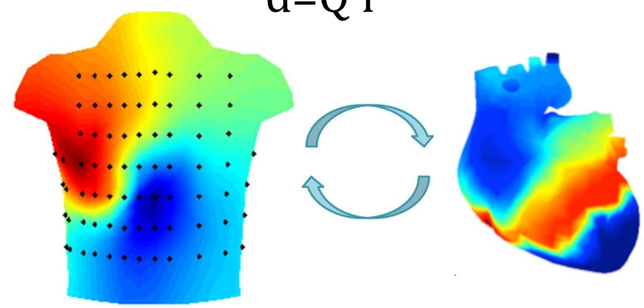
origin: dipole moment

$d = Q \cdot l$

Electric signals on the bidy surface (diagnostics):

- Electrocardiography (EKG)
- Electroencephalography (EEG)
- Electromyography (EMG)
- Electrooculography (EOG)
- Electroretinography (ERG)

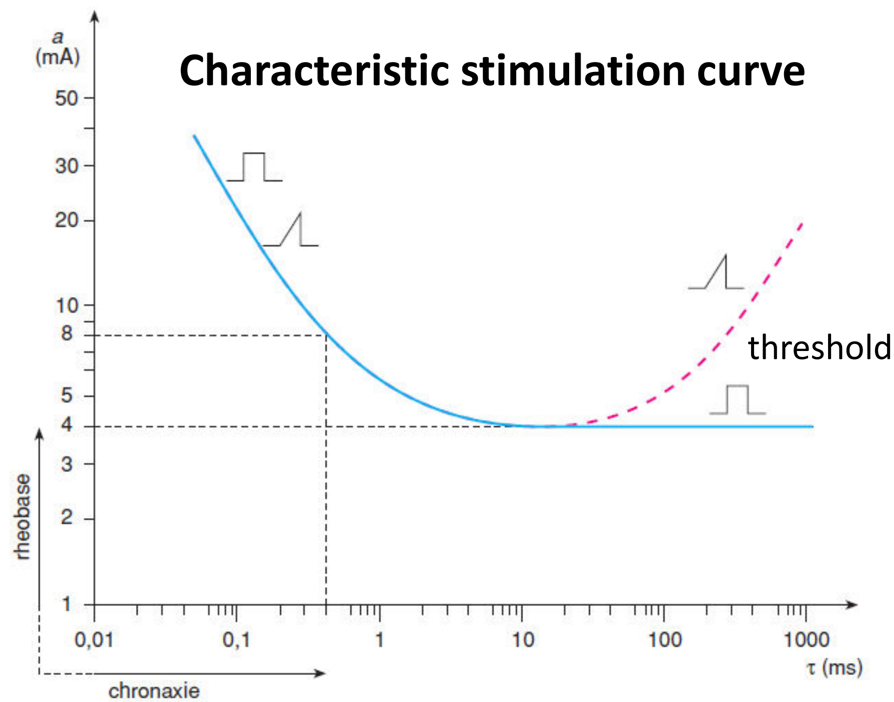




Elektromos ingerlés (terápia):

- Galvanic treatment (DC)
- Iontophoresis (DC)
- HF-thermotherapy (AC)
- Electric stimulus therapy (pulse)
- Defibrillator (pulse)
- Pacemaker (pulse)

- **rheobase:** minimal electric current that elicit stimulation
- **chronaxie:** time to 2x rheobase

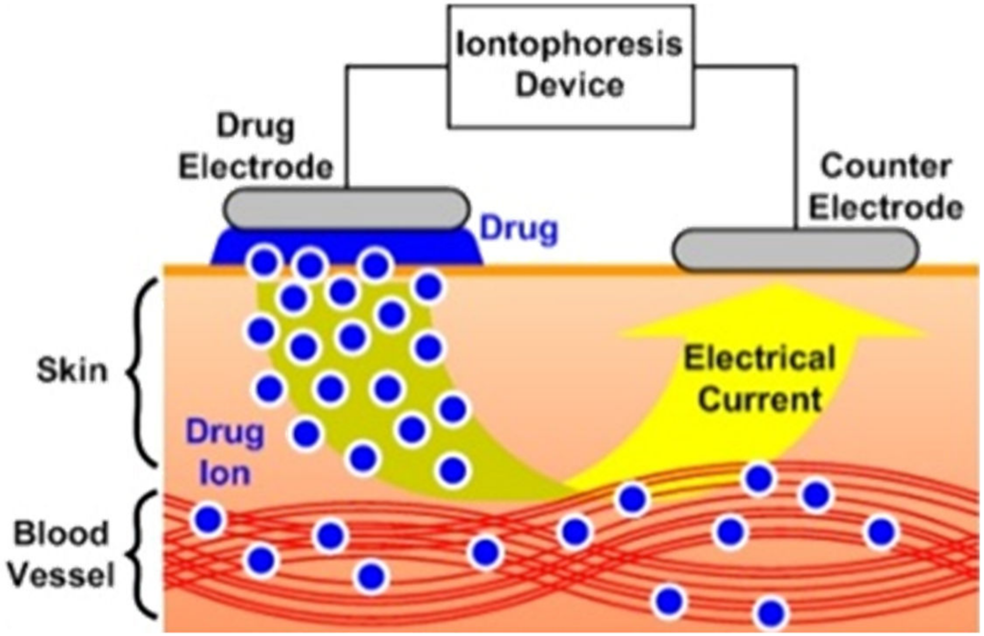


The Application of Direct Current

<u>Galvanic treatment</u>

- I ~ mA, t ~ 10 min
- analgesia
- improving circulation
- improving metabolism

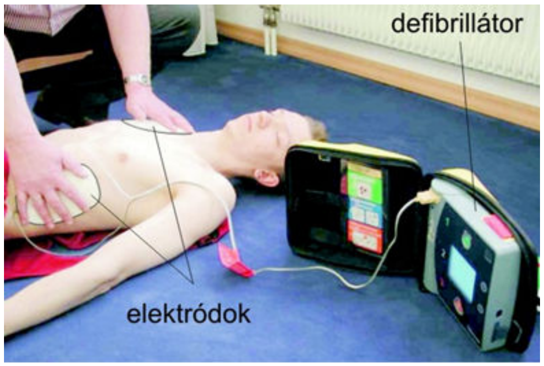
<u>Iontophoresis</u>



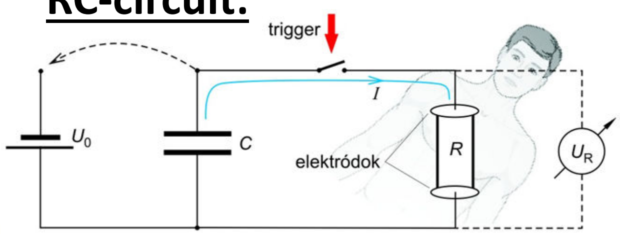
- I ~ mA, t ~ 10 min
- a charged substance (e.g. medication) propelled rapidly through the dermis
- the polarity of the drug electrode should match the charge of the substance

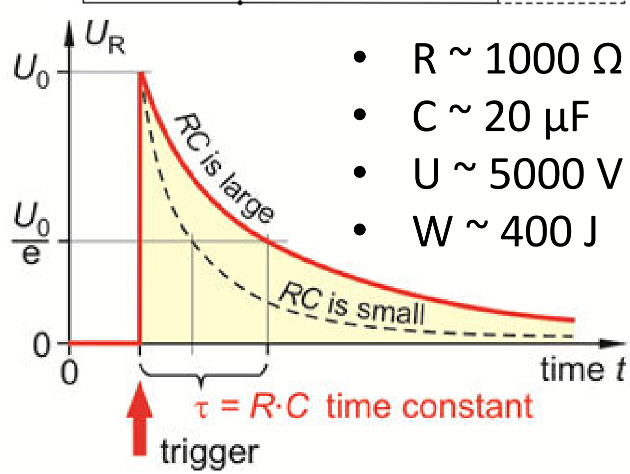
Therapy with electric stimuli

Defibrillator



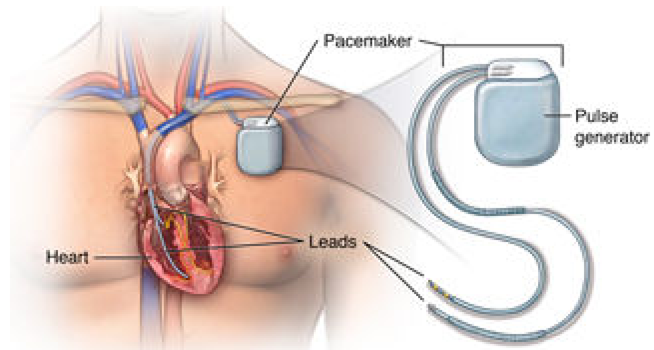
RC-circuit:

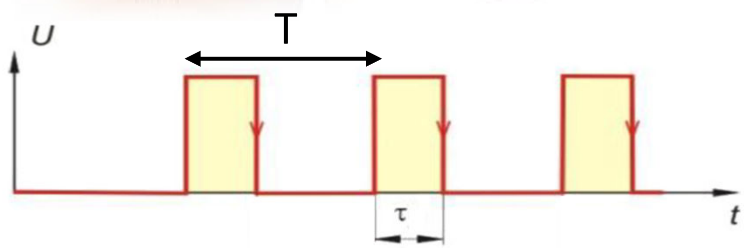




- R ~ 1000 Ω
- C ~ 20 μF
- U ~ 5000 V
- W ~ 400 J

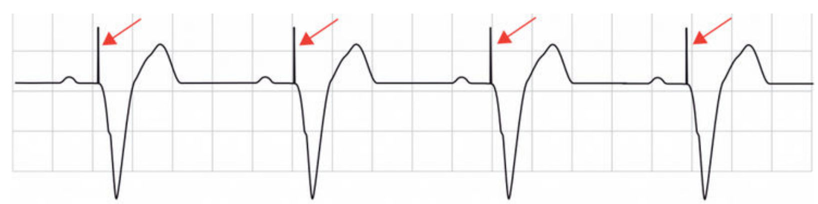
Pacemaker





- τ ~ ms
- T ~ s

- U ~ 1 V
- R ~ 200 Ω
- I ~ 5 mA



Thank you for your attention!

Electrophysiology:

Phases of the Cardiac Cycle