

Modelling of ligand-protein binding

II. Approximate methods for estimating thermodynamic quantities

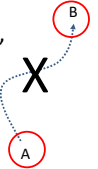
Ferenczy György

Semmelweis University
Department of Biophysics and Radiation Biology

ferenczy.gyorgy@med.semmelweis-univ.hu

Outline

- Calculation of binding free energy by „endpoint” methods
- $\Delta G = G_b - G_A$
 - $G/\Delta G$ cannot be accurately calculated
 - Approximate methods:
 - MM-PBSA (Molecular Mechanics Poisson – Boltzmann Surface Area) – not discussed
 - Docking and scoring



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Docking and scoring

Scoring function

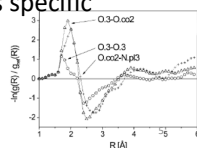
- Crude estimation of ligand-protein binding free energy
- Free energy vs. scoring
- Very fast – (several) ligand(s)/second
- Typically a single configuration is considered
- Accompanied by docking
 - Generating the structure of complexes using minimal preliminary information

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Knowledge-based scoring function

- Derived from the statistical analysis of experimental structural data
 - $E_i = -kT \ln(p_i)$ – energy \sim observed frequency
- Protein Data Bank: over 180000 structures in November 2020
- Binding affinity data not required
- Long-range sampling – solvent effect included
- Short-range sampling – emphasizes specific interactions
- Incomplete repulsion



Docking - scoring

- Generating and ranking ligand-protein complex structures
 - Single ligand-protein pair
 - finding binding mode
 - Multiple ligands and a single protein
 - Virtual screening
 - Binding mode identification
 - Ranking ligands by docking score
- Without preliminary structural information (in principle)
- Application in pharmaceutical research – see later

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Approximations of docking-scoring

Selected approximations:

- Protein is rigid or has limited flexibility
- Protonation state
- Interaction with and structure of water
- Entropy
- Temperature
- ...

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Protein flexibility – docking-scoring

- Role of protein flexibility in ligand binding
 - Selection of protein conformation advantageous for ligand binding
 - Population shift
 - Induced fit
 - Binding to a protein conformation not available for the free protein
 - No strict distinction between the above two mechanisms

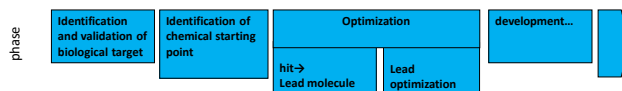
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Protein flexibility – docking-scoring

- Docking with taking into account protein flexibility
 - Using multiple static protein structures
 - Experimental structure – complexes with various ligands, NMR
 - Structures generated by computation (MD, MC)
 - Increased computational requirements
 - „Soft” protein structure
 - Single averaged structure derived from several structures and containing damped interactions
 - Unable to describe large movements
 - Increased binding pocket
 - Mutually exclusive binding sites appear simultaneously
 - Protein conformations generated upon binding (eg. MD)

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Application of docking-scoring

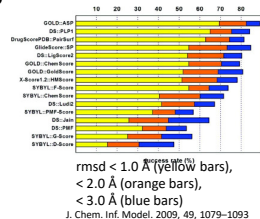


- Virtual screening
 - Identification of chemical starting point
- Docking – Binding mode identification
 - hit to lead

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Docking

- Protein structure
 - X-ray crystallography
 - homology model
- Ligand structure
 - Model
- Complex structure
 - Fitting the ligand into the protein binding pocket - docking
 - Ranking of binding modes using scoring functions
 - Limited protein flexibility
 - Efficient exploration of ligand conformational space
- RMSD of docked ligand < 2 Å – 70-80% in favourable cases



Success rate (%)

rmsd < 1.0 Å (yellow bars),
< 2.0 Å (orange bars),
< 3.0 Å (blue bars)

J. Chem. Inf. Model. 2009, 49, 1079–1093

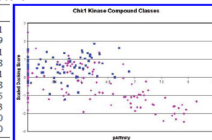
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Ranking

- Docking compounds into a protein and ranking the complexes (ligands) by scoring functions
- Studying similar compounds – lead optimization
- ***weak correlation between score and experimental affinity***

Best Correlation Coefficient r between the
-log Affinity (pAffinity) and Docking Score

program	Chk1	FX
Dock4	-0.33	-0.3
DockIt	-0.49	-0.1
FlexX	-0.57	-0.3
Flot+	-0.44	-0.3
Fred	-0.14	0.0
Glide	-0.47	-0.0
Gold	-0.42	-0.0
LigandFit	-0.45	-0.1
MOEDock	-0.29	0.0
MVP	-0.26	0.1



J. Med. Chem. 2006, 49, 5912

Correlation Between the Scores and Experimental Binding Affinities^a

model	Experimental mean	Experimental std	Simulated mean	Simulated std
code 1	0.76 (0.80-0.71)	0.74 (0.79-0.68)		
code 2	0.77 (0.79-0.76)	0.77 (0.78-0.77)		
code 3	0.67 (0.72-0.60)	0.68 (0.74-0.61)		
code 4	0.64 (0.70-0.58)	0.64 (0.70-0.56)		
code 5	0.63 (0.69-0.56)	0.64 (0.71-0.57)		
code 6	0.62 (0.68-0.55)	0.61 (0.68-0.53)		
code 7	0.61 (0.68-0.55)	0.61 (0.68-0.53)		
code 8	0.61 (0.67-0.56)	0.61 (0.67-0.53)		
code 9	0.61 (0.67-0.53)	0.60 (0.67-0.52)		
code 10	0.60 (0.66-0.52)	0.60 (0.67-0.52)		
code 11	0.59 (0.66-0.52)	0.57 (0.64-0.49)		
code 12	0.57 (0.63-0.49)	0.57 (0.65-0.49)		
code 13	0.56 (0.63-0.48)	0.56 (0.67-0.52)		
code 14	0.54 (0.62-0.46)	0.54 (0.66-0.48)		
code 15	0.53 (0.63-0.48)	0.56 (0.67-0.47)		
code 16	0.53 (0.60-0.45)	0.53 (0.61-0.44)		
code 17	0.53 (0.64-0.42)	0.53 (0.66-0.42)		

J. Chem. Inf. Model. 2011, 51, 2115

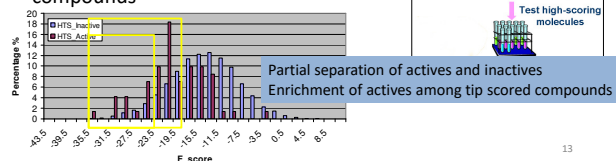
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Virtual screening

- Identification of chemical starting point
- Computation:
 - Docking a large number of structurally diverse compounds
 - Ranking the complexes (compounds) by score



- Experimental testing of top scored compounds



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Identification of chemical starting point and virtual screening

- High throughput screening (HTS) - experimental
 - Finding compounds with the required effect on a target protein
 - Biochemical/biophysical methods
 - receptor binding
 - Enzyme inhibition
 - ...
 - Testing 10^5 - 10^6 compounds
 - Number of hits: $\sim 10^2$
 - Hit rate: 0.1% ($10^2/10^5$)
- Virtual screening
 - Objective: increase HTS hit rate by computational (cheap) prescreening
 - Docking and scoring $\sim 10^6$ compounds
 - Experimental testing of top $\sim 10^3$ compounds; typical hit rate: 1-10 %

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Slide 14

GF1 György Ferenczy, 05/12/2019

Summary

- Endpoint methods – Approximate schemes for estimating ΔG
 - MM-PBSA – not discussed
 - fast (lesser extent)
 - Fair correlation with experimental values
 - Well fitted to improved virtual screening results
 - Docking - scoring
 - Very fast
 - Good quality binding mode prediction
 - Weak correlation between score and experimental affinity
 - Virtual screening is an established tool in chemical starting point identification
 - Intensively applied in pharmaceutical research

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