

# Ligand-protein interactions and binding thermodynamics

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## Subject

- Focus on ligand-protein binding
  - Qualitative and quantitative characterisation
  - Thermodynamics (and kinetics)
  - How to interact/influence - drug design
  - Computational support to drug discovery

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## Outline

- Basic relationships
  - Measurements and computations
  - Analysis of ligand-protein binding
  - Role of water
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- Computations
    - Molecular dynamics (MD)
      - Tool for quantitative description
    - MD based applications to characterize ligand-protein binding
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- Very fast estimation of ligand-protein interactions
    - Docking-scoring

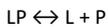
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## Ligand-protein binding

- Signal transduction
    - G-protein coupled receptors (GPCRs)
  - Enzymatic catalysis
    - Cytochrome P450
  - Transcription
    - Nuclear receptors...
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- Endogenous and exogenous (e.g. drugs) ligands

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## Few basic relationships



$$K_d = \frac{[L][P]}{[LP]}; pK_d = -\log(K_d)$$

$$\Delta G_{\text{bind}} = RT \ln(K_d/C_{\text{ref}})$$

$$\Delta G = \Delta H - T\Delta S \quad (\text{Gibbs) typical experimental conditions (NPT)}$$

$$\Delta F = \Delta U - T\Delta S \quad (\text{Helmholtz) calculations for solutions, often used (NVT, canonical)}$$

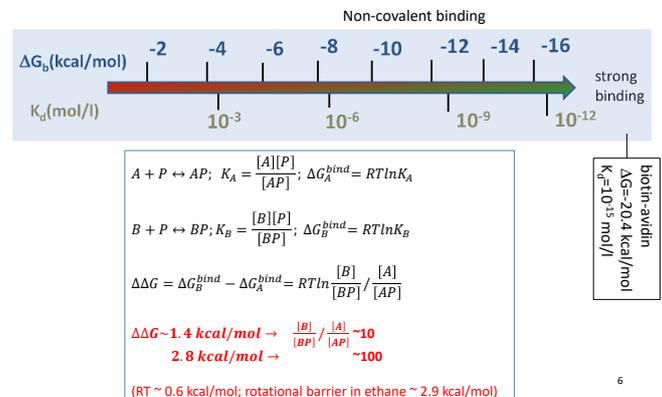
$$F = -k_B T \ln Z,$$

$$Z = \sum_i e^{-\frac{E_i}{k_B T}} - \text{partition function} \quad (\sim \int e^{-\frac{E(r,p)}{k_B T}} dr dp)$$

can be calculated for simple systems only

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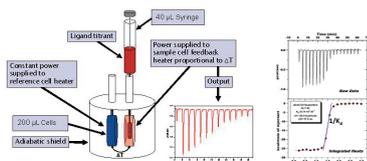
## Free energy – Equilibrium constant



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## Measuring binding thermodynamics

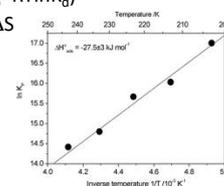
- Isothermal titration calorimetry
  - $n, K_d, \Delta H \rightarrow \Delta G, \Delta S$
  - limits:
    - solutions
    - protein quantity (10-100  $\mu\text{g}$ )
    - throughput



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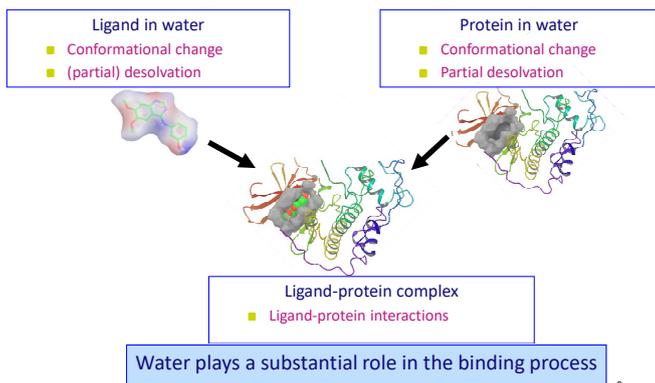
## Measuring binding thermodynamics

- Van't Hoff analysis
  - $\ln K_d = \frac{\Delta H_b}{RT} - \frac{\Delta S_b}{R}$  ( $\Delta G_b = \Delta H_b - T\Delta S_b = RT \ln K_d$ )
  - Measure  $K_d$  at various T  $\rightarrow \Delta H$  és  $\Delta S$
  - Experimental techniques
    - Radioligand displacement
    - Mass spectrometry
    - Chromatography
    - Surface plasmon resonance (SPR)
  - limits
    - $\Delta H$  depends on T
    - extrapolation ( $\Delta S: 1/T=0$ )



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## Ligand-protein binding “steps”



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## Qualitative binding thermodynamics

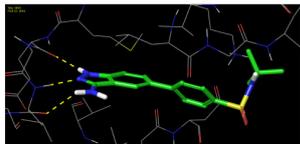
- desolvation (ligand+protein)
  - beneficial  $\Delta S$  (change in water structure)
  - disadvantageous  $\Delta H$
- Conformational change (ligand+protein)
  - disadvantageous  $\Delta H$  (optimal before binding)
- Ligand-protein interactions
  - beneficial  $\Delta H$  (polar and van der Waals interactions)
  - disadvantageous  $\Delta S$  (restricted motion)

$\Delta G$  is a sum of several terms with positive and negative signs

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## Qualitative binding thermodynamics

- $\Delta G, \Delta H, \Delta S$  can be assigned to steps from one state to another – state functions
- Assigning  $\Delta G, \Delta H, \Delta S$  to structural elements is problematic
  - Limited additivity
    - $\Delta H$  additivity – good approximation
    - $\Delta S$  additivity – bad approximation

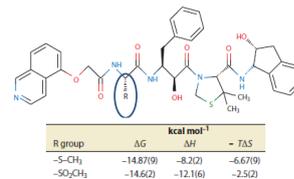
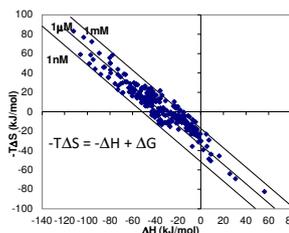


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## Enthalpy-entropy compensation

Small structural changes of a ligand-protein complex results in significant  $\Delta\Delta H$  és  $\Delta(T\Delta S)$  changes of opposite sign and a small change in  $\Delta\Delta G$

- The compensation is observed for a wide range of phenomena
- Both in water and in apolar solvents



$\Delta G$  changes are limited (within ~35kJ/mol)

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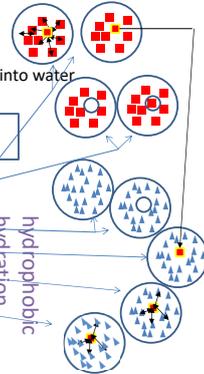
## Hydrophobic effect

### Hydrophobic effect:

Bringing an apolar substance from its apolar solvent into water (hydrocarbon → water)

analogy: desolvation upon ligand-protein binding (inverse)  
apolar moieties - solvent → self-interactions

- Breaking apolar contacts and removing molecule
- Filling empty space in the apolar medium
- Hole formation in water
- Inserting the apolar substance
- Formation of solute-solvent interactions
- Reorganization of water structure



$\Delta G$  positive

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## Hydrophobic effect

### $\Delta G$ increase

– 20°C

- $\Delta H$  (advantageous) and  $T\Delta S$  (disadvantageous) decrease;  $T\Delta S$  change dominates

– higher T

- small change in  $\Delta G$
- $\Delta H$  increases and becomes dominant
  - disadvantageous for enthalpy
  - advantageous for free energy
  - interactions are sacrificed for increased disorder

– Explanation: focuses on hydrophobic hydration

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## Hydrophobic hydration

### Key factors in entropy decrease: Water structure perturbed

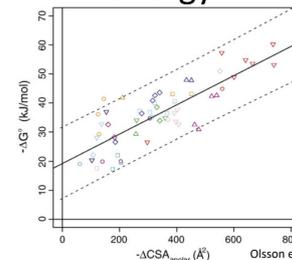
- Hole formation - small size of water molecules
- Water H-bonds near to the apolar solute
  - Stronger and more H-bonds – iceberg model
  - Stronger, but less H-bonds – „two-state“ model

### How do these factors contribute to $\Delta H$ and $T\Delta S$ changes?

No general quantitative model available!

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## Apolar surface and binding free energy



- Ligand-protein binding free energy correlates with apolar surface buried in the binding ( $R^2=0.65$ ).
- Shape fitting and polar/apolar feature mapping give significant contribution to binding
  - Directional interactions do not contribute importantly to the above correlation

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## Affinity and molecular size

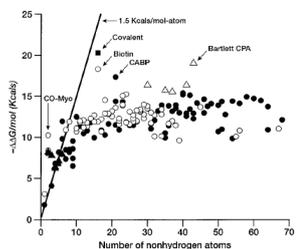


FIG. 1. Free energy of binding (in kcal/mol) for ligands and enzyme inhibitors plotted as a function of the number of nonhydrogen atoms in the ligand. See Table 1. A line with slope of 1.5 kcal/mol and an intercept of 0 is included as a visual aid to analysis.  $\Delta$ , Metal ions or metalloenzymes;  $\square$ , small anions;  $\circ$ , natural ligands;  $\diamond$ , enzyme inhibitors.

PNAS 1999, 96, 9997

Available binding affinity is

- limited
- limit does not increase with size above ~25 nonhydrogen atoms

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## Summary

- Binding thermodynamics – characteristic to ligand-protein interactions
- Key elements of binding: polar interactions and apolar desolvation
- Related phenomena: hydrophobic effect, enthalpy-entropy compensation
- Ligand size affects maximal available binding free energy
  - $\Delta G_{\max}$  – available binding free energy increase fast with ligand size for small ligands and is insensitive to size for larger ligands

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