

Local change of membrane-potential

Laws of sensation

for pharmacy students

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Lecture topics

Topics

- Resting membrane potential (recap)
- Change of RMP
 - Graded membrane potential
 - Equivalent circuit model of cell membrane
 - Propagation of potential change
 - Action potential
 - Phases, ion currents
 - Propagation
 - (Voltage clamp, patch clamp)
- Process of sensation
 - Receptors
 - Sensory nerve
 - Psychophysical laws

Related practices

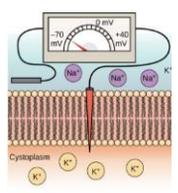
- Diffusion
- ECC
- Sensor
- Audiometry

Textbook chapters:

- III/4.3. Changes of membrane potential due to Stimuli below the depolarization threshold
- III/4.4. Membrane potential changes in the excited state: The action potential
- IV/1. General laws of the perception



Membrane potential

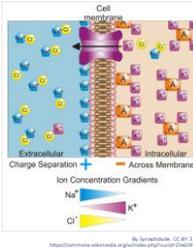


Transmembrane potential / Membrane voltage / „Resting membrane potential“

- Electric potential difference between inner and outer surface of the membrane
- Present in all living cell
- Varies among cell types (-30 mV to -90 mV)
- Negative sign: cell interior is negative compared to extracellular space
- Functions:
 - providing power to operate a variety of “molecular devices” embedded in the membrane (cell as battery)
 - in electrically excitable cells such as neurons and muscle cells, it is used for transmitting signals between different parts of a cell



Membrane potential



Two sides of the membrane has different ionic composition

Cell type	Intracellular concentration [mM]			Extracellular concentration [mM]		
	Na ⁺	K ⁺	Cl ⁻	Na ⁺	K ⁺	Cl ⁻
Squid giant axon	72	345	61	455	10	540
Frog muscle	20	139	3,8	120	2,5	120
Rat muscle	12	180	3,8	150	4,5	110

- Large phosphate and protein anions inside – p - 0
- p is different for the different ions
- Electric and chemical potential difference occurs between the two sides.



Generation of membrane potential I.

Model I

Presumptions:

- Closed thermodynamic system
- Membrane permeable to ions
- Cytoplasm and extracellular space are in thermodynamic equilibrium – for each ion!
- No net transport of ions
- Thermodynamic force is 0
- Electrochemical potential is the same at the two sides for each type of ion:

$$\mu_{ext}^{int} - \mu_{ext}^{ext} = 0$$

$$\mu_0 + RT \ln c_{int}^{int} + zF\phi_{int}^{ext} = \mu_0 + RT \ln c_{int}^{int} + zF\phi_{int}^{int}$$

$$\phi_{int}^{int} - \phi_{int}^{ext} = U_b = \frac{RT}{zF} \ln \frac{c_{int}^{ext}}{c_{int}^{int}} \quad \text{Nerst equation}$$

Electric potential of *i*th ion in equilibrium = equilibrium potential = Electromotive force of a concentration cell of the *i*th ion

	Squid giant axon	Frog muscle
U _{measured}	-62 mV	-92 mV
U _{Nernst}	47 mV	46 mV
U _{Na+}	-91 mV	-103 mV
U _{Cl-}	-56 mV	-88 mV

Results: model failed

- Nerst equation is inadequate to interpret resting potential
- It is not a closed system in equilibrium
- Transport of individual ions is not independent



Generation of membrane potential III.

Electrodifusion model

- Passive-ion diffusion maintains an electric potential difference
- Permeability is different for different ions
- Flux of individual ions ≠ 0
- At rest the transmembrane potential difference is constant → total electric charge and particle flux must be 0.
- Thus, flux of ions may depend on each other

Steady state electrodiffusion:

- Constant electrochemical potential gradient → constant flow of ions → electrically compensate each other → no net charge accumulation
- Na⁺/K⁺ pumps maintains constant ion gradients
- Result: good agreement with experimental data

flux density of *k*th particle

$$J_k = -u_k k \frac{\Delta c_k}{\Delta x} + c_k \frac{z_k F \Delta \phi}{RT \Delta x}$$

electric potential difference

electrochemical potential gradient

$$\sum J_k = 0$$

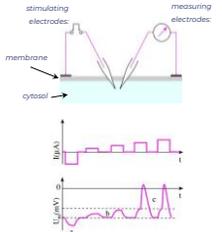
Goldman-Hodgkin-Katz equation:

$$U = \Delta \phi = \frac{RT}{F} \ln \frac{p_K [K^+]_{ext} + p_{Na^+} [Na^+]_{ext} + p_{Cl^-} [Cl^-]_{int}}{p_K [K^+]_{int} + p_{Na^+} [Na^+]_{int} + p_{Cl^-} [Cl^-]_{ext}}$$

	Squid giant axon	Frog muscle
U _{measured}	-62 mV	-92 mV
U _{calc.}	-61,3 mV	-89,2 mV



Electric perturbation of RMP



Experiment

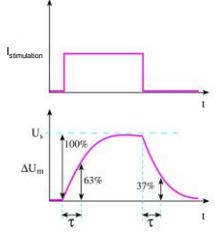
- Current is driven into the cell
- Square-wave pulses
- Direction and magnitude can be controlled

Results:

- **Graded membrane potential**
 - Hyperpolarization / depolarization
 - Size and direction can be controlled
 - Analogue
 - Localized
 - Up to a threshold: RMP changes proportionally
- **Action potential:** reaching a stimulation threshold
 - large, uniform pulse (action potential)

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Electric perturbation of RMP II.



Depolarizing current below the threshold

- Square-wave current pulse
- Lower than the depolarization threshold

Result:

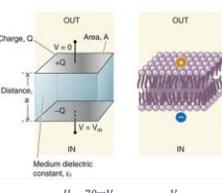
- Delayed, saturating depolarization
- Decay after stimulation ceases
- Shape resembling charging and discharging an RC circuit

Conclusion:

- Cell membrane could be modelled by a parallel RC circuit.

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Electric circuit model of the membrane



Membrane as an RC circuit I.

Lipid bilayer:

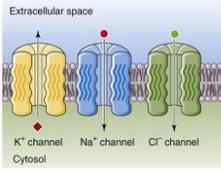
- Good insulator
- Charges can accumulate on both sides
- Behaves as a capacitor
- Specific capacity
 - constant in time
 - by numbers (for a mammal neuron):
 - $C_m = 10 \text{ nF/mm}^2$
 - $4 \cdot 10^{10} \text{ charge/cm}^2$
 - $A_{\text{neuron}} = 0.01\text{-}0.1 \text{ mm}^2$
 - $C_{\text{neuron}} = 0.1\text{-}1 \text{ nF}$

$$C_m = \frac{C}{A} = \frac{Q}{UA}$$

$$E = \frac{U}{d} = \frac{70 \text{ mV}}{5 \text{ nm}} = 1.4 \cdot 10^7 \frac{\text{V}}{\text{m}}$$

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Electric circuit model of the membrane



Membrane as an RC circuit II.

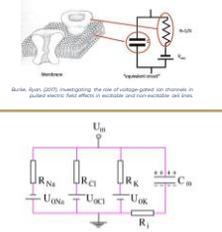
ion channels:

- Ion-selective permeability → ion currents
- Behave as resistors
- Ion-specific conductivities
 - Constant in resting state
 - Changes upon excitation
- $C_m / C_{\text{total}} = 1 / 0.04 / 0.45$ (in brain neurons)

$$R_i = \frac{1}{G_i} \quad \frac{1}{R_{\text{total}}} = \sum_{i=1}^n \frac{1}{R_i}$$

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Electric circuit model of the membrane



Equivalent circuit model

- Parallel RC circuit
- For each ion type:
 - permeability → resistance ($R=1/C$)
 - equilibrium potential → electromotive force (U_0)

Nernst equation: $U_{0i} = \frac{RT}{z_i F} \ln \frac{c_i^{\text{out}}}{c_i^{\text{in}}}$

Conductive current of the i th ion: $I_j = \frac{U_m - U_{0j}}{R_j}$

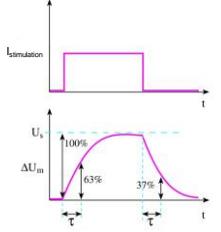
capacitive current: $I_C = C_m \frac{\Delta U_m}{\Delta t}$

GHK: total charge flux = 0

$$\Sigma I = I_C + \Sigma I_j - I_{\text{stimulation}} = 0$$

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Electric perturbation of RMP III.



Changes of membrane potential upon stimulation

- (1-)exponential time course:

$$U_m(t) = U_0 \left[1 - e^{-\frac{t}{\tau}} \right]$$

U_0 : saturation MP
 R_m : membrane resistance
 C_m : membrane capacitance
 τ : relaxed time

- Amplitude of change (ΔU_m) is proportional to stimulus amplitude
- **Time constant of the membrane:** time required to reach 63% of saturation value.

$$\tau = R_m C_m$$

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Electric perturbation of RMP IV.

Changes of membrane potential following stimulation

- Exponential decay course:

$$U_m(t) = U_s \cdot e^{-\frac{t}{R_m C_m}}$$
- U_s : saturation MP
- R_m : membrane resistance
- C_m : membrane capacitance
- t : elapsed time

Time constant of the membrane: time required to reach 37% (U_0/e) of saturation value.

$$\tau = R_m C_m$$

- Actual membrane potential during stimulation depends on:
 - time constant
 - size and direction of stimulation.

Propagation of a potential change

$$U_m(x) - U_m(x_0) = e^{-\frac{x}{\lambda}}$$

space constant of the membrane

$$\lambda \propto \sqrt{\frac{R_m}{R_i}}$$

Space constant of membrane: distance in which the maximal value of the induced potential change decreases to its e-th level (37%)

Change of MP: decreases exponentially with distance

Propagation of a potential change

Model of a larger membrane section:

Action potential

Action potential:

- Sudden, fast, transitory and propagating change of the RMP
- Only in excitable cells: neurons, muscle cells
- Trigger: RMP exceeding the depolarization threshold
- Digital: „all or nothing“ response
- Characteristic time-dependent shape
- Does not depend on the size or duration of trigger pulse
- few ms duration; cca. 100 mV amplitude
- Propagates along the cell/fibre unidirectionally

Action potential

Phases:

- #1 **hypopolarization** (slow, up to threshold)
- #2 **depolarization phase** (rapid)
- #3 **peak potential**
- #4 **repolarization** (rapid)
- #5 **hyperpolarizing afterpotential**

- Shape, time course, peak potential: cell-type dependent
- Depolarizing threshold: changes during AP
 - Absolute refractive period**: large threshold, no excitation; around peak potential
 - Relative refractive period**: higher excitation threshold, mainly during hyperpolarization

Action potential - examples

The first intracellular recording of an action potential, from squid axon. Time calibration, 2 ms. (origins: A. L. H. Huxley, A. F. Hodgkin, 1944, 1952, 1953)

Action potential – ionic currents

Sodium channels

- **Deactivated state**: at RMP
- **Activated state**: reaching depolarizing threshold → voltage gated Na⁺-channels open → Na⁺ influx → depolarization → more Na⁺-channels open → further depolarization (+ feedback, Hodgkin cycle)
- **Inactivated state**: reached shortly after opening → channels closed → absolute refractory period

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Action potential – ionic currents

Potassium channels

- **Deactivated state**: at RMP
- **Activated state**: reaching depolarizing threshold → voltage gated K⁺-channels open (delayed kinetics) → K⁺ outflux → repolarization
- **Inactivated state**: channels close slower → relatively large K⁺ permeability → transient hyperpolarization → relative refractory period

Na⁺ / K⁺ ATPase

- Restores cytosolic concentration of the cations
- 3 Na⁺ out / 2 K⁺ in

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Action potential – modelled

Modified equivalent circuit model (Hodgkin)

- Permeabilities of ions are not constant → nor the resistances are
- Conductivity depends on membrane potential
- Conductivities can be measured by voltage-clamp technique
- G(U) functions were determined
- Substituted into the modell AP function could be calculated
- Showed good agreement with experimental data.

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Action potential – propagation

Propagation of AP along non-myelinated axon(al region)

- Exponential decay of peak potential in space – determined by **space constant**
- Large enough depolarizations in nearby regions to exceed threshold → AP is generated.
- Na⁺ channels become inactive in previously activated areas → absolute refractor period.
- By the time the refractory periods cease depolarisation wave travelled far enough to avoid re-excitation.
- Unidirectional propagation
- v=1-30 m/s (mammal neurons)

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Action potential – propagation II.

Space constant: $\lambda \propto \sqrt{\frac{R_m}{R_i}}$

How is it possible to increase the speed of propagation?

- Increase R_m: **myelination** (decreases C_m and thus the time constant as well)
- Decrease R_i: enlargement of cellular (axonal) diameter – non advantageous

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Action potential – propagation III.

Measurement of the propagation of a nervous impulse. (Adapted from A. F. Huxley and R. Stämpfli, J. Physiol., 1952/53, 194a.)

Optical measurement of action potential speed. (A) Series of images of the fluorescence in the cell as a function of time. (B) A series of equally spaced (15 μm) measurement points along three different processes are used to measure the arrival time of a propagating action potential. (C) Arrival times for the three processes shown in part (B).

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Voltage clamp

Experimental setup

Working principle

- MP kept constant regardless transmembrane ionic currents
- Intracellular electrode #1: records actual MP
- Intracellular electrode #2:
 - conducts current to set MP to command V value → **MP is clamped**
 - Current amplitude is equal to membrane current (I)
- Channels can be blocked (Na⁺: tetrodotoxin; K⁺: tetraethylammonium) → specific currents can be calculated
- MP-dependent conductivity of ions can be determined

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Patch clamp

Experimental setup

Working principle

- Class micropipette ($d_{tip} \approx 0.5-1 \mu m$) attached to a cell
- Large electrical and mechanical resistance contact („giga-seal“)
- Cell Attached Patch (CAP)
- Membrane patch with 1-few ion channels
- Measuring electrode in the electrolyte inside pipette
- Voltage-clamp setup → currents (pA-s) are measured
- Single opening – unitary current jump
- 10^3-10^7 ions / few ms; conductance: ~pS
- **Applications:**
 - Ion-channel studies (drug effects; receptor-ligand binding; kinetics, activation, inhibition, etc.)

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Patch clamp

Recording methods

Single channel currents (human blood lymphocyte, CAP)

Whole-cell K⁺ currents during depolarizations of a T-lymphocyte

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Process of sensation

Modality: types of sensation, eg: vision, hearing, olfaction, taste, touch, pain, thirst, muscle fatigue.
Quality: psychophysical sensation given by modality, eg: loudness, colour shade
Quantity: degrees of a quality, eg: son-loudness; brightness
Perception: interpretation of the sensation, eg: experience of vision, voice recognition

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Receptors

Receptor cell

- Transducer: converts physical/chemical stimulus energy into receptor potential
- **Receptor potential** (generator potential): stimulus-triggered, local change of MP
- **Amplitude coding:** amplitude depends on stimulus intensity
- Specifically energy sensitive
- Stimulus energy – wide range
- **Dynamic compression/expansion:** weak stimuli results in relatively larger/smaller change of MP than strong stimuli
- **Adaptation:** constant stimulus intensity – decreasing receptor potential

Graded responses of a muscle spindle receptor to stretch. Graded stretches are indicated by the stretch monitor in the lower traces; graded generator potentials are shown in the upper traces. (Oatman D and Shepherd GM. Cold Spring Harbor Symp Quant Biol 36:105-114, 1971.)

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Receptors

Classification	Receptor cell/group	What does it sense?	Example
According to stimulus	Photoreceptor	Light (D.A)	Retina
	Chemoreceptor	Chemical substances (concentration)	Taste buds (tongue); olfactory receptors (nose)
	Thermoreceptor	ΔT	Skin thermoreceptors
	Mechanoreceptor	p (touch)	Skin, hair
	Baroreceptor	p	Blood vessel wall
According to location	Exteroreceptor	External conditions	Eye, ear, nose, tongue, skin
	Interoreceptor	Internal conditions	Blood vessel wall; baroreceptors, pain sensors
	Proprioceptor	Position of body parts	Muscle, joint
According to complexity	General sensory organs	Individual or grouped receptor cells	Hat, touch
	Specialized sensory organs	Complex sensory organs (large number of receptor cells)	Eye, ear, nose, tongue

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Sensory nerve

Nerve cell

- Attached to receptor cell.
- Answers with AP when receptor potential exceeds depolarisation threshold.
- AP propagates fast alongside nerve fiber to reach the sensory cortex.
- Frequency coding: frequency of AP proportional to the size of receptor potential.

The diagram illustrates the process of signal transmission in a sensory nerve cell. It consists of three vertically stacked graphs sharing a common time axis (t):

- Stimulus:** Shows a series of rectangular pulses of varying heights. The top horizontal line is labeled $\Phi_{\text{threshold}}$.
- Receptor potential:** Shows a series of sub-threshold depolarizations that increase in amplitude with the stimulus. The top horizontal line is labeled U_{gen} and the bottom horizontal line is labeled $U_{\text{threshold}}$.
- Action potential:** Shows a series of all-or-none spikes that occur whenever the receptor potential reaches the threshold. The top horizontal line is labeled U_{active} and the bottom horizontal line is labeled 0 mV and -70 mV .

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Psychophysical laws – I.

Weber-Fechner law

$$\psi = a \cdot \log_b \frac{\Phi}{\Phi_0}$$

ψ : strenght of sensation
 a and b : constants
 Φ : absolute stimulus intensity
 Φ_0 : absolute threshold stimulus

- Sensation is proportional to the logarithm of relative stimulus intensity (Φ/Φ_0)
- Very limited validity

Stevens law

$$\psi = a \cdot \left[\frac{\Phi}{\Phi_0} \right]^n$$

a and n : constants

- Sensation is proportional to the power of relative stimulus intensity (Φ/Φ_0)
- Valid over wide ranges of stimulus intensities
- $n < 1$ → compressive function (pl. loudness: $n=0,3$)
- $n > 1$ → expansive function (pl. electric shock: $n=3,5$)

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Psychophysical laws – II.

This graph plots 'Magnitude estimate' on the y-axis (0 to 80) against 'Stimulus intensity' on the x-axis (0 to 100). Three curves are shown: a red curve for 'Electric shock' (n > 1, expansive), a green curve for 'Apparent line length' (n = 1, linear), and a blue curve for 'Brightness' (n < 1, compressive).

This log-log plot shows 'Log magnitude estimate' on the y-axis (0 to 2.0) against 'Log stimulus intensity' on the x-axis (0 to 2.0). Three lines are shown: a red line for 'Electric shock' (n > 1, steeper slope), a green line for 'Apparent line length' (n = 1, 45-degree slope), and a blue line for 'Brightness' (n < 1, shallower slope).

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Thanks for your attention!

Dr. Tamás Bozó