Evaluation of CEEPUS Intensive Course 2024 BIO- AND SINGLE MOLECULAR ANALYSIS FROM BASICS TO PRACTICE

Jolan Harsfalvi



Semmelweis University
Faculty of Medicine
Institute of Biophysics and Radiation
Biology
Tuzolto str 37-47, 1094, Budapest,
Hungary

harsfalvi.jolan@semmelweis.hu

2023/24 New grant for Intensive Course

- personal visit and proposal at the Hungarian NCO, April 2023 (Donath Nagy Gabriella, Jolan Harsfalvi)
- call December 2023
 https://tka.hu/palyazatok/607/palyazati-dokumentumok
- Inquiry letters to coordinators, December 2023 May 2024
- completed Grant application sent for check to NCO at 24th December 2023
- Submission of the corrected Grant application, 10th February 2024

Structure of the form – questions to answer

National Application form international requirement (I translated to English. NCO)

Administrative data (University and organizing Unit)

A. Body of the Course:

- planned # of participants (students and teachers national and international);
- 2. from which countries the guests are expected;
- 3. which partners will be involved;
- 4. how and where the course will be advertised;
- 5. how the participants will be selected;

ATTACH the CALL for THE COURSE in ENGLISH.

Structure of the form – questions to answer, Cont.

B. Content of the Course

- 1. Aims, 1000-2000 characters (Hungarian)
- 2. international dimensions (list of partners involved)
- how the tasks will be shared among the involved partners;
- Why the number of students and teachers is necessary for the realization of the course, and what is the ratio between students and teachers;
- 5. Why the number of the days are necessary for the course

Structure of the form – questions to answer, Cont.

- 6. how the interactivity of the course is ensured;
- 7. what kind of innovative pedagogical methods will be utilized;
- 8. how the not CEEPUS teachers will be donated;
 - 9. how the achievement of the participants will be measured;
- 10. how many credit points can be given;
- 11. how the course will be evaluated; how the result of the course will be distributed.

ATTACH the DETAILED PROGRAM of THE COURSE in English

Structure of the form – questions to answer, Cont.

- C. Financial plan
- D. Attachments
 - a) CALL for THE COURSE in English
 - b) DETAILED PROGRAM of THE COURSE in English
- E. Earlier courses in Hungary during the existence of the Network:
 - a) when
 - b) where (2020 Semmelweis, 2022 Pécs)
 - c) how many was the donation

04.23.2024 awarded **CEEPUS Intensive Course**

Administration 22-31.05.2024

- Letter to the Rector
- 2. Supporting documentation
- Informal institutional data form, statements
- 4. Instruction
- Anonym review1
- 6. Anonym review2

My information and call to our Network's coordinators, Teachers, BSc, MSc, PhD students examination period

Realization CEEPUS Intensive Course

22-31.05.2024

Aim is to learn bioanalysis in an overall aspect from separation/isolation, and identification to visualization of molecules; from basic knowledge to innovation; with a focus on combining classical and new single molecular techniques.

Lectures, hands-on training and "depth over breadth" way of teaching.

Encourage PhD students to continue their studies at the partner Doctoral Schools of the partner Universities and obtain joint PhD degree.

BIO- AND SINGLE MOLECULAR ANALYSIS FROM BASICS TO PRACTICE

Semmelweis University Basic Medical Science Center Room 1.205, Intensive Course 2024 BIFI 22-31 May 2024 CEEPUS, In the frame of RO-0010-18-2324 network, Teaching, and Learning & Joanalysis CEEPUS Network (coordinator Jolan Harsfalvi)

	Learning Bioanalysis CEEPUS Network (Coordinator Joian Harsfalvi)							
2024	*HOUR scheduled for the beginning of the daily teaching programs.							
	Days were ended with quizzes of the lecturers and demonstrators - the student gave the most good answer awarded a Semmelweis flask- and closed at 5 pm.							
May	9 - 10	1011		11 - 12	12 - 13 - 14	14 - 15	15-16	
22				ARRIVAL to Budapest				
23	Welcome Gabriella Donath Periodic phenomena and pattern		Protein structure determination in solution with Nanomanipulation of single viruses (Bálint					
	Nagy, Jolán Hársfalvi,	formation in chemical and biological		X-ray and neutron small-angle scattering. Hands- Kiss)				
	Introduction of the Students	systems (<i>Gabriella Donáth-Nagy</i>)		on experience in interpreting structural biology				
			data (Bence Fehér)					
24	From single molecules to the livi	ng organism.	S e mm elwe is	Resonance theory and	practice in small Békésy G	yösgy Reserach Center Ato	mic force microscopy	
	Passion for discovery and value-driven leadership (Miklós Buda		Budapest	groups at 7 working places (<i>Ádám</i> (AFM) (<i>Ádám Zolcsák</i>)				
	Kellermayer)		(Bálint	Zolcsák)	Practice: preparation of a multimer protein and its AFM			
			Budavári)			sis (Jolán Hársfalvi)		
25	Explore links between bioanalysis and biophysics, force sensing during pri			imary hemostasis.		ersal method for synthesis of artificial gel antibodies by the imprinting		
	Practice: preparation of a multimer protein and its AFM analysis				pproach combined with a unique electrophoresis technique for detection			
	(Jolán Hársfalvi), Atomic force microscopy (AFM) (Ádám Zoksák)				of minute structural differences of proteins, viruses, and bacteria (Anikó			
					Takátsy)			
26	Visiting museums: Natural History Museum; Semmelweis History of Medicine; Hospital in The Rock Nuclear Bunker. (Show contexts of the Course and science in Hungary.							
27	All atom simulation of In silico biophysics of transmembrane		Analytical and	Demonstration of	Békésy György Reserach Center,			
	biological macromolecules (Erika Balog) proteins (Tamás Hege düs		5)	biomedical	electrochemical sensing	Demonstrations and measurement options (Optical tweezers, Optical microscopy, Optical spectroscopy (<i>Balázs Kretzer</i>)		
				applications of	using a portable			
				molecularly	electrochemical system			
				imprinted polymers	(Bogdan Cezar Iacob)			
				(Ede Bodoki)				
28	Phosphorylation-dependent	Janochemistry and its medical applications (Angéla Jedlovszky-				**How to develop an	**Live	
BIFI	structure of titin (Zsolt	Laboratory visit & work & Ákos Juhász)	Bálint Budavári, Kristóf Molnár, Sarolta Halmóczki, Veronika Pálos,			HPLC separation method demonstration of		
library	Mártonfalvi)				for pharmaceutical how to compare			
						quality control (Martin	an alysis results of	
						Schmid)	real Viagra samples	
							with fakes (IMartin	
							Schmid)	
29	**Recent advances in chiral analysis (Gergő Tóth)				**Practical method	Showing quantitative in vivo molecular Imagin		
	HPLC measurement by students in 4 groups in the laboratory (Gergő Tóth					center with explanation of		
					electrophoresis (Tomas	presentation of an anima	l experiment (<i>Noémi</i>	
					Krizek)	Kovács),		
30		Békésy Gyösgy Reserach Center Fluorescent- and Optical microscopy, Spectroscopy (<i>Jokin Hársfalvi</i>)						
31				LEAVE for home				

^{*}To meet the students' requirements, the duration of the lectures and demonstrations are not limited strictly, but a short lunchtime before 2 pm is obligatory.

^{**} ALL IN <u>Department</u> of Pharmaceutical Chemistry, Faculty of Pharmacy, Semmelweis University, 1092 Budapest, Hőgyes Endre u. 9

Certificate of attendance

Name's of the Student

for participation at the Intensive Course on Bio- and Single Molecule Analysis from Basics to Practice, organised in Budapest, between 22th and 31th of May 2024, by Institute of Biophysics and Radiation Biology, Semmelweis University in the frame of CEEPUS RO-0010-18-2324.



Kellermayer Miklós Head of the institute



Thanks for all the <u>contributors!</u>

Links to the program and lectures

Questions of teachers & students:

Same Time, Next year? Same Place? Same



